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(54) **PESTICIDAL GENES FROM  
BREVIBACILLUS AND METHODS FOR  
THEIR USE**

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(58) **Field of Classification Search**

None

See application file for complete search history.

(56) **References Cited**

**U.S. PATENT DOCUMENTS**

7,091,399 B2 \* 8/2006 Boets et al. .... 800/279

**FOREIGN PATENT DOCUMENTS**

EP 1277763 1/2003  
WO 0009697 2/2000  
WO 01/87931 11/2001

**OTHER PUBLICATIONS**

de Maagd, et al (Trends in Genetics, 2001, 17(4), pp. 193-199).

De Oliveira E.J., et al. Molecular characterization of *Brevibacillus*  
*laterosporus* and its potential use in biological control, *Applied and*  
*Environmental Microbiology* US, Nov. 2004, vol. 70, No. 11, pp.  
6657-6664.

Favret, Montgomery E., et al., Insecticidal Activity of *Bacillus*  
*laterosporus*, *Journal of Invertebrate Pathology*, 1985, vol. 45, pp.  
195-203.

Orlova Margarita V., et al., Insecticidal activity of *Bacillus*  
*laterosporus*, *Applied and Environmental Microbiology*, Jul. 1998,  
vol. 64, No. 7, pp. 2723-2725.

Prasanna, et al. (Appl. Microbiol Biotechnol, Apr. 29, 2012).

Ruii Luca, et al., Lethal and Sublethal Effects of *Brevibacillus*  
*Laterosporus* on the Housefly (*Musca domestica*), *Entomologia*  
*Experimentalis Et Applicata*, Kluwer Academic Publishers,  
Dordrecht, NL, Feb. 2006, vol. 118, No. 2, pp. 137-144.

Ruii, et al., Toxicity of a *Brevibacillus Laterosporus* Strain Lacking  
Parasporal Crystals Against *Musca domestica* and *Aedes Aegypti*,  
*Biological Control*, San Diego, CA, US, Sep. 2007, vol. 43, pp.  
136-143.

Singer, Samuel, The Utility of Strains of Morphological Group II  
*Bacillus*, *Advances in Applied Microbiology*, vol. 42, pp. 219-261.

Tounsi, et al. (*J. Appl Microbiol* 95:23-28; 2003).

Zahner, Viviane, et al., Geotypic Diversity among *Brevibacillus*  
*laterosporus* Strains, *Applied and Environmental Microbiology*, Nov.  
1999, vol. 65, No. 11, pp. 5182-5185.

Partial European Search Report for Application No. 13182993.9-  
1401 dated Jan. 17, 2014.

Partial International Search Report for PCT/US2009/069144, mailed  
Mar. 30, 2010 (3 pages).

\* cited by examiner

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(57) **ABSTRACT**

Compositions and methods for conferring insecticidal activ-  
ity to bacteria, plants, plant cells, tissues and seeds are pro-  
vided. Compositions including a coding sequence for a *Brevi-*  
*bacillus*-derived delta-endotoxin polypeptide are provided.  
The coding sequences can be used in DNA constructs or  
expression cassettes for transformation and expression in  
plants and bacteria. Compositions also include transformed  
bacteria, plants, plant cells, tissues, and seeds. In particular,  
isolated delta-endotoxin nucleic acid molecules are provided.  
Additionally, amino acid sequences corresponding to the  
polynucleotides are encompassed, and antibodies specifically  
binding to those amino acid sequences. In particular, the  
present invention provides for isolated nucleic acid molecules  
having nucleotide sequences encoding the amino acid  
sequence shown in SEQ ID NO:2, 4, 7, or 10, or the nucle-  
otide sequence set forth in SEQ ID NO:1, 3, 5, 6, 8, 9, 11, 12,  
13, 14, or 15, as well as variants and fragments thereof.

**21 Claims, No Drawings**

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# PESTICIDAL GENES FROM BREVIBACILLUS AND METHODS FOR THEIR USE

## CROSS REFERENCE TO RELATED APPLICATION

This application is a continuation of U.S. patent application Ser. No. 12/644,632, filed Dec. 22, 2009, which claims the benefit of U.S. Provisional Application Ser. No. 61/139,947, filed Dec. 22, 2008, the contents of which are herein incorporated by reference in their entirety.

## REFERENCE TO SEQUENCE LISTING SUBMITTED AS A TEXT FILE VIA EFS-WEB

The official copy of the sequence listing is submitted concurrently with the specification as an ASCII formatted text file via EFS-Web, with a file name of "APA059SEQLIST.txt", a creation date of Feb. 27, 2013, and a size of 98 kilobytes. The sequence listing filed via EFS-Web is part of the specification and is hereby incorporated in its entirety by reference herein.

## FIELD OF THE INVENTION

This invention relates to the field of molecular biology. Provided are novel genes that encode insecticidal proteins. These proteins and the nucleic acid sequences that encode them are useful in preparing insecticidal formulations and in the production of transgenic insect-resistant plants.

## BACKGROUND OF THE INVENTION

*Brevibacillus* is a spore-forming bacterium that has been suggested for probiotic effects. For example, *Brevibacillus brevis* is now well established as a biocontrol agent in many areas, and has been shown to have efficacy against *Botrytis* and powdery mildew disease (Edwards and Seddon, Edwards and Seddon (1992) Recent Advances in *Botrytis* Research. The Netherlands: Pudoc Scientific Publications; *Bacillus brevis* as a Biocontrol Agent against *Botrytis cinerea* on Protected Chinese Cabbage; pp. 267-271) and *Fusarium* head blight (FHB) (Zhang et al. (2005) J Zhejiang Univ Sci B. 6(8):770-777). By comparing the activity of *B. brevis* Nagano against *Botrytis cinerea* with that of pure gramicidin S and the antibiotic-negative mutant *B. brevis* E-1, Edwards and Seddon ((2001) J Appl Microbiol. 91:652-659) showed that the mode of antagonism exhibited was antibiosis due to the presence of gramicidin S. There are some other antibiotics (for example tyrocidins and gramicidin D) reported to be produced by *B. brevis* (Saito et al., 1970, *Adv Enzymol.* 33:337-380).

*Brevibacillus laterosporus* comb. nov. (Shida (1996) Int. J. Syst. Bacteriol. 46:939-946), previously classified as *Bacillus laterosporus*, is an aerobic spore-forming bacterium that can also demonstrate pathogenicity to insects. In common with *B. sphaericus* and *B. thuringiensis*, *B. laterosporus* produces parasporal bodies, which in this species may be canoe-shaped and which serve to cradle the spore (Hanney (1957) J. Biophys. Biochem. Cytol. 3:1001-1010) or can even be present in different shapes (Smirnova et al. (1996) Res. Microbiol. 147:343-350). However, these parasporal bodies were not considered to have any entomocidal activity (Favret and Yousten (1985) J. Invertebr. Pathol. 45:195-203) until Orlova et al. ((1998) Appl. Environ. Microbiol. 64:2723-

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2725) demonstrated that some crystals produced during sporulation are highly toxic to *Aedes aegypti* and *Anopheles stephensi* larvae.

Some *B. laterosporus* strains show no apparent toxic activity to any test organism, and the observed toxicity is not homogeneous among toxic isolates (Favret and Yousten (1985) J. Invertebr. Pathol. 45:195-203, Rivers et al. (1991) J. Invertebr. Pathol. 58:444-447, and Singer (1996) Adv. Appl. Microbiol. 42:219-261). The results of the first bioassays with *B. laterosporus* demonstrated that some strains presented a larvicidal activity which was 1,000 times lower than that of the *B. thuringiensis* var. *israelensis* standard (Favret and Yousten (1985), Rivers et al. (1991)). These results discouraged the use of *B. laterosporus* in biological control. No entomocidal activity has been demonstrated against plant pathogens using isolates of *Brevibacillus* sp.

## SUMMARY OF INVENTION

Compositions and methods for conferring insect resistance to bacteria, plants, plant cells, tissues and seeds are provided. Compositions include nucleic acid molecules encoding sequences for *Brevibacillus*-derived delta-endotoxin polypeptides, vectors comprising those nucleic acid molecules, and host cells comprising the vectors. Compositions also include the polypeptide sequences of the endotoxin, and antibodies to those polypeptides. The nucleotide sequences can be used in DNA constructs or expression cassettes for transformation and expression in organisms, including microorganisms and plants. The nucleotide or amino acid sequences may be synthetic sequences that have been designed for expression in an organism including, but not limited to, a microorganism or a plant. Compositions also comprise transformed bacteria, plants, plant cells, tissues, and seeds.

In particular, isolated nucleic acid molecules corresponding to delta-endotoxin nucleic acid sequences are provided. Additionally, amino acid sequences corresponding to the polynucleotides are encompassed. In particular, the present invention provides for an isolated nucleic acid molecule comprising a nucleotide sequence encoding the amino acid sequence shown in any of SEQ ID NO:2, 4, 7, or 10, or a nucleotide sequence set forth in any of SEQ ID NO:1, 3, 5, 6, 8, or 9, as well as variants and fragments thereof. Nucleotide sequences that are complementary to a nucleotide sequence of the invention, or that hybridize to a sequence of the invention are also encompassed.

The compositions and methods of the invention are useful for the production of organisms with resistance to plant pests, specifically bacteria and plants with resistance to these pests. These organisms and compositions derived from them are desirable for agricultural purposes. The compositions of the invention are also useful for generating altered or improved delta-endotoxin proteins that have pesticidal activity, or for detecting the presence of delta-endotoxin proteins or nucleic acids in products or organisms.

## DETAILED DESCRIPTION

The present invention is drawn to compositions and methods for regulating insect resistance in organisms, particularly plants or plant cells. The methods involve transforming organisms with a nucleotide sequence encoding a delta-endotoxin protein of the invention. In particular, the nucleotide sequences of the invention are useful for preparing plants and microorganisms that possess insecticidal activity. Thus, transformed bacteria, plants, plant cells, plant tissues and

seeds are provided. Compositions are delta-endotoxin nucleic acids and proteins derived from a *Brevibacillus* organism. By "derived from" is intended that the nucleic acid or polypeptide is cloned or otherwise isolated from a *Brevibacillus* organism, or is a sequence that has been cloned or otherwise isolated from the *Brevibacillus* organism and subsequently altered (e.g., by making nucleotide and/or amino acid changes). The sequences find use in the construction of expression vectors for subsequent transformation into organisms of interest, as probes for the isolation of other delta-endotoxin genes, and for the generation of altered insecticidal proteins by methods known in the art, such as domain swapping or DNA shuffling. The proteins find use in controlling or killing lepidopteran, coleopteran, nematode, and other insect populations that are pathogenic to plants, and for producing compositions with insecticidal activity.

Exemplary *Brevibacillus* organisms from which the delta-endotoxin sequences encompassed by the present invention can be derived include *Brevibacillus* agri, *Brevibacillus borstelensis*, *Brevibacillus brevis*, *Brevibacillus centrosporus*, *Brevibacillus choshinensis*, *Brevibacillus formosus*, *Brevibacillus ginsengisoli*, *Brevibacillus invocatus*, *Brevibacillus laterosporus*, *Bacillus laterosporus*, *Brevibacillus levickii*, *Brevibacillus limnophilus*, *Brevibacillus parabrevis*, *Brevibacillus reuszeri*, *Brevibacillus* sp., and *Brevibacillus* thermoruber.

For the purposes of the present invention, a "*Brevibacillus*-derived delta-endotoxin" is intended a toxin from *Brevibacillus* sp. that has toxic activity against one or more pests, including, but not limited to, members of the Lepidoptera and Coleoptera orders, or a protein that has homology to such a protein. Delta-endotoxin proteins have been isolated from other organisms, including *Bacillus thuringiensis*, *Clostridium bifermentans* and *Paenibacillus popilliae*. However, prior to the present invention, these sequences were not known to exist in *Brevibacillus* organisms. Thus, the present invention provides a new source for identifying and isolating genes of agricultural significance.

Delta-endotoxin proteins include amino acid sequences deduced from the full-length nucleotide sequences disclosed herein, and amino acid sequences that are shorter than the full-length sequences, either due to the use of an alternate downstream start site, or due to processing that produces a shorter protein having insecticidal activity. Processing may occur in the organism the protein is expressed in, or in the pest after ingestion of the protein.

Further encompassed herein are any delta-endotoxin sequence derived from a *Brevibacillus* organism. Delta-endotoxins include proteins identified as cry1 through cry43, cyt1 and cyt2, and Cyt-like toxin. There are currently over 250 known species of delta-endotoxins with a wide range of specificities and toxicities. For an expansive list see Crickmore et al. (1998), *Microbiol. Mol. Biol. Rev.* 62:807-813, and for regular updates see Crickmore et al. (2003) "*Bacillus thuringiensis* toxin nomenclature," on the world wide web at [biols.susx.ac.uk/Home/Neil\\_Crickmore/Bt/index](http://biols.susx.ac.uk/Home/Neil_Crickmore/Bt/index).

Provided herein are novel isolated nucleotide sequences that confer pesticidal activity against plant-pathogenic pests. Also provided are the amino acid sequences of the delta-endotoxin proteins. The protein resulting from translation of this gene allows cells to control or kill insects that ingest it. Isolated Nucleic Acid Molecules, and Variants and Fragments Thereof

One aspect of the invention pertains to isolated or recombinant nucleic acid molecules comprising nucleotide sequences encoding delta-endotoxin proteins and polypeptides or biologically active portions thereof, as well as nucleic

acid molecules sufficient for use as hybridization probes to identify delta-endotoxin encoding nucleic acids. As used herein, the term "nucleic acid molecule" is intended to include DNA molecules (e.g., recombinant DNA, cDNA or genomic DNA) and RNA molecules (e.g., mRNA) and analogs of the DNA or RNA generated using nucleotide analogs. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA.

An "isolated" nucleic acid sequence (or DNA) is used herein to refer to a nucleic acid sequence (or DNA) that is no longer in its natural environment, for example in an in vitro or in a recombinant bacterial or plant host cell. In some embodiments, an "isolated" nucleic acid is free of sequences (preferably protein encoding sequences) that naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For purposes of the invention, "isolated" when used to refer to nucleic acid molecules excludes isolated chromosomes. For example, in various embodiments, the isolated delta-endotoxin encoding nucleic acid molecule can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb, or 0.1 kb of nucleotide sequences that naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. In various embodiments, a delta-endotoxin protein that is substantially free of cellular material includes preparations of protein having less than about 30%, 20%, 10%, or 5% (by dry weight) of non-delta-endotoxin protein (also referred to herein as a "contaminating protein").

Nucleotide sequences encoding the proteins of the present invention include any delta-endotoxin derived from *Brevibacillus*. In various embodiments, the delta-endotoxin nucleotide sequence comprises the sequence set forth in SEQ ID NO:3, 6, or 9, and variants, fragments, and complements thereof. In some embodiments, the nucleotide sequence comprising SEQ ID NO:3, 6, or 9 is set forth in SEQ ID NO:1, 5, 8, 11, 12, 13, 14, or 15. In other embodiments, the variants and fragments of SEQ ID NO:3, 6, or 9 include the sequences corresponding to nucleotides 160-3819 of SEQ ID NO:1, nucleotides 4-1059 of SEQ ID NO:6, nucleotides 13-1059 of SEQ ID NO:6, nucleotides 151-1059 of SEQ ID NO:6, nucleotides 19-1893 of SEQ ID NO:9, nucleotides 46-1893 of SEQ ID NO:9, and nucleotides 52-1893 of SEQ ID NO:9, as well as variants and fragments of those sequences.

By "complement" is intended a nucleotide sequence that is sufficiently complementary to a given nucleotide sequence such that it can hybridize to the given nucleotide sequence to thereby form a stable duplex. The corresponding amino acid sequence for the delta-endotoxin protein encoded by this nucleotide sequence are set forth in SEQ ID NO:2, 4, 7, and 10.

Nucleic acid molecules that are fragments of these delta-endotoxin encoding nucleotide sequences are also encompassed by the present invention. By "fragment" is intended a portion of the nucleotide sequence encoding a delta-endotoxin protein. A fragment of a nucleotide sequence may encode a biologically active portion of a delta-endotoxin protein, or it may be a fragment that can be used as a hybridization probe or PCR primer using methods disclosed below. Nucleic acid molecules that are fragments of a delta-endotoxin nucleotide sequence comprise at least about 50, 100, 200, 300, 400, 500, 600, 700, 800, 900, 1000, 1050, 1100, 1150, 1200, 1250, 1300, 1350, 1400, 1450, 1500, 1550, 1600, 1650, 1700, 1750, 1800, 1850, 1900, 1950, 2000, 2050, 2100, 2150, 2200, 2250, 2300, 2350, 2400, 2450, 2500, 2550, 2600, 2650, 2700, 2750, 2800, 2850, 2900, 2950, 3000, 3050, 3100, 3150, 3200, 3250, 3300, 3350 contiguous nucleotides, or up

to the number of nucleotides present in a full-length delta-endotoxin encoding nucleotide sequence disclosed herein depending upon the intended use. By "contiguous" nucleotides is intended nucleotide residues that are immediately adjacent to one another. Fragments of the nucleotide sequences of the present invention will encode protein fragments that retain the biological activity of the delta-endotoxin protein and, hence, retain insecticidal activity. By "retains activity" is intended that the fragment will have at least about 30%, at least about 50%, at least about 70%, 80%, 90%, 95% or higher of the insecticidal activity of the delta-endotoxin protein. Methods for measuring insecticidal activity are well known in the art. See, for example, Czapla and Lang (1990) *J. Econ. Entomol.* 83:2480-2485; Andrews et al. (1988) *Biochem. J.* 252:199-206; Marrone et al. (1985) *J. of Economic Entomology* 78:290-293; and U.S. Pat. No. 5,743,477, all of which are herein incorporated by reference in their entirety.

A fragment of a delta-endotoxin encoding nucleotide sequence that encodes a biologically active portion of a protein of the invention will encode at least about 15, 25, 30, 50, 75, 100, 125, 150, 175, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1050, 1100 contiguous amino acids, or up to the total number of amino acids present in a full-length delta-endotoxin protein of the invention.

Preferred delta-endotoxin proteins of the present invention are encoded by a nucleotide sequence sufficiently identical to any of the nucleotide sequences disclosed herein. By "sufficiently identical" is intended an amino acid or nucleotide sequence that has at least about 60% or 65% sequence identity, about 70% or 75% sequence identity, about 80% or 85% sequence identity, about 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or greater sequence identity compared to a reference sequence using one of the alignment programs described herein using standard parameters. One of skill in the art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like.

To determine the percent identity of two amino acid sequences or of two nucleic acids, the sequences are aligned for optimal comparison purposes. The percent identity between the two sequences is a function of the number of identical positions shared by the sequences (i.e., percent identity = number of identical positions/total number of positions (e.g., overlapping positions) × 100). In one embodiment, the two sequences are the same length. In another embodiment, the comparison is across the entirety of the reference sequence (e.g., across the entirety of one of SEQ ID NO:1-11). The percent identity between two sequences can be determined using techniques similar to those described below, with or without allowing gaps. In calculating percent identity, typically exact matches are counted.

The determination of percent identity between two sequences can be accomplished using a mathematical algorithm. A nonlimiting example of a mathematical algorithm utilized for the comparison of two sequences is the algorithm of Karlin and Altschul (1990) *Proc. Natl. Acad. Sci. USA* 87:2264, modified as in Karlin and Altschul (1993) *Proc. Natl. Acad. Sci. USA* 90:5873-5877. Such an algorithm is incorporated into the BLASTN and BLASTX programs of Altschul et al. (1990) *J. Mol. Biol.* 215:403. BLAST nucleotide searches can be performed with the BLASTN program, score=100, wordlength=12, to obtain nucleotide sequences homologous to delta-endotoxin-like nucleic acid molecules of the invention. BLAST protein searches can be performed

with the BLASTX program, score=50, wordlength=3, to obtain amino acid sequences homologous to delta-endotoxin protein molecules of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST (in BLAST 2.0) can be utilized as described in Altschul et al. (1997) *Nucleic Acids Res.* 25:3389. Alternatively, PSI-Blast can be used to perform an iterated search that detects distant relationships between molecules. See Altschul et al. (1997) *supra*. When utilizing BLAST, Gapped BLAST, and PSI-Blast programs, the default parameters of the respective programs (e.g., BLASTX and BLASTN) can be used. Alignment may also be performed manually by inspection.

Another non-limiting example of a mathematical algorithm utilized for the comparison of sequences is the ClustalW algorithm (Higgins et al. (1994) *Nucleic Acids Res.* 22:4673-4680). ClustalW compares sequences and aligns the entirety of the amino acid or DNA sequence, and thus can provide data about the sequence conservation of the entire amino acid sequence. The ClustalW algorithm is used in several commercially available DNA/amino acid analysis software packages, such as the ALIGNX module of the Vector NTI Program Suite (Invitrogen Corporation, Carlsbad, Calif.). After alignment of amino acid sequences with ClustalW, the percent amino acid identity can be assessed. A non-limiting example of a software program useful for analysis of ClustalW alignments is GENEDOC™. GENEDOC™ (Karl Nicholas) allows assessment of amino acid (or DNA) similarity and identity between multiple proteins. Another non-limiting example of a mathematical algorithm utilized for the comparison of sequences is the algorithm of Myers and Miller (1988) *CABIOS* 4:11-17. Such an algorithm is incorporated into the ALIGN program (version 2.0), which is part of the GCG Wisconsin Genetics Software Package, Version 10 (available from Accelrys, Inc., 9685 Scranton Rd., San Diego, Calif., USA). When utilizing the ALIGN program for comparing amino acid sequences, a PAM120 weight residue table, a gap length penalty of 12, and a gap penalty of 4 can be used.

Unless otherwise stated, GAP Version 10, which uses the algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48(3):443-453, will be used to determine sequence identity or similarity using the following parameters: % identity and % similarity for a nucleotide sequence using GAP Weight of 50 and Length Weight of 3, and the nws gapdna.cmp scoring matrix; % identity or % similarity for an amino acid sequence using GAP weight of 8 and length weight of 2, and the BLOSUM62 scoring program. Equivalent programs may also be used. By "equivalent program" is intended any sequence comparison program that, for any two sequences in question, generates an alignment having identical nucleotide residue matches and an identical percent sequence identity when compared to the corresponding alignment generated by GAP Version 10.

The invention also encompasses variant nucleic acid molecules. "Variants" of the delta-endotoxin encoding nucleotide sequences include those sequences that encode the delta-endotoxin proteins disclosed herein but that differ conservatively because of the degeneracy of the genetic code as well as those that are sufficiently identical as discussed above. Naturally occurring allelic variants can be identified with the use of well-known molecular biology techniques, such as polymerase chain reaction (PCR) and hybridization techniques as outlined below. Variant nucleotide sequences also include synthetically derived nucleotide sequences that have been generated, for example, by using site-directed mutagenesis but which still encode the delta-endotoxin proteins disclosed in the present invention as discussed below. Variant proteins

encompassed by the present invention are biologically active, that is they continue to possess the desired biological activity of the native protein, that is, retaining insecticidal activity. By “retains activity” is intended that the variant will have at least about 30%, at least about 50%, at least about 70%, or at least about 80% of the insecticidal activity of the native protein. Methods for measuring insecticidal activity are well known in the art. See, for example, Czapla and Lang (1990) *J. Econ. Entomol.* 83: 2480-2485; Andrews et al. (1988) *Biochem. J.* 252:199-206; Marrone et al. (1985) *J. of Economic Entomology* 78:290-293; and U.S. Pat. No. 5,743,477, all of which are herein incorporated by reference in their entirety.

The skilled artisan will further appreciate that changes can be introduced by mutation of the nucleotide sequences of the invention thereby leading to changes in the amino acid sequence of the encoded delta-endotoxin proteins, without altering the biological activity of the proteins. Thus, variant isolated nucleic acid molecules can be created by introducing one or more nucleotide substitutions, additions, or deletions into the corresponding nucleotide sequence disclosed herein, such that one or more amino acid substitutions, additions or deletions are introduced into the encoded protein. Mutations can be introduced by standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis. Such variant nucleotide sequences are also encompassed by the present invention.

For example, conservative amino acid substitutions may be made at one or more predicted, nonessential amino acid residues. A “nonessential” amino acid residue is a residue that can be altered from the wild-type sequence of a delta-endotoxin protein without altering the biological activity, whereas an “essential” amino acid residue is required for biological activity. A “conservative amino acid substitution” is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (e.g., lysine, arginine, histidine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine, phenylalanine, tryptophan, histidine).

Delta-endotoxins generally have five conserved sequence domains, and three conserved structural domains (see, for example, de Maagd et al. (2001) *Trends Genetics* 17:193-199). The first conserved structural domain consists of seven alpha helices and is involved in membrane insertion and pore formation. Domain II consists of three beta-sheets arranged in a Greek key configuration, and domain III consists of two antiparallel beta-sheets in “jelly-roll” formation (de Maagd et al., 2001, supra). Domains II and III are involved in receptor recognition and binding, and are therefore considered determinants of toxin specificity.

Amino acid substitutions may be made in nonconserved regions that retain function. In general, such substitutions would not be made for conserved amino acid residues, or for amino acid residues residing within a conserved motif, where such residues are essential for protein activity. Examples of residues that are conserved and that may be essential for protein activity include, for example, residues that are identical between all proteins contained in an alignment of the amino acid sequences of the present invention and known delta-endotoxin sequences. Examples of residues that are conserved but that may allow conservative amino acid sub-

stitutions and still retain activity include, for example, residues that have only conservative substitutions between all proteins contained in an alignment of the amino acid sequences of the present invention and known delta-endotoxin sequences. However, one of skill in the art would understand that functional variants may have minor conserved or nonconserved alterations in the conserved residues.

Alternatively, variant nucleotide sequences can be made by introducing mutations randomly along all or part of the coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for ability to confer delta-endotoxin activity to identify mutants that retain activity. Following mutagenesis, the encoded protein can be expressed recombinantly, and the activity of the protein can be determined using standard assay techniques.

Using methods such as PCR, hybridization, and the like corresponding delta-endotoxin sequences can be identified, such sequences having substantial identity to the sequences of the invention. See, for example, Sambrook and Russell (2001) *Molecular Cloning: A Laboratory Manual*. (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.) and Innis, et al. (1990) *PCR Protocols: A Guide to Methods and Applications* (Academic Press, NY).

In a hybridization method, all or part of the delta-endotoxin nucleotide sequence can be used to screen cDNA or genomic libraries. Methods for construction of such cDNA and genomic libraries are generally known in the art and are disclosed in Sambrook and Russell, 2001, supra. The so-called hybridization probes may be genomic DNA fragments, cDNA fragments, RNA fragments, or other oligonucleotides, and may be labeled with a detectable group such as  $^{32}\text{P}$ , or any other detectable marker, such as other radioisotopes, a fluorescent compound, an enzyme, or an enzyme co-factor. Probes for hybridization can be made by labeling synthetic oligonucleotides based on the known delta-endotoxin-encoding nucleotide sequence disclosed herein. Degenerate primers designed on the basis of conserved nucleotides or amino acid residues in the nucleotide sequence or encoded amino acid sequence can additionally be used. The probe typically comprises a region of nucleotide sequence that hybridizes under stringent conditions to at least about 12, at least about 25, at least about 50, 75, 100, 125, 150, 175, 200, 250, 300, 350, or 400 consecutive nucleotides of delta-endotoxin encoding nucleotide sequence of the invention or a fragment or variant thereof. Methods for the preparation of probes for hybridization are generally known in the art and are disclosed in Sambrook and Russell, 2001, supra herein incorporated by reference.

For example, an entire delta-endotoxin sequence disclosed herein, or one or more portions thereof, may be used as a probe capable of specifically hybridizing to corresponding delta-endotoxin-like sequences and messenger RNAs. To achieve specific hybridization under a variety of conditions, such probes include sequences that are unique and are preferably at least about 10 nucleotides in length, or at least about 20 nucleotides in length. Such probes may be used to amplify corresponding delta-endotoxin sequences from a chosen organism by PCR. This technique may be used to isolate additional coding sequences from a desired organism or as a diagnostic assay to determine the presence of coding sequences in an organism. Hybridization techniques include hybridization screening of plated DNA libraries (either plaques or colonies; see, for example, Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.).

Hybridization of such sequences may be carried out under stringent conditions. By “stringent conditions” or “stringent

hybridization conditions" is intended conditions under which a probe will hybridize to its target sequence to a detectably greater degree than to other sequences (e.g., at least 2-fold over background). Stringent conditions are sequence-dependent and will be different in depending upon circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences that are 100% complementary to the probe can be identified (homologous probing). Alternatively, stringency conditions can be adjusted to allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous probing). Generally, a probe is less than about 1000 nucleotides in length, preferably less than 500 nucleotides in length.

Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30° C. for short probes (e.g., 10 to 50 nucleotides) and at least about 60° C. for long probes (e.g., greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37° C., and a wash in 1× to 2×SSC (20×SSC=3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55° C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1.0 M NaCl, 1% SDS at 37° C., and a wash in 0.5× to 1×SSC at 55 to 60° C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37° C., and a wash in 0.1×SSC at 60 to 65° C. Optionally, wash buffers may comprise about 0.1% to about 1% SDS. Duration of hybridization is generally less than about 24 hours, usually about 4 to about 12 hours.

Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the  $T_m$  can be approximated from the equation of Meinkoth and Wahl (1984) *Anal. Biochem.* 138:267-284:  $T_m = 81.5^\circ \text{C} + 16.6 (\log M) + 0.41 (\% \text{ GC}) - 0.61 (\% \text{ form}) - 500/L$ ; where M is the molarity of monovalent cations, % GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The  $T_m$  is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridizes to a perfectly matched probe.  $T_m$  is reduced by about 1° C. for each 1% of mismatching; thus,  $T_m$ , hybridization, and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with >90% identity are sought, the  $T_m$  can be decreased 10° C. Generally, stringent conditions are selected to be about 5° C. lower than the thermal melting point ( $T_m$ ) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4° C. lower than the thermal melting point ( $T_m$ ); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10° C. lower than the thermal melting point ( $T_m$ ); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20° C. lower than the thermal melting point ( $T_m$ ). Using the equation, hybridization and wash compositions, and desired  $T_m$ , those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a  $T_m$  of less than 45° C. (aqueous solution) or 32° C. (formamide solution), it is preferred to increase the SSC concentration so that a higher temperature can be used. An

extensive guide to the hybridization of nucleic acids is found in Tijssen (1993) *Laboratory Techniques in Biochemistry and Molecular Biology—Hybridization with Nucleic Acid Probes*, Part I, Chapter 2 (Elsevier, N.Y.); and Ausubel et al., eds. (1995) *Current Protocols in Molecular Biology*, Chapter 2 (Greene Publishing and Wiley-Interscience, New York). See Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.).

#### Isolated Proteins and Variants and Fragments Thereof

Delta-endotoxin proteins are also encompassed within the present invention. By "delta-endotoxin protein" is intended a protein having the amino acid sequence set forth in SEQ ID NO:2, 4, 7, or 10. Fragments, biologically active portions, and variants thereof are also provided, and may be used to practice the methods of the present invention. An "isolated protein" is used to refer to a protein that is no longer in its natural environment, for example in vitro or in a recombinant bacterial or plant host cell.

"Fragments" or "biologically active portions" include polypeptide fragments comprising amino acid sequences sufficiently identical to the amino acid sequence set forth in any of SEQ ID NO:2, 4, 7, or 10 and that exhibit insecticidal activity. A biologically active portion of a delta-endotoxin protein can be a polypeptide that is, for example, 10, 25, 50, 100 or more amino acids in length. Such biologically active portions can be prepared by recombinant techniques and evaluated for insecticidal activity. Methods for measuring insecticidal activity are well known in the art. See, for example, Czaplak and Lang (1990) *J. Econ. Entomol.* 83:2480-2485; Andrews et al. (1988) *Biochem. J.* 252:199-206; Marrone et al. (1985) *J. of Economic Entomology* 78:290-293; and U.S. Pat. No. 5,743,477, all of which are herein incorporated by reference in their entirety. As used here, a fragment comprises at least 8 contiguous amino acids of SEQ ID NO:2, 4, 7, or 10. In various embodiments, the fragments correspond to amino acids 54-1272 of SEQ ID NO:2, amino acids 21-651 of SEQ ID NO:4, amino acids 54-651 of SEQ ID NO:4, amino acids 2-352 of SEQ ID NO:7, amino acids 5-352 of SEQ ID NO:7, amino acids 51-352 of SEQ ID NO:7, amino acids 7-630 of SEQ ID NO:10, amino acids 16-630 of SEQ ID NO:10, amino acids 18-630 of SEQ ID NO:10, as well as variants and fragments thereof. The invention encompasses other fragments, however, such as any fragment in the protein greater than about 10, 20, 30, 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1050, 1100, 1150, 1200, 1250, or 1300 amino acids.

By "variants" is intended proteins or polypeptides having an amino acid sequence that is at least about 60%, 65%, about 70%, 75%, about 80%, 85%, about 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98% or 99% identical to the amino acid sequence of any of SEQ ID NO:2, 4, 7, or 10. Variants also include polypeptides encoded by a nucleic acid molecule that hybridizes to the nucleic acid molecule of SEQ ID NO:1, 3, 5, 6, 8, 9, 11, 12, 13, 14, or 15, or a complement thereof, under stringent conditions. Variants include polypeptides that differ in amino acid sequence due to mutagenesis. Variant proteins encompassed by the present invention are biologically active, that is they continue to possess the desired biological activity of the native protein, that is, retaining insecticidal activity. Methods for measuring insecticidal activity are well known in the art. See, for example, Czaplak and Lang (1990) *J. Econ. Entomol.* 83:2480-2485; Andrews et al. (1988) *Biochem. J.* 252:199-206; Marrone et al. (1985) *J. of*

*Economic Entomology* 78:290-293; and U.S. Pat. No. 5,743, 477, all of which are herein incorporated by reference in their entirety.

Bacterial genes, such as the axmi genes of this invention, quite often possess multiple methionine initiation codons in proximity to the start of the open reading frame. Often, translation initiation at one or more of these start codons will lead to generation of a functional protein. These start codons can include ATG codons. However, bacteria such as *Bacillus* sp. also recognize the codon GTG as a start codon, and proteins that initiate translation at GTG codons contain a methionine at the first amino acid. On rare occasions, translation in bacterial systems can initiate at a TTG codon, though in this event the TTG encodes a methionine. Furthermore, it is not often determined a priori which of these codons are used naturally in the bacterium. Thus, it is understood that use of one of the alternate methionine codons may also lead to generation of delta-endotoxin proteins that encode insecticidal activity. These delta-endotoxin proteins are encompassed in the present invention and may be used in the methods of the present invention. For example, the amino acid sequences corresponding to amino acids 54-1272 of SEQ ID NO:2, amino acids 21-651 of SEQ ID NO:4, amino acids 54-651 of SEQ ID NO:4, amino acids 2-352 of SEQ ID NO:7, amino acids 5-352 of SEQ ID NO:7, amino acids 51-352 of SEQ ID NO:7, amino acids 7-630 of SEQ ID NO:10, amino acids 16-630 of SEQ ID NO:10, amino acids 18-630 of SEQ ID NO:10, as well as variants and fragments thereof are encompassed herein. It will be understood that, when expressed in plants, it will be necessary to alter the alternate start codon to ATG for proper translation.

Antibodies to the polypeptides of the present invention, or to variants or fragments thereof, are also encompassed. Methods for producing antibodies are well known in the art (see, for example, Harlow and Lane (1988) *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.; U.S. Pat. No. 4,196,265).

#### Altered or Improved Variants

It is recognized that DNA sequences of a delta-endotoxin may be altered by various methods, and that these alterations may result in DNA sequences encoding proteins with amino acid sequences different than that encoded by a delta-endotoxin of the present invention. This protein may be altered in various ways including amino acid substitutions, deletions, truncations, and insertions of one or more amino acids of SEQ ID NO:2, 4, 7, or 10, including up to about 2, about 3, about 4, about 5, about 6, about 7, about 8, about 9, about 10, about 15, about 20, about 25, about 30, about 35, about 40, about 45, about 50, about 55, about 60, about 65, about 70, about 75, about 80, about 85, about 90, about 100, about 105, about 110, about 115, about 120, about 125, about 130 or more amino acid substitutions, deletions or insertions.

Methods for such manipulations are generally known in the art. For example, amino acid sequence variants of a delta-endotoxin protein can be prepared by mutations in the DNA. This may also be accomplished by one of several forms of mutagenesis and/or in directed evolution. In some aspects, the changes encoded in the amino acid sequence will not substantially affect the function of the protein. Such variants will possess the desired insecticidal activity. However, it is understood that the ability of a delta-endotoxin to confer insecticidal activity may be improved by the use of such techniques upon the compositions of this invention. For example, one may express a delta-endotoxin in host cells that exhibit high rates of base misincorporation during DNA replication, such as XL-1 Red (Stratagene). After propagation in such strains, one can isolate the delta-endotoxin DNA (for example by

preparing plasmid DNA, or by amplifying by PCR and cloning the resulting PCR fragment into a vector), culture the delta-endotoxin mutations in a non-mutagenic strain, and identify mutated delta-endotoxin genes with insecticidal activity, for example by performing an assay to test for insecticidal activity. Generally, the protein is mixed and used in feeding assays. See, for example Marrone et al. (1985) *J. of Economic Entomology* 78:290-293. Such assays can include contacting plants with one or more insects and determining the plant's ability to survive and/or cause the death of the insects. Examples of mutations that result in increased toxicity are found in Schnepf et al. (1998) *Microbiol. Mol. Biol. Rev.* 62:775-806.

Alternatively, alterations may be made to the protein sequence of many proteins at the amino or carboxy terminus without substantially affecting activity. This can include insertions, deletions, or alterations introduced by modern molecular methods, such as PCR, including PCR amplifications that alter or extend the protein coding sequence by virtue of inclusion of amino acid encoding sequences in the oligonucleotides utilized in the PCR amplification. Alternatively, the protein sequences added can include entire protein-coding sequences, such as those used commonly in the art to generate protein fusions. Such fusion proteins are often used to (1) increase expression of a protein of interest (2) introduce a binding domain, enzymatic activity, or epitope to facilitate either protein purification, protein detection, or other experimental uses known in the art (3) target secretion or translation of a protein to a subcellular organelle, such as the periplasmic space of Gram-negative bacteria, or the endoplasmic reticulum of eukaryotic cells, the latter of which often results in glycosylation of the protein.

Variant nucleotide and amino acid sequences of the present invention also encompass sequences derived from mutagenic and recombinogenic procedures such as DNA shuffling. With such a procedure, one or more different delta-endotoxin protein coding regions can be used to create a new delta-endotoxin protein possessing the desired properties. In this manner, libraries of recombinant polynucleotides are generated from a population of related sequence polynucleotides comprising sequence regions that have substantial sequence identity and can be homologously recombined in vitro or in vivo. For example, using this approach, sequence motifs encoding a domain of interest may be shuffled between a delta-endotoxin gene of the invention and other known delta-endotoxin genes to obtain a new gene coding for a protein with an improved property of interest, such as an increased insecticidal activity. Strategies for such DNA shuffling are known in the art. See, for example, Stemmer (1994) *Proc. Natl. Acad. Sci. USA* 91:10747-10751; Stemmer (1994) *Nature* 370:389-391; Cramer et al. (1997) *Nature Biotech.* 15:436-438; Moore et al. (1997) *J. Mol. Biol.* 272:336-347; Zhang et al. (1997) *Proc. Natl. Acad. Sci. USA* 94:4504-4509; Cramer et al. (1998) *Nature* 391:288-291; and U.S. Pat. Nos. 5,605,793 and 5,837,458.

Domain swapping or shuffling is another mechanism for generating altered delta-endotoxin proteins. Domains II and III may be swapped between delta-endotoxin proteins, resulting in hybrid or chimeric toxins with improved insecticidal activity or target spectrum. Methods for generating recombinant proteins and testing them for insecticidal activity are well known in the art (see, for example, Naimov et al. (2001) *Appl. Environ. Microbiol.* 67:5328-5330; de Maagd et al. (1996) *Appl. Environ. Microbiol.* 62:1537-1543; Ge et al. (1991) *J. Biol. Chem.* 266:17954-17958; Schnepf et al. (1990) *J. Biol. Chem.* 265:20923-20930; Rang et al. 91999) *Appl. Environ. Microbiol.* 65:2918-2925).

## Vectors

A delta-endotoxin sequence of the invention may be provided in an expression cassette for expression in a plant of interest. By "plant expression cassette" is intended a DNA construct that is capable of resulting in the expression of a protein from an open reading frame in a plant cell. Typically these contain a promoter and a coding sequence. Often, such constructs will also contain a 3' untranslated region. Such constructs may contain a "signal sequence" or "leader sequence" to facilitate co-translational or post-translational transport of the peptide to certain intracellular structures such as the chloroplast (or other plastid), endoplasmic reticulum, or Golgi apparatus.

By "signal sequence" is intended a sequence that is known or suspected to result in cotranslational or post-translational peptide transport across the cell membrane. In eukaryotes, this typically involves secretion into the Golgi apparatus, with some resulting glycosylation. By "leader sequence" is intended any sequence that when translated, results in an amino acid sequence sufficient to trigger co-translational transport of the peptide chain to a sub-cellular organelle. Thus, this includes leader sequences targeting transport and/or glycosylation by passage into the endoplasmic reticulum, passage to vacuoles, plastids including chloroplasts, mitochondria, and the like.

By "plant transformation vector" is intended a DNA molecule that is necessary for efficient transformation of a plant cell. Such a molecule may consist of one or more plant expression cassettes, and may be organized into more than one "vector" DNA molecule. For example, binary vectors are plant transformation vectors that utilize two non-contiguous DNA vectors to encode all requisite cis- and trans-acting functions for transformation of plant cells (Hellens and Mulineaux (2000) *Trends in Plant Science* 5:446-451). "Vector" refers to a nucleic acid construct designed for transfer between different host cells. "Expression vector" refers to a vector that has the ability to incorporate, integrate and express heterologous DNA sequences or fragments in a foreign cell. The cassette will include 5' and 3' regulatory sequences operably linked to a sequence of the invention. By "operably linked" is intended a functional linkage between a promoter and a second sequence, wherein the promoter sequence initiates and mediates transcription of the DNA sequence corresponding to the second sequence. Generally, operably linked means that the nucleic acid sequences being linked are contiguous and, where necessary to join two protein coding regions, contiguous and in the same reading frame. The cassette may additionally contain at least one additional gene to be cotransformed into the organism. Alternatively, the additional gene(s) can be provided on multiple expression cassettes.

"Promoter" refers to a nucleic acid sequence that functions to direct transcription of a downstream coding sequence. The promoter together with other transcriptional and translational regulatory nucleic acid sequences (also termed "control sequences") are necessary for the expression of a DNA sequence of interest.

Such an expression cassette is provided with a plurality of restriction sites for insertion of the delta-endotoxin sequence to be under the transcriptional regulation of the regulatory regions.

The expression cassette will include in the 5'-3' direction of transcription, a transcriptional and translational initiation region (i.e., a promoter), a DNA sequence of the invention, and a translational and transcriptional termination region (i.e., termination region) functional in plants. The promoter may be native, or analogous, or foreign or heterologous, to the

plant host and/or to the DNA sequence of the invention. Additionally, the promoter may be the natural sequence or alternatively a synthetic sequence. Where the promoter is "native" or "homologous" to the plant host, it is intended that the promoter is found in the native plant into which the promoter is introduced. Where the promoter is "foreign" or "heterologous" to the DNA sequence of the invention, it is intended that the promoter is not the native or naturally occurring promoter for the operably linked DNA sequence of the invention.

The termination region may be native with the transcriptional initiation region, may be native with the operably linked DNA sequence of interest, may be native with the plant host, or may be derived from another source (i.e., foreign or heterologous to the promoter, the DNA sequence of interest, the plant host, or any combination thereof). Convenient termination regions are available from the Ti-plasmid of *A. tumefaciens*, such as the octopine synthase and nopaline synthase termination regions. See also Guerineau et al. (1991) *Mol. Gen. Genet.* 262:141-144; Proudfoot (1991) *Cell* 64:671-674; Sanfacon et al. (1991) *Genes Dev.* 5:141-149; Mogen et al. (1990) *Plant Cell* 2:1261-1272; Munroe et al. (1990) *Gene* 91:151-158; Ballas et al. (1989) *Nucleic Acids Res.* 17:7891-7903; and Joshi et al. (1987) *Nucleic Acid Res.* 15:9627-9639.

Where appropriate, the gene(s) may be optimized for increased expression in the transformed host cell. That is, the genes can be synthesized using host cell-preferred codons for improved expression, or may be synthesized using codons at a host-preferred codon usage frequency. Generally, the GC content of the gene will be increased. See, for example, Campbell and Gowri (1990) *Plant Physiol.* 92:1-11 for a discussion of host-preferred codon usage. Methods are available in the art for synthesizing plant-preferred genes. See, for example, U.S. Pat. Nos. 5,380,831, and 5,436,391, and Murray et al. (1989) *Nucleic Acids Res.* 17:477-498, herein incorporated by reference.

In one embodiment, the delta-endotoxin is targeted to the chloroplast for expression. In this manner, where the delta-endotoxin is not directly inserted into the chloroplast, the expression cassette will additionally contain a nucleic acid encoding a transit peptide to direct the delta-endotoxin to the chloroplasts. Such transit peptides are known in the art. See, for example, Von Heijne et al. (1991) *Plant Mol. Biol. Rep.* 9:104-126; Clark et al. (1989) *J. Biol. Chem.* 264:17544-17550; Della-Cioppa et al. (1987) *Plant Physiol.* 84:965-968; Romer et al. (1993) *Biochem. Biophys. Res. Commun.* 196:1414-1421; and Shah et al. (1986) *Science* 233:478-481.

The delta-endotoxin gene to be targeted to the chloroplast may be optimized for expression in the chloroplast to account for differences in codon usage between the plant nucleus and this organelle. In this manner, the nucleic acids of interest may be synthesized using chloroplast-preferred codons. See, for example, U.S. Pat. No. 5,380,831, herein incorporated by reference.

## Plant Transformation

Methods of the invention involve introducing a nucleotide construct into a plant. By "introducing" is intended to present to the plant the nucleotide construct in such a manner that the construct gains access to the interior of a cell of the plant. The methods of the invention do not require that a particular method for introducing a nucleotide construct to a plant is used, only that the nucleotide construct gains access to the interior of at least one cell of the plant. Methods for introducing nucleotide constructs into plants are known in the art



including, but not limited to, stable transformation methods, transient transformation methods, and virus-mediated methods.

By "plant" is intended whole plants, plant organs (e.g., leaves, stems, roots, etc.), seeds, plant cells, propagules, embryos and progeny of the same. Plant cells can be differentiated or undifferentiated (e.g. callus, suspension culture cells, protoplasts, leaf cells, root cells, phloem cells, pollen).

"Transgenic plants" or "transformed plants" or "stably transformed" plants or cells or tissues refers to plants that have incorporated or integrated exogenous nucleic acid sequences or DNA fragments into the plant cell. These nucleic acid sequences include those that are exogenous, or not present in the untransformed plant cell, as well as those that may be endogenous, or present in the untransformed plant cell.

"Heterologous" generally refers to the nucleic acid sequences that are not endogenous to the cell or part of the native genome in which they are present, and have been added to the cell by infection, transfection, microinjection, electroporation, microporation, or the like.

Transformation of plant cells can be accomplished by one of several techniques known in the art. The delta-endotoxin gene of the invention may be modified to obtain or enhance expression in plant cells. Typically a construct that expresses such a protein would contain a promoter to drive transcription of the gene, as well as a 3' untranslated region to allow transcription termination and polyadenylation. The organization of such constructs is well known in the art. In some instances, it may be useful to engineer the gene such that the resulting peptide is secreted, or otherwise targeted within the plant cell. For example, the gene can be engineered to contain a signal peptide to facilitate transfer of the peptide to the endoplasmic reticulum. It may also be preferable to engineer the plant expression cassette to contain an intron, such that mRNA processing of the intron is required for expression.

Typically this "plant expression cassette" will be inserted into a "plant transformation vector". This plant transformation vector may be comprised of one or more DNA vectors needed for achieving plant transformation. For example, it is a common practice in the art to utilize plant transformation vectors that are comprised of more than one contiguous DNA segment. These vectors are often referred to in the art as "binary vectors". Binary vectors as well as vectors with helper plasmids are most often used for *Agrobacterium*-mediated transformation, where the size and complexity of DNA segments needed to achieve efficient transformation is quite large, and it is advantageous to separate functions onto separate DNA molecules. Binary vectors typically contain a plasmid vector that contains the cis-acting sequences required for T-DNA transfer (such as left border and right border), a selectable marker that is engineered to be capable of expression in a plant cell, and a "gene of interest" (a gene engineered to be capable of expression in a plant cell for which generation of transgenic plants is desired). Also present on this plasmid vector are sequences required for bacterial replication. The cis-acting sequences are arranged in a fashion to allow efficient transfer into plant cells and expression therein. For example, the selectable marker gene and the delta-endotoxin are located between the left and right borders. Often a second plasmid vector contains the trans-acting factors that mediate T-DNA transfer from *Agrobacterium* to plant cells. This plasmid often contains the virulence functions (Vir genes) that allow infection of plant cells by *Agrobacterium*, and transfer of DNA by cleavage at border sequences and vir-mediated DNA transfer, as is understood in the art (Hellen and Mullineaux (2000) *Trends in Plant Science* 5:446-

451). Several types of *Agrobacterium* strains (e.g. LBA4404, GV3101, EHA101, EHA105, etc.) can be used for plant transformation. The second plasmid vector is not necessary for transforming the plants by other methods such as microprojection, microinjection, electroporation, polyethylene glycol, etc.

In general, plant transformation methods involve transferring heterologous DNA into target plant cells (e.g. immature or mature embryos, suspension cultures, undifferentiated callus, protoplasts, etc.), followed by applying a maximum threshold level of appropriate selection (depending on the selectable marker gene) to recover the transformed plant cells from a group of untransformed cell mass. Explants are typically transferred to a fresh supply of the same medium and cultured routinely. Subsequently, the transformed cells are differentiated into shoots after placing on regeneration medium supplemented with a maximum threshold level of selecting agent. The shoots are then transferred to a selective rooting medium for recovering rooted shoot or plantlet. The transgenic plantlet then grows into a mature plant and produces fertile seeds (e.g. Hiei et al. (1994) *The Plant Journal* 6:271-282; Ishida et al. (1996) *Nature Biotechnology* 14:745-750). Explants are typically transferred to a fresh supply of the same medium and cultured routinely. A general description of the techniques and methods for generating transgenic plants are found in Ayres and Park (1994) *Critical Reviews in Plant Science* 13:219-239 and Bommineni and Jauhar (1997) *Maydica* 42:107-120. Since the transformed material contains many cells; both transformed and non-transformed cells are present in any piece of subjected target callus or tissue or group of cells. The ability to kill non-transformed cells and allow transformed cells to proliferate results in transformed plant cultures. Often, the ability to remove non-transformed cells is a limitation to rapid recovery of transformed plant cells and successful generation of transgenic plants.

Transformation protocols as well as protocols for introducing nucleotide sequences into plants may vary depending on the type of plant or plant cell, i.e., monocot or dicot, targeted for transformation. Generation of transgenic plants may be performed by one of several methods, including, but not limited to, microinjection, electroporation, direct gene transfer, introduction of heterologous DNA by *Agrobacterium* into plant cells (*Agrobacterium*-mediated transformation), bombardment of plant cells with heterologous foreign DNA adhered to particles, ballistic particle acceleration, aerosol beam transformation (U.S. Published Application No. 20010026941; U.S. Pat. No. 4,945,050; International Publication No. WO 91/00915; U.S. Published Application No. 2002015066), Lecl transformation, and various other non-particle direct-mediated methods to transfer DNA.

Methods for transformation of chloroplasts are known in the art. See, for example, Svab et al. (1990) *Proc. Natl. Acad. Sci. USA* 87:8526-8530; Svab and Maliga (1993) *Proc. Natl. Acad. Sci. USA* 90:913-917; Svab and Maliga (1993) *EMBO J.* 12:601-606. The method relies on particle gun delivery of DNA containing a selectable marker and targeting of the DNA to the plastid genome through homologous recombination. Additionally, plastid transformation can be accomplished by transactivation of a silent plastid-borne transgene by tissue-preferred expression of a nuclear-encoded and plastid-directed RNA polymerase. Such a system has been reported in McBride et al. (1994) *Proc. Natl. Acad. Sci. USA* 91:7301-7305.

Following integration of heterologous foreign DNA into plant cells, one then applies a maximum threshold level of appropriate selection in the medium to kill the untransformed cells and separate and proliferate the putatively transformed

cells that survive from this selection treatment by transferring regularly to a fresh medium. By continuous passage and challenge with appropriate selection, one identifies and proliferates the cells that are transformed with the plasmid vector. Molecular and biochemical methods can then be used to confirm the presence of the integrated heterologous gene of interest into the genome of the transgenic plant.

The cells that have been transformed may be grown into plants in accordance with conventional ways. See, for example, McCormick et al. (1986) *Plant Cell Reports* 5:81-84. These plants may then be grown, and either pollinated with the same transformed strain or different strains, and the resulting hybrid having constitutive expression of the desired phenotypic characteristic identified. Two or more generations may be grown to ensure that expression of the desired phenotypic characteristic is stably maintained and inherited and then seeds harvested to ensure expression of the desired phenotypic characteristic has been achieved. In this manner, the present invention provides transformed seed (also referred to as "transgenic seed") having a nucleotide construct of the invention, for example, an expression cassette of the invention, stably incorporated into their genome.

#### Evaluation of Plant Transformation

Following introduction of heterologous foreign DNA into plant cells, the transformation or integration of heterologous gene in the plant genome is confirmed by various methods such as analysis of nucleic acids, proteins and metabolites associated with the integrated gene.

PCR analysis is a rapid method to screen transformed cells, tissue or shoots for the presence of incorporated gene at the earlier stage before transplanting into the soil (Sambrook and Russell (2001) *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.). PCR is carried out using oligonucleotide primers specific to the gene of interest or *Agrobacterium* vector background, etc.

Plant transformation may be confirmed by Southern blot analysis of genomic DNA (Sambrook and Russell, 2001, supra). In general, total DNA is extracted from the transformant, digested with appropriate restriction enzymes, resolved in an agarose gel and transferred to a nitrocellulose or nylon membrane. The membrane or "blot" is then probed with, for example, radiolabeled <sup>32</sup>P target DNA fragment to confirm the integration of introduced gene into the plant genome according to standard techniques (Sambrook and Russell, 2001, supra).

In Northern blot analysis, RNA is isolated from specific tissues of transformant, fractionated in a formaldehyde agarose gel, and blotted onto a nylon filter according to standard procedures that are routinely used in the art (Sambrook and Russell, 2001, supra). Expression of RNA encoded by the delta-endotoxin is then tested by hybridizing the filter to a radioactive probe derived from a delta-endotoxin, by methods known in the art (Sambrook and Russell, 2001, supra).

Western blot, biochemical assays and the like may be carried out on the transgenic plants to confirm the presence of protein encoded by the delta-endotoxin gene by standard procedures (Sambrook and Russell, 2001, supra) using antibodies that bind to one or more epitopes present on the delta-endotoxin protein.

#### Insecticidal Activity in Plants

In another aspect of the invention, one may generate transgenic plants expressing a delta-endotoxin that has insecticidal activity. Methods described above by way of example may be utilized to generate transgenic plants, but the manner in which the transgenic plant cells are generated is not critical to this invention. Methods known or described in the art such as

*Agrobacterium*-mediated transformation, biolistic transformation, and non-particle-mediated methods may be used at the discretion of the experimenter. Plants expressing a delta-endotoxin may be isolated by common methods described in the art, for example by transformation of callus, selection of transformed callus, and regeneration of fertile plants from such transgenic callus. In such process, one may use any gene as a selectable marker so long as its expression in plant cells confers ability to identify or select for transformed cells.

A number of markers have been developed for use with plant cells, such as resistance to chloramphenicol, the aminoglycoside G418, hygromycin, or the like. Other genes that encode a product involved in chloroplast metabolism may also be used as selectable markers. For example, genes that provide resistance to plant herbicides such as glyphosate, bromoxynil, or imidazolinone may find particular use. Such genes have been reported (Stalker et al. (1985) *J. Biol. Chem.* 263:6310-6314 (bromoxynil resistance nitrilase gene); and Sathasivan et al. (1990) *Nucl. Acids Res.* 18:2188 (AHAS imidazolinone resistance gene). Additionally, the genes disclosed herein are useful as markers to assess transformation of bacterial or plant cells. Methods for detecting the presence of a transgene in a plant, plant organ (e.g., leaves, stems, roots, etc.), seed, plant cell, propagule, embryo or progeny of the same are well known in the art. In one embodiment, the presence of the transgene is detected by testing for insecticidal activity.

Fertile plants expressing a delta-endotoxin may be tested for insecticidal activity, and the plants showing optimal activity selected for further breeding. Methods are available in the art to assay for insect activity. Generally, the protein is mixed and used in feeding assays. See, for example Marrone et al. (1985) *J. of Economic Entomology* 78:290-293.

The present invention may be used for transformation of any plant species, including, but not limited to, monocots and dicots. Examples of plants of interest include, but are not limited to, corn (maize), sorghum, wheat, sunflower, tomato, crucifers, peppers, potato, cotton, rice, soybean, sugarbeet, sugarcane, tobacco, barley, and oilseed rape, *Brassica* sp., alfalfa, rye, millet, safflower, peanuts, sweet potato, cassava, coffee, coconut, pineapple, citrus trees, cocoa, tea, banana, avocado, fig, guava, mango, olive, papaya, cashew, *macadamia*, almond, oats, vegetables, ornamentals, and conifers.

Vegetables include, but are not limited to, tomatoes, lettuce, green beans, lima beans, peas, and members of the genus *Curcumis* such as cucumber, cantaloupe, and musk melon. Ornamentals include, but are not limited to, azalea, hydrangea, hibiscus, roses, tulips, daffodils, petunias, carnation, poinsettia, and chrysanthemum. Preferably, plants of the present invention are crop plants (for example, maize, sorghum, wheat, sunflower, tomato, crucifers, peppers, potato, cotton, rice, soybean, sugarbeet, sugarcane, tobacco, barley, oilseed rape, etc.).

#### Use in Insect Control

General methods for employing strains comprising a nucleotide sequence of the present invention, or a variant thereof, in insect control or in engineering other organisms as insecticidal agents are known in the art. See, for example U.S. Pat. No. 5,039,523 and EP 0480762A2.

The *Bacillus* strains containing a nucleotide sequence of the present invention, or a variant thereof, or the microorganisms that have been genetically altered to contain an insecticidal gene and protein may be used for protecting agricultural crops and products from insects. In one aspect of the invention, whole, i.e., unlysed, cells of a toxin (insecticide)-producing organism are treated with reagents that prolong the

activity of the toxin produced in the cell when the cell is applied to the environment of target insect(s).

Alternatively, the insecticide is produced by introducing a delta-endotoxin gene into a cellular host. Expression of the delta-endotoxin gene results, directly or indirectly, in the intracellular production and maintenance of the insecticide. In one aspect of this invention, these cells are then treated under conditions that prolong the activity of the toxin produced in the cell when the cell is applied to the environment of target insect(s). The resulting product retains the toxicity of the toxin. These naturally encapsulated insecticides may then be formulated in accordance with conventional techniques for application to the environment hosting a target insect, e.g., soil, water, and foliage of plants. See, for example EPA 0192319, and the references cited therein. Alternatively, one may formulate the cells expressing a gene of this invention such as to allow application of the resulting material as a insecticide.

#### Insecticidal Compositions

The active ingredients of the present invention are normally applied in the form of compositions and can be applied to the crop area or plant to be treated, simultaneously or in succession, with other compounds. These compounds can be fertilizers, weed killers, cryoprotectants, surfactants, detergents, insecticidal soaps, dormant oils, polymers, and/or time-release or biodegradable carrier formulations that permit long-term dosing of a target area following a single application of the formulation. They can also be selective herbicides, chemical insecticides, virucides, microbicides, amoebicides, pesticides, fungicides, bacteriocides, nematocides, molluscicides or mixtures of several of these preparations, if desired, together with further agriculturally acceptable carriers, surfactants or application-promoting adjuvants customarily employed in the art of formulation. Suitable carriers and adjuvants can be solid or liquid and correspond to the substances ordinarily employed in formulation technology, e.g. natural or regenerated mineral substances, solvents, dispersants, wetting agents, tackifiers, binders or fertilizers. Likewise the formulations may be prepared into edible "baits" or fashioned into pest "traps" to permit feeding or ingestion by a target pest of the insecticidal formulation.

Methods of applying an active ingredient of the present invention or an agrochemical composition of the present invention that contains at least one of the insecticidal proteins produced by the bacterial strains of the present invention include leaf application, seed coating and soil application. The number of applications and the rate of application depend on the intensity of infestation by the corresponding insect.

The composition may be formulated as a powder, dust, pellet, granule, spray, emulsion, colloid, solution, or such like, and may be prepared by such conventional means as desiccation, lyophilization, homogenation, extraction, filtration, centrifugation, sedimentation, or concentration of a culture of cells comprising the polypeptide. In all such compositions that contain at least one such insecticidal polypeptide, the polypeptide may be present in a concentration of from about 1% to about 99% by weight.

Lepidopteran, coleopteran, or other insects may be killed or reduced in numbers in a given area by the methods of the invention, or may be prophylactically applied to an environmental area to prevent infestation by a susceptible insect. Preferably the insect ingests, or is contacted with, a insecticidally-effective amount of the polypeptide. By "insecticidally-effective amount" is intended an amount of the insecticide that is able to bring about death to at least one insect, or to noticeably reduce insect growth, feeding, or normal physiological development. This amount will vary depending on

such factors as, for example, the specific target insects to be controlled, the specific environment, location, plant, crop, or agricultural site to be treated, the environmental conditions, and the method, rate, concentration, stability, and quantity of application of the insecticidally-effective polypeptide composition. The formulations may also vary with respect to climatic conditions, environmental considerations, and/or frequency of application and/or severity of insect infestation.

The insecticide compositions described may be made by formulating either the bacterial cell, crystal and/or spore suspension, or isolated protein component with the desired agriculturally-acceptable carrier. The compositions may be formulated prior to administration in an appropriate means such as lyophilized, freeze-dried, desiccated, or in an aqueous carrier, medium or suitable diluent, such as saline or other buffer. The formulated compositions may be in the form of a dust or granular material, or a suspension in oil (vegetable or mineral), or water or oil/water emulsions, or as a wettable powder, or in combination with any other carrier material suitable for agricultural application. Suitable agricultural carriers can be solid or liquid and are well known in the art. The term "agriculturally-acceptable carrier" covers all adjuvants, inert components, dispersants, surfactants, tackifiers, binders, etc. that are ordinarily used in insecticide formulation technology; these are well known to those skilled in insecticide formulation. The formulations may be mixed with one or more solid or liquid adjuvants and prepared by various means, e.g., by homogeneously mixing, blending and/or grinding the insecticidal composition with suitable adjuvants using conventional formulation techniques. Suitable formulations and application methods are described in U.S. Pat. No. 6,468,523, herein incorporated by reference.

"Pest" includes but is not limited to, insects, fungi, bacteria, nematodes, mites, ticks, and the like. Insect pests include insects selected from the orders Coleoptera, Diptera, Hymenoptera, Lepidoptera, Mallophaga, Homoptera, Hemiptera, Orthoptera, Thysanoptera, Dermaptera, Isoptera, Anoplura, Siphonaptera, Trichoptera, etc., particularly Coleoptera, Lepidoptera, and Diptera. In various embodiments, the pest does not include Dipteran pests.

The order Coleoptera includes the suborders Adephaga and Polyphaga. Suborder Adephaga includes the superfamilies Caraboidea and Gyrinoidea, while suborder Polyphaga includes the superfamilies Hydrophiloidea, Staphylinoidea, Cantharoidea, Cleroidea, Elateroidea, Dascilloidea, Dryopoidea, Byrrhoidea, Cucujoidea, Meloidea, Mordelloidea, Tenebrionoidea, Bostrichoidea, Scarabaeoidea, Cerambycoidea, Chrysomeloidea, and Curculionoidea. Superfamily Caraboidea includes the families Cicindelidae, Carabidae, and Dytiscidae. Superfamily Gyrinoidea includes the family Gyrinidae. Superfamily Hydrophiloidea includes the family Hydrophilidae. Superfamily Staphylinoidea includes the families Silphidae and Staphylinidae. Superfamily Cantharoidea includes the families Cantharidae and Lampyridae. Superfamily Cleroidea includes the families Cleridae and Dermestidae. Superfamily Elateroidea includes the families Elateridae and Buprestidae. Superfamily Cucujoidea includes the family Coccinellidae. Superfamily Meloidea includes the family Meloidea. Superfamily Tenebrionoidea includes the family Tenebrionidae. Superfamily Scarabaeoidea includes the families Passalidae and Scarabaeidae. Superfamily Cerambycoidea includes the family Cerambycidae. Superfamily Chrysomeloidea includes the family Chrysomelidae. Superfamily Curculionoidea includes the families Curculionidae and Scolytidae.

The order Diptera includes the Suborders Nematocera, Brachycera, and Cyclorrhapha. Suborder Nematocera

includes the families Tipulidae, Psychodidae, Culicidae, Ceratopogonidae, Chironomidae, Simuliidae, Bibionidae, and Cecidomyiidae. Suborder Brachycera includes the families Stratiomyidae, Tabanidae, Therevidae, Asilidae, Mydidae, Bombyliidae, and Dolichopodidae. Suborder Cyclorrhapha includes the Divisions Aschiza and Aschiza. Division Aschiza includes the families Phoridae, Syrphidae, and Conopidae. Division Aschiza includes the Sections Acalyptratae and Calyptratae. Section Acalyptratae includes the families Otitidae, Tephritidae, Agromyzidae, and Drosophilidae. Section Calyptratae includes the families Hippoboscidae, Oestridae, Tachinidae, Anthomyiidae, Muscidae, Caliphoridae, and Sarcophagidae.

The order Lepidoptera includes the families Papilionidae, Pieridae, Lycaenidae, Nymphalidae, Danaidae, Satyridae, Hesperidae, Sphingidae, Saturniidae, Geometridae, Arctidae, Noctuidae, Lymantriidae, Sesiidae, Crambidae, and Tineidae.

Nematodes include parasitic nematodes such as root-knot, cyst, and lesion nematodes, including *Heterodera* spp., *Meloidogyne* spp., and *Globodera* spp.; particularly members of the cyst nematodes, including, but not limited to, *Heterodera glycines* (soybean cyst nematode); *Heterodera schachtii* (beet cyst nematode); *Heterodera avenae* (cereal cyst nematode); and *Globodera rostochiensis* and *Globodera pallida* (potato cyst nematodes). Lesion nematodes include *Pratylenchus* spp.

Insect pests of the invention for the major crops include: Maize: *Ostrinia nubilalis*, European corn borer; *Agrotis ipsilon*, black cutworm; *Helicoverpa zea*, corn earworm; *Spodoptera frugiperda*, fall armyworm; *Diatraea grandiosella*, southwestern corn borer; *Elasmopalpus lignosellus*, lesser cornstalk borer; *Diatraea saccharalis*, sugarcane borer; *Diabrotica virgifera*, western corn rootworm; *Diabrotica longicornis barberi*, northern corn rootworm; *Diabrotica undecimpunctata howardi*, southern corn rootworm; *Melanotus* spp., wireworms; *Cyclocephala borealis*, northern masked chafer (white grub); *Cyclocephala immaculata*, southern masked chafer (white grub); *Popillia japonica*, Japanese beetle; *Chaetocnema pulicaria*, corn flea beetle; *Sphenophorus maidis*, maize billbug; *Rhopalosiphum maidis*, corn leaf aphid; *Anuraphis maidiradicis*, corn root aphid; *Blissus leucopterus leucopterus*, chinch bug; *Melanoplus femurrubrum*, redlegged grasshopper; *Melanoplus sanguinipes*, migratory grasshopper; *Hylemya platura*, seedcorn maggot; *Agromyza parvicornis*, corn blot leafminer; *Anaphothrips obscurus*, grass thrips; *Solenopsis milesta*, thief ant; *Tetranychus urticae*, twospotted spider mite; *Sorghum: Chilo partellus*, sorghum borer; *Spodoptera frugiperda*, fall armyworm; *Helicoverpa zea*, corn earworm; *Elasmopalpus lignosellus*, lesser cornstalk borer; *Feltia subterranea*, granulate cutworm; *Phyllophaga crinita*, white grub; *Eleodes*, *Conoderus*, and *Aeolus* spp., wireworms; *Oulema melanopus*, cereal leaf beetle; *Chaetocnema pulicaria*, corn flea beetle; *Sphenophorus maidis*, maize billbug; *Rhopalosiphum maidis*, corn leaf aphid; *Sipha flava*, yellow sugarcane aphid; *Blissus leucopterus leucopterus*, chinch bug; *Contarinia sorghicola*, sorghum midge; *Tetranychus cinnabarinus*, carmine spider mite; *Tetranychus urticae*, twospotted spider mite; Wheat: *Pseudaletia unipunctata*, army worm; *Spodoptera frugiperda*, fall armyworm; *Elasmopalpus lignosellus*, lesser cornstalk borer; *Agrotis orthogonia*, western cutworm; *Elasmopalpus lignosellus*, lesser cornstalk borer; *Oulema melanopus*, cereal leaf beetle; *Hypera punctata*, clover leaf weevil; *Diabrotica undecimpunctata howardi*, southern corn rootworm; Russian wheat aphid; *Schizaphis graminum*, greenbug; *Macrosiphum avenae*,

English grain aphid; *Melanoplus femurrubrum*, redlegged grasshopper; *Melanoplus differentialis*, differential grasshopper; *Melanoplus sanguinipes*, migratory grasshopper; *Mayetiola destructor*, Hessian fly; *Sitodiplosis mosellana*, wheat midge; *Meromyza americana*, wheat stem maggot; *Hylemya coarctata*, wheat bulb fly; *Frankliniella fusca*, tobacco thrips; *Cephus cinctus*, wheat stem sawfly; *Aceria tulipae*, wheat curl mite; Sunflower: *Suleima helianthana*, sunflower bud moth; *Homoeosoma electellum*, sunflower moth; *zyogramma exclamationis*, sunflower beetle; *Bothyrus gibbosus*, carrot beetle; *Neolasioptera murtfeldtiana*, sunflower seed midge; Cotton: *Heliothis virescens*, cotton budworm; *Helicoverpa zea*, cotton bollworm; *Spodoptera exigua*, beet armyworm; *Pectinophora gossypiella*, pink bollworm; *Anthonomus grandis*, boll weevil; *Aphis gossypii*, cotton aphid; *Pseudatomoscelis seriatus*, cotton fleahopper; *Trialeurodes abutilonea*, bandedwinged whitefly; *Lygus lineolaris*, tarnished plant bug; *Melanoplus femurrubrum*, redlegged grasshopper; *Melanoplus differentialis*, differential grasshopper; *Thrips tabaci*, onion thrips; *Frankliniella fusca*, tobacco thrips; *Tetranychus cinnabarinus*, carmine spider mite; *Tetranychus urticae*, twospotted spider mite; Rice: *Diatraea saccharalis*, sugarcane borer; *Spodoptera frugiperda*, fall armyworm; *Helicoverpa zea*, corn earworm; *Colaspis brunnea*, grape colaspis; *Lissorhoptrus oryzophilus*, rice water weevil; *Sitophilus oryzae*, rice weevil; *Nephotettix nigropictus*, rice leafhopper; *Blissus leucopterus leucopterus*, chinch bug; *Acrosternum hilare*, green stink bug; Soybean: *Pseudoplusia includens*, soybean looper; *Anticarsia gemmatilis*, velvetbean caterpillar; *Plathypena scabra*, green cloverworm; *Ostrinia nubilalis*, European corn borer; *Agrotis ipsilon*, black cutworm; *Spodoptera exigua*, beet armyworm; *Heliothis virescens*, cotton budworm; *Helicoverpa zea*, cotton bollworm; *Epilachna varivestis*, Mexican bean beetle; *Myzus persicae*, green peach aphid; *Empoasca fabae*, potato leafhopper; *Acrosternum hilare*, green stink bug; *Melanoplus femurrubrum*, redlegged grasshopper; *Melanoplus differentialis*, differential grasshopper; *Hylemya platura*, seedcorn maggot; *Sericothrips variabilis*, soybean thrips; *Thrips tabaci*, onion thrips; *Tetranychus turkestanii*, strawberry spider mite; *Tetranychus urticae*, twospotted spider mite; Barley: *Ostrinia nubilalis*, European corn borer; *Agrotis ipsilon*, black cutworm; *Schizaphis graminum*, greenbug; *Blissus leucopterus leucopterus*, chinch bug; *Acrosternum hilare*, green stink bug; *Euschistus servus*, brown stink bug; *Delia platura*, seedcorn maggot; *Mayetiola destructor*, Hessian fly; *Petrobia latens*, brown wheat mite; Oil Seed Rape: *Brevicoryne brassicae*, cabbage aphid; *Phyllotreta cruciferae*, Flea beetle; *Mamestra configurata*, Bertha armyworm; *Plutella xylostella*, Diamond-back moth; *Delia* spp., Root maggots.

#### Methods for Increasing Plant Yield

Methods for increasing plant yield are provided. The methods comprise introducing into a plant or plant cell a polynucleotide comprising an insecticidal sequence disclosed herein. As defined herein, the "yield" of the plant refers to the quality and/or quantity of biomass produced by the plant. By "biomass" is intended any measured plant product. An increase in biomass production is any improvement in the yield of the measured plant product. Increasing plant yield has several commercial applications. For example, increasing plant leaf biomass may increase the yield of leafy vegetables for human or animal consumption. Additionally, increasing leaf biomass can be used to increase production of plant-derived pharmaceutical or industrial products. An increase in yield can comprise any statistically significant increase including, but not limited to, at least a 1% increase, at least a

3% increase, at least a 5% increase, at least a 10% increase, at least a 20% increase, at least a 30%, at least a 50%, at least a 70%, at least a 100% or a greater increase in yield compared to a plant not expressing the insecticidal sequence.

In specific methods, plant yield is increased as a result of improved insect resistance of a plant expressing an insecticidal protein disclosed herein. Expression of the insecticidal protein results in a reduced ability of an insect to infest or feed on the plant, thus improving plant yield.

The plants can also be treated with one or more chemical compositions, including one or more herbicide, insecticides, or fungicides. Exemplary chemical compositions include: Fruits/Vegetables Herbicides: Atrazine, Bromacil, Diuron, Glyphosate, Linuron, Metribuzin, Simazine, Trifluralin, Fluzifop, Glufosinate, Halosulfuron Gowan, Paraquat, Propyzamide, Sethoxydim, Butafenacil, Halosulfuron, Indaziflam; Fruits/Vegetables Insecticides: Aldicarb, *Bacillus thuringiensis*, Carbaryl, Carbofuran, Chlorpyrifos, Cypermethrin, Deltamethrin, Diazinon, Malathion, Abamectin, Cyfluthrin/beta-cyfluthrin, Esfenvalerate, Lambda-cyhalothrin, Acequinocyl, Bifenazate, Methoxyfenozide, Novaluron, Chromafenozide, Thiacloprid, Dinotefuran, Flucyprym, Tolfenpyrad, Clothianidin, Spirodiclofen, Gamma-cyhalothrin, Spiromesifen, Spinosad, Rynaxypyr, Cyazypyr, Spinotram, Triflumuron, Spirotetramat, Imidacloprid, Flubendiamide, Thiodicarb, Metaflumizone, Sulfoxaflor, Cyflumetofen, Cyanopyrafen, Imidacloprid, Clothianidin, Thiamethoxam, Spinotram, Thiodicarb, Flonicamid, Methiocarb, Emamectin-benzoate, Indoxacarb, Forthiazate, Fenamiphos, Cadusaphos, Pyriproxifen, Fenbutatin-oxid, Hexthiazox, Methomyl, 4-[[[(6-Chlorpyridin-3-yl)methyl](2,2-difluorethyl)amino]furan-2(5H)-on]; Fruits/Vegetables Fungicides: Carbendazim, Chlorothalonil, EBDCs, Sulphur, Thiophanate-methyl, Azoxystrobin, Cymoxanil, Fluzazinam, Fosetyl, Iprodione, Kresoxim-methyl, Metalaxyl/mefenoxam, Trifloxystrobin, Ethaboxam, Iprovalicarb, Trifloxystrobin, Fenhexamid, Oxpoconazole fumarate, Cyazofamid, Fenamidone, Zoxamide, Picoxystrobin, Pyraclostrobin, Cyflufenamid, Boscalid; Cereals Herbicides: Isoproturon, Bromoxynil, Ioxynil, Phenoxies, Chlorsulfuron, Clodinafop, Diclofop, Diflufenican, Fenoxaprop, Florasulam, Fluoroxypyr, Metsulfuron, Triasulfuron, Flucarbazone, Iodosulfuron, Propoxycarbazone, Picolinafen, Mesosulfuron, Befflbutamid, Pinoxaden, Amidosulfuron, Thifensulfuron, Tribenuron, Flupyrasulfuron, Sulfosulfuron, Pyrasulfotole, Pyroxulam, Flufenacet, Tralkoxydim, Pyroxasulfon; Cereals Fungicides: Carbendazim, Chlorothalonil, Azoxystrobin, Cyproconazole, Cyprodinil, Fenpropimorph, Epoxiconazole, Kresoxim-methyl, Quinoxifen, Tebuconazole, Trifloxystrobin, Simeconazole, Picoxystrobin, Pyraclostrobin, Dimoxystrobin, Prothioconazole, Fluoxastrobin; Cereals Insecticides: Dimethoate, Lambda-cyhalothrin, Deltamethrin, alpha-Cypermethrin, beta-cyfluthrin, Bifenthrin, Imidacloprid, Clothianidin, Thiamethoxam, Thiacloprid, Acetamiprid, Dinotefuran, Chlorpyrifos, Metamidophos, Oxidemethon-methyl, Pirimicarb, Methiocarb; Maize Herbicides: Atrazine, Alachlor, Bromoxynil, Acetochlor, Dicamba, Clopyralid, (S-)Dimethenamid, Glufosinate, Glyphosate, Isoxaflutole, (S-)Metolachlor, Mesotrione, Nicosulfuron, Primisulfuron, Rimsulfuron, Sulcotrione, Foramsulfuron, Topramezone, Tembotrione, Saflufenacil, Thiencarbazone, Flufenacet, Pyroxasulfon; Maize Insecticides Carbofuran, Chlorpyrifos, Bifenthrin, Fipronil, Imidacloprid, Lambda-Cyhalothrin, Tefluthrin, Terbufos, Thiamethoxam, Clothianidin, Spiromesifen, Flubendiamide, Triflumuron, Rynaxypyr, Deltamethrin, Thiodicarb, beta-Cyfluthrin, Cypermethrin, Bifenthrin, Lufenuron, Triflumuron, Tefluthrin, Tebupirim-

phos, Ethiprole, Cyazypyr, Thiacloprid, Acetamiprid, Dinotefuran, Avermectin, Methiocarb, Spirodiclofen, Spirotetramat; Maize Fungicides: Fenitropan, Thiram, Prothioconazole, Tebuconazole, Trifloxystrobin; Rice Herbicides: Butachlor, Propanil, Azimsulfuron, Bensulfuron, Cyhalofop, Daimuron, Fentrazamide, Imazosulfuron, Mefenacet, Oxaziclomefone, Pyrazosulfuron, Pyributicarb, Quinclorac, Thiobencarb, Indanofan, Flufenacet, Fentrazamide, Halosulfuron, Oxaziclomefone, Benzobicyclon, Pyrifalid, Penoxsulam, Bispyribac, Oxadiargyl, Ethoxysulfuron, Pretilachlor, Mesotrione, Tefuryltrione, Oxadiazone, Fenoxaprop, Pyrimisulfan; Rice Insecticides: Diazinon, Fenitrothion, Fenobucarb, Monocrotophos, Benfuracarb, Buprofezin, Dinotefuran, Fipronil, Imidacloprid, Isoprocarb, Thiacloprid, Chromafenozide, Thiacloprid, Dinotefuran, Clothianidin, Ethiprole, Flubendiamide, Rynaxypyr, Deltamethrin, Acetamiprid, Thiamethoxam, Cyazypyr, Spinosad, Spinotram, Emamectin-Benzoate, Cypermethrin, Chlorpyrifos, Cartap, Methamidophos, Etofenprox, Triazophos, 4-[[[(6-Chlorpyridin-3-yl)methyl](2,2-difluorethyl)amino]furan-2(5H)-on, Carbofuran, Benfuracarb; Rice Fungicides: Thiophanate-methyl, Azoxystrobin, Carpropamid, Edifenphos, Ferimzone, Iprobenfos, Isoprothiolane, Pencycuron, Probenazole, Pyroquilon, Tricyclazole, Trifloxystrobin, Diclocymet, Fenoxanil, Simeconazole, Tiadinil; Cotton Herbicides: Diuron, Fluometuron, MSMA, Oxyfluorfen, Prometryn, Trifluralin, Carfentrazone, Clethodim, Fluzifop-butyl, Glyphosate, Norflurazon, Pendimethalin, Pyriothibac-sodium, Trifloxysulfuron, Tepraloxym, Glufosinate, Flumioxazin, Thidiazuron; Cotton Insecticides: Acephate, Aldicarb, Chlorpyrifos, Cypermethrin, Deltamethrin, Malathion, Monocrotophos, Abamectin, Acetamiprid, Emamectin Benzoate, Imidacloprid, Indoxacarb, Lambda-Cyhalothrin, Spinosad, Thiodicarb, Gamma-Cyhalothrin, Spiromesifen, Pyridalyl, Flonicamid, Flubendiamide, Triflumuron, Rynaxypyr, Beta-Cyfluthrin, Spirotetramat, Clothianidin, Thiamethoxam, Thiacloprid, Dinotefuran, Flubendiamide, Cyazypyr, Spinosad, Spinotram, gamma Cyhalothrin, 4-[[[(6-Chlorpyridin-3-yl)methyl](2,2-difluorethyl)amino]furan-2(5H)-on, Thiodicarb, Avermectin, Flonicamid, Pyridalyl, Spiromesifen, Sulfoxaflor, Profenophos, Thiazophos, Endosulfan; Cotton Fungicides: Etridiazole, Metalaxyl, Quintozene; Soybean Herbicides: Alachlor, Bentazone, Trifluralin, Chlorimuron-Ethyl, Cloransulam-Methyl, Fenoxaprop, Fomesafen, Fluzifop, Glyphosate, Imazamox, Imazaquin, Imazethapyr, (S-)Metolachlor, Metribuzin, Pendimethalin, Tepraloxym, Glufosinate; Soybean Insecticides: Lambda-cyhalothrin, Methomyl, Parathion, Thiocarb, Imidacloprid, Clothianidin, Thiamethoxam, Thiacloprid, Acetamiprid, Dinotefuran, Flubendiamide, Rynaxypyr, Cyazypyr, Spinosad, Spinotram, Emamectin-Benzoate, Fipronil, Ethiprole, Deltamethrin, beta-Cyfluthrin, gamma and lambda Cyhalothrin, 4-[[[(6-Chlorpyridin-3-yl)methyl](2,2-difluorethyl)amino]furan-2(5H)-on, Spirotetramat, Spinodiclofen, Triflumuron, Flonicamid, Thiodicarb, beta-Cyfluthrin; Soybean Fungicides: Azoxystrobin, Cyproconazole, Epoxiconazole, Flutriafol, Pyraclostrobin, Tebuconazole, Trifloxystrobin, Prothioconazole, Tetraconazole; Sugarbeet Herbicides: Chloridazon, Desmedipham, Ethofumesate, Phenmedipham, Triallate, Clopyralid, Fluzifop, Lenacil, Metamitron, Quinmerac, Cycloxydim, Triflusulfuron, Tepraloxym, Quizalofop; Sugarbeet Insecticides: Imidacloprid, Clothianidin, Thiamethoxam, Thiacloprid, Acetamiprid, Dinotefuran, Deltamethrin, beta-Cyfluthrin, gamma/lambda Cyhalothrin, 4-[[[(6-Chlorpyridin-3-yl)methyl](2,2-difluorethyl)amino]furan-2(5H)-on, Tefluthrin, Rynaxypyr, Cyaxypyr, Fipronil,

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Carbofuran; Canola Herbicides: Clopyralid, Diclofop, Fluzifop, Glufosinate, Glyphosate, Metazachlor, Trifluralin Ethametsulfuron, Quinmerac, Quizalofop, Clethodim, Tepraloxymid; Canola Fungicides: Azoxystrobin, Carbendazim, Fludioxonil, Iprodione, Prochloraz, Vinclozolin; Canola Insecticides: Carbofuran, Organophosphates, Pyrethroids, Thiocloprid, Deltamethrin, Imidacloprid, Clothianidin, Thiamethoxam, Acetamiprid, Dinotofuran,  $\beta$ -Cyfluthrin, gamma and lambda Cyhalothrin, tau-Fluvalerate, Ethiprole, Spinosad, Spinetoram, Flubendiamide, Rynaxypyr, Cyazypyr, 4-[[[(6-Chlorpyridin-3-yl)methyl](2,2-difluoroethyl)amino]furan-2(5H)-on.

The following examples are offered by way of illustration and not by way of limitation.

EXPERIMENTAL

Example 1

Identification of Axmi-134 from *Brevibacillus* Strain ATX15530

The complete gene sequence was identified as follows:  
Preparation of extrachromosomal DNA from the strain. Extrachromosomal DNA contains a mixture of some or all of the following: plasmids of various size; phage chromosomes; genomic DNA fragments not separated by the purification protocol; other uncharacterized extrachromosomal molecules.  
Mechanical or enzymatic shearing of the extrachromosomal DNA to generate size-distributed fragments.  
Sequencing of the fragmented DNA by high-throughput pyrosequencing methods. Identification of putative toxin genes via homology and/or other computational analyses.  
When required, sequence finishing of the gene of interest by one of several PCR or cloning strategies (e.g. TAIL-PCR). The sequence of the axmi-134 open reading frame is provided herein as SEQ ID NO:1 and encodes the AXMI-134 protein (SEQ ID NO: 2. Comparison of AXMI-134 vs protein databases identified the following homologies:  
Known homologs and approximate percent identity:  
Cry43Aa1—60%  
Cry43Aa1—60%  
Cry43Ba1—55%

Example 2

Heterologous Expression of AXMI-134

The complete ORF of axmi-134 (3.82 kb which encodes 1272 amino acid long protein) was amplified from *Brevibacillus* strain ATX15530 using appropriate primers. It was cloned into an *E. coli* expression vector based on pRSF1b (to give pAX5479) and a *Bacillus* vector based on pAX916 (to give pAX5481). The resulting clones were confirmed by restriction analysis and finally, by complete sequencing of the cloned gene. For expression in *E. coli*, BL21\*DE3 was transformed with pAX5479. Single colony was inoculated in LB supplemented with kanamycin and grown overnight at 37° C. The following day, fresh medium was inoculated in duplicate with 1% of overnight culture and grown at 37° C. to logarithmic phase. Subsequently, cultures were induced with 1 mM IPTG for 3 hours at 37° C. or overnight at 20° C. Each cell pellet was suspended in 50 mM sodium carbonate buffer, pH 10.5 supplemented with 1 mM DTT and sonicated. Analysis by SDS-PAGE detected expression of a 144 kD protein corresponding to Axmi134. For expression in *Bacillus*, a labo-

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ratory strain of *Bacillus* was transformed with pAX5481 and a single colony was grown in CYS-glu medium for 3 days to sporulation. Cell pellet was then extracted with 50 mM sodium carbonate buffer, pH 10.5 supplemented with 1 mM DTT. Soluble fraction showed presence of a ~144 kD Axmi134 protein. Trypsinization of Axmi134 cleaved the full length protein, resulting in two protein fragments of about 85 kDa and 65 kDa.

Example 3

Pesticidal Activity of AXMI-134

Soluble extracts containing Axmi134 were tested separately in insect assays with appropriate controls at an approximate concentration of 200 ng/ul. In this assay, a 5 day read of the plates showed AXMI-134 to have pesticidal activity on diamondback moth (DBM) and Colorado potato beetle (CPB) (see Table 2 below). Trypsin treatment of AXMI-134 resulted in strong mortality and stunting on DBM and CPB, (Table 2) respectively. Table 1 shows a description of the scoring assignments used herein

TABLE 1

Description of Scoring System	
Score	Description
0	no effect observed
1	mild non-uniform stunting
2	moderate non-uniform stunting
3	moderate to severe uniform stunting
4	mortality (<100%) with uniform stunting
5	complete mortality

TABLE 2

Activity of AXMI134 on Diamondback moth ( <i>Plutella xylostella</i> ) and Colorado potato beetle ( <i>Leptinotarsa decemlineata</i> ).		
Pest	AXMI134	AXMI134 (Trypsin treated)
Diamondback Moth	4 - Uniform stunt, 60% mortality	4 - Uniform stunt, 80% mortality
Colorado Potato Beetle	3 - Light feeding on leaf disks, 100% mortality	5 - Little or no feeding on leaf disks, 100% mortality

Example 4

N-Terminal Deletions of AXMI-134

A series of N-terminal deletions of AXMI-134 were planned and performed using PCR-based cloning. The deletion mutants thus generated were cloned and expressed, and resulting protein products were tested for activity against selected agricultural pests. An 'ATG' start codon, encoding methionine (M), was added at the beginning of each variant. Listed here are the start sites of the deletion variants and their starting amino acid sequences (corresponding to SEQ ID NO:2).

TABLE 3

Location of Protein start sites in N-terminal Deletions	
Deletion	Starting position Relative to SEQ ID NO:2
D3	21
D5	39
D6	55
D7	63
D8	73
D9	83
D11	103
D12	119

All deletion mutants were PCR amplified from construct pAX5481 (which contains the full-length axmi-134 gene) and cloned into the BamHI site in the expression vector pAX916. All constructs were transformed into host cells and grown to measure the expression of the truncated variants. Soluble protein was extracted in 50 mM sodium carbonate buffer, pH 10.5. Whole culture (W) and soluble (S) fractions were analyzed by SDS-PAGE.

Deletions at the N-terminus had differing effects on the solubility of AXMI-134. Variant D3, cloned into pAX4245 and starting at position 21 of the native AXMI-134 amino acid sequence, showed enhanced solubility as estimated by PAGE analysis. Variants D5, D6, D7, D8, D9, and D12 expressed poorly and produced little or no soluble protein under the conditions tested. Variant D11 expressed and produced soluble protein at levels comparable to the wild-type AXMI-134 protein.

Soluble fractions were assayed at three concentrations (100, 50, and 10 ng per microliter) against Colorado Potato Beetle (CPB) using leaf-dip assays. Results are presented in Table 3. Scores are presented as means of 3 replicate experiments, with individual component scores presented parenthetically. The scoring key for Table 4 is the same as for Table 1 above.

Variant D3 (cloned into pAX4245) showed enhanced activity versus the wild-type against CPB in replicate assays. The LC50 for variant D3 was found to be 52.5 ng/ml and the LC50 for the wild-type was found to be 800 ng/ml for CPB. The remaining variants showed no activity in this assay.

TABLE 4

Activity of Axmi134 variants against Colorado Potato Beetle			
Form of Axmi134	100 ng/ul	50 ng/ul	10 ng/ul
Full length (pAX5481)	2.33(2,2,3)	2.67(2,3,3)	0
Deleted form D3 (pAX4245)	4.67(5,4,5)	3.67(3,4,4)	2(2,2,2)

N-terminal deletions may be of general utility in designing active variants of Cry-type delta endotoxins. The N-terminal portion of AXMI-134 contained multiple asparagine (N) and glutamine (Q) residues. Other Cry-type proteins have sequences near the N terminus that contain similar polar residues. Removal of this portion of the protein may facilitate toxin activation.

Example 5

#### Identification of Axmi-159 from *Brevibacillus* Strain ATX15530

The complete gene sequence was identified as follows:

Preparation of extrachromosomal DNA from the strain. Extrachromosomal DNA contains a mixture of some or all of the following: plasmids of various size; phage chromosomes; genomic DNA fragments not separated by the purification protocol; other uncharacterized extrachromosomal molecules.

Mechanical or enzymatic shearing of the extrachromosomal DNA to generate size-distributed fragments.

Sequencing of the fragmented DNA by high-throughput pyrosequencing methods. Identification of putative toxin genes via homology and/or other computational analyses.

When required, sequence finishing of the gene of interest by one of several PCR or cloning strategies (e.g. TAIL-PCR).

The sequence of the axmi-159 open reading frame is provided herein as SEQ ID NO:6 and encodes the AXMI-159 protein (SEQ ID NO:7). Comparison of AXMI-159 vs protein databases identified the following homologies:

AXMI-159 homologs and approximate percent identity:

Axmi012—24.0%

Cry35Ba1—21.1%

Cry35Ac1—20.5%

Example 6

#### Identification of Axmi-160 from *Brevibacillus* Strain ATX15530

The complete gene sequence was identified as follows:

Preparation of extrachromosomal DNA from the strain. Extrachromosomal DNA contains a mixture of some or all of the following: plasmids of various size; phage chromosomes; genomic DNA fragments not separated by the purification protocol; other uncharacterized extrachromosomal molecules.

Mechanical or enzymatic shearing of the extrachromosomal DNA to generate size-distributed fragments.

Sequencing of the fragmented DNA by high-throughput pyrosequencing methods. Identification of putative toxin genes via homology and/or other computational analyses.

When required, sequence finishing of the gene of interest by one of several PCR or cloning strategies (e.g. TAIL-PCR).

The sequence of the axmi-160 open reading frame is provided herein as SEQ ID NO:9 and encodes the AXMI-160 protein (SEQ ID NO:10). Comparison of AXMI-160 vs protein databases identified the following homologies:

Known homologs and approximate percent identity:

lethal factor—20.5%

Vip2Ba1—20.7%

Example 7

#### Synthetic Genes

Alternate DNA sequences encoding the proteins of the invention were developed. They are provided as follows:

Encoded Protein	SEQ ID NO of	
	Synthetic Gene	Corresponding amino acid encoded
AXMI-134(v01)	12	amino acids 1-677 of SEQ ID NO:2
AXMI-134(v02)	13	amino acids 21-662 of SEQ ID NO:2 with N-terminal methionine addition
AXMI-159	14	amino acids 1-352 of SEQ ID NO:7
AXMI-160	15	amino acids 1-630 of SEQ ID NO:10

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## Example 8

## Additional Assays for Pesticidal Activity

The ability of an insecticidal protein to act as a pesticide upon a pest is often assessed in a number of ways. One way well known in the art is to perform a feeding assay. In such a feeding assay, one exposes the pest to a sample containing either compounds to be tested, or control samples. Often this is performed by placing the material to be tested, or a suitable dilution of such material, onto a material that the pest will ingest, such as an artificial diet. The material to be tested may be in a liquid, solid, or slurry form. The material to be tested may be placed upon the surface and then allowed to dry or incorporate into the diet. Alternatively, the material to be tested may be mixed with a molten artificial diet, then dispensed into the assay chamber. The assay chamber may be, for example, a cup, a dish, or a well of a microtiter plate.

Assays for sucking pests (for example aphids) may involve separating the test material from the insect by a partition, ideally a portion that can be pierced by the sucking mouth parts of the sucking insect, to allow ingestion of the test material. Often the test material is mixed with a feeding stimulant, such as sucrose, to promote ingestion of the test compound.

Other types of assays can include microinjection of the test material into the mouth, or gut of the pest, as well as development of transgenic plants, followed by test of the ability of the pest to feed upon the transgenic plant. Plant testing may involve isolation of the plant parts normally consumed, for example, small cages attached to a leaf, or isolation of entire plants in cages containing insects.

Other methods and approaches to assay pests are known in the art, and can be found, for example in Robertson, J. L. & H. K. Preisler. 1992. *Pesticide bioassays with arthropods*. CRC, Boca Raton, Fla. Alternatively, assays are commonly described in the journals "Arthropod Management Tests" and "Journal of Economic Entomology" or by discussion with members of the Entomological Society of America (ESA).

## Example 9

## Vectoring of the Insecticidal Genes of the Invention for Plant Expression

Each of the coding regions of the genes of the invention is connected independently with appropriate promoter and terminator sequences for expression in plants. Such sequences are well known in the art and may include the rice actin promoter or maize ubiquitin promoter for expression in monocots, the *Arabidopsis* UBQ3 promoter or CaMV 35S promoter for expression in dicots, and the nos or PinII terminators. Techniques for producing and confirming promoter-gene-terminator constructs also are well known in the art.

## Example 10

Transformation of the Genes of the Invention into Plant Cells by *Agrobacterium*-Mediated Transformation

Ears are collected 8-12 days after pollination. Embryos are isolated from the ears, and those embryos 0.8-1.5 mm in size are used for transformation. Embryos are plated scutellum side-up on a suitable incubation media, and incubated overnight at 25° C. in the dark. However, it is not necessary per se to incubate the embryos overnight. Embryos are contacted

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with an *Agrobacterium* strain containing the appropriate vectors for Ti plasmid mediated transfer for 5-10 min, and then plated onto co-cultivation media for 3 days (25° C. in the dark). After co-cultivation, explants are transferred to recovery period media for five days (at 25° C. in the dark). Explants are incubated in selection media for up to eight weeks, depending on the nature and characteristics of the particular selection utilized. After the selection period, the resulting callus is transferred to embryo maturation media, until the formation of mature somatic embryos is observed. The resulting mature somatic embryos are then placed under low light, and the process of regeneration is initiated as known in the art. The resulting shoots are allowed to root on rooting media, and the resulting plants are transferred to nursery pots and propagated as transgenic plants.

## Example 11

## Transformation of Maize Cells with the Insecticidal Genes of the Invention

Maize ears are collected 8-12 days after pollination. Embryos are isolated from the ears, and those embryos 0.8-1.5 mm in size are used for transformation. Embryos are plated scutellum side-up on a suitable incubation media, such as DN62A5S media (3.98 g/L N6 Salts; 1 mL/L (of 1000x Stock) N6 Vitamins; 800 mg/L L-Asparagine; 100 mg/L Myo-inositol; 1.4 g/L L-Proline; 100 mg/L Casaminoacids; 50 g/L sucrose; 1 mL/L (of 1 mg/mL Stock) 2,4-D), and incubated overnight at 25° C. in the dark.

The resulting explants are transferred to mesh squares (30-40 per plate), transferred onto osmotic media for 30-45 minutes, then transferred to a beaming plate (see, for example, PCT Publication No. WO/0138514 and U.S. Pat. No. 5,240,842).

DNA constructs designed to express the genes of the invention in plant cells are accelerated into plant tissue using an aerosol beam accelerator, using conditions essentially as described in PCT Publication No. WO/0138514. After beaming, embryos are incubated for 30 min on osmotic media, then placed onto incubation media overnight at 25° C. in the dark. To avoid unduly damaging beamed explants, they are incubated for at least 24 hours prior to transfer to recovery media. Embryos are then spread onto recovery period media, for 5 days, 25° C. in the dark, then transferred to a selection media. Explants are incubated in selection media for up to eight weeks, depending on the nature and characteristics of the particular selection utilized. After the selection period, the resulting callus is transferred to embryo maturation media, until the formation of mature somatic embryos is observed. The resulting mature somatic embryos are then placed under low light, and the process of regeneration is initiated by methods known in the art. The resulting shoots are allowed to root on rooting media, and the resulting plants are transferred to nursery pots and propagated as transgenic plants.

## Materials

DN62A5S Media		
Components	per liter	Source
Chu's N6 Basal Salt Mixture (Prod. No. C 416)	3.98 g/L	Phytotechnology Labs
Chu's N6 Vitamin Solution (Prod. No. C 149)	1 mL/L (of 1000x Stock)	Phytotechnology Labs
L-Asparagine	800 mg/L	Phytotechnology Labs
Myo-inositol	100 mg/L	Sigma



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-continued

DN62A5S Media		
Components	per liter	Source
L-Proline	1.4 g/L	Phytotechnology Labs
Casaminoacids	100 mg/L	Fisher Scientific
Sucrose	50 g/L	Phytotechnology Labs
2,4-D (Prod. No. D-7299)	1 mL/L	Sigma
(of 1 mg/mL Stock)		

Adjust the pH of the solution to pH to 5.8 with 1N KOH/1N KCl, add Gelrite (Sigma) to 3 g/L, and autoclave. After cool-

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ing to 50° C., add 2 ml/1 of a 5 mg/ml stock solution of Silver Nitrate (Phytotechnology Labs). This recipe yields about 20 plates.

5 All publications and patent applications mentioned in the specification are indicative of the level of skill of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

10 Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

## SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 15

<210> SEQ ID NO 1

<211> LENGTH: 3819

<212> TYPE: DNA

<213> ORGANISM: Brevibacillus

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (1) ... (3819)

<400> SEQUENCE: 1

```

atg aat caa aat caa aat cag aat cag aat caa aat aaa aat gaa ctt      48
Met Asn Gln Asn Gln Asn Gln Asn Gln Asn Gln Asn Lys Asn Glu Leu
  1                               5                               10                               15

caa atc ata gaa cct tca agc gat tct ttt ctt tat agt cac aac aat      96
Gln Ile Ile Glu Pro Ser Ser Asp Ser Phe Leu Tyr Ser His Asn Asn
                20                25                30

tat ccg tat tcc act gat cca aat aca gta tta cac ggt agg aat tac      144
Tyr Pro Tyr Ser Thr Asp Pro Asn Thr Val Leu His Gly Arg Asn Tyr
                35                40                45

aaa gag tgg cta aac atg tgt aca ggt aca gac gat tca cga ggt ccc      192
Lys Glu Trp Leu Asn Met Cys Thr Gly Thr Asp Asp Ser Arg Gly Pro
  50                55                60

gaa gct gct tct act gca aga tca gct ata tcg gtt gcg att act ata      240
Glu Ala Ala Ser Thr Ala Arg Ser Ala Ile Ser Val Ala Ile Thr Ile
  65                70                75                80

agc acc aca att ctt ggc tta cta ggt gtt ccg ttt gca tct cag atc      288
Ser Thr Thr Ile Leu Gly Leu Leu Gly Val Pro Phe Ala Ser Gln Ile
                85                90                95

ggg gca ttt tat aac ttc gta ttg aat acg gta tgg cct cag gga aat      336
Gly Ala Phe Tyr Asn Phe Val Leu Asn Thr Val Trp Pro Gln Gly Asn
  100                105                110

aac caa tgg gaa gag ttc atg aga cat gta gaa aat ctc ata aac gaa      384
Asn Gln Trp Glu Glu Phe Met Arg His Val Glu Asn Leu Ile Asn Glu
  115                120                125

cga ata gct gat tat gca aga agt aag gca ctt gca gaa tta acg ggt      432
Arg Ile Ala Asp Tyr Ala Arg Ser Lys Ala Leu Ala Glu Leu Thr Gly
  130                135                140

tta ggt aat aac tta aat tta tat aga gag gct ttt gaa gat tgg aga      480
Leu Gly Asn Asn Leu Asn Leu Tyr Arg Glu Ala Phe Glu Asp Trp Arg
  145                150                155                160

cga aat cct act agt caa gaa gct aaa acc cgc gta ata gat aga ttc      528
Arg Asn Pro Thr Ser Gln Glu Ala Lys Thr Arg Val Ile Asp Arg Phe
                165                170                175

cgt ata cta gat ggc tta ttt gaa gca tat atg cca tca ttt gca gta      576
Arg Ile Leu Asp Gly Leu Phe Glu Ala Tyr Met Pro Ser Phe Ala Val
                180                185                190

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caa ggt ttt gaa gta caa tta tta aca gtg tat gca tcc gct gca aat Gln Gly Phe Glu Val Gln Leu Leu Thr Val Tyr Ala Ser Ala Ala Asn 195 200 205	624
atc cat tta ttt tta tta aga gat agc tct att tac ggt ttg gat tgg Ile His Leu Phe Leu Leu Arg Asp Ser Ser Ile Tyr Gly Leu Asp Trp 210 215 220	672
gga tta agt caa act aat gtt aac gaa aat tac aat cgc caa ata agg Gly Leu Ser Gln Thr Asn Val Asn Glu Asn Tyr Asn Arg Gln Ile Arg 225 230 235 240	720
cac acc gca acg tat gca aat cat tgt aca act tgg tat caa act ggt His Thr Ala Thr Tyr Ala Asn His Cys Thr Thr Trp Tyr Gln Thr Gly 245 250 255	768
tta caa aga ttg caa ggt acc aat gct acc agt tgg ggc gct tat aat Leu Gln Arg Leu Gln Gly Thr Asn Ala Thr Ser Trp Gly Ala Tyr Asn 260 265 270	816
aga ttt aga agg gaa atg acg tta aca gta tta gat att agt tca tta Arg Phe Arg Arg Glu Met Thr Leu Thr Val Leu Asp Ile Ser Ser Leu 275 280 285	864
ttt tca aat tat gat tat cgt agt tat cca aca gag gta agg gga gag Phe Ser Asn Tyr Asp Tyr Arg Ser Tyr Pro Thr Glu Val Arg Gly Glu 290 295 300	912
ctt acg aga gaa att tat acg gat cca gta ggc ttt ggc tgg cag aat Leu Thr Arg Glu Ile Tyr Thr Asp Pro Val Gly Phe Gly Trp Gln Asn 305 310 315 320	960
aat gca cca tca ttc gct gaa ata gaa aat cta gca att agg gca cca Asn Ala Pro Ser Phe Ala Glu Ile Glu Asn Leu Ala Ile Arg Ala Pro 325 330 335	1008
aga acc gtt act tgg tta aat tca aca aga att cat aca ggg acc ttg Arg Thr Val Thr Trp Leu Asn Ser Thr Arg Ile His Thr Gly Thr Leu 340 345 350	1056
cag ggc tgg agt ggt tct aac aga tat tgg gca gct cac atg caa aac Gln Gly Trp Ser Gly Ser Asn Arg Tyr Trp Ala Ala His Met Gln Asn 355 360 365	1104
ttt tca gaa acc aat tca gga aat ata aga ttt gac ggt cct ctc tat Phe Ser Glu Thr Asn Ser Gly Asn Ile Arg Phe Asp Gly Pro Leu Tyr 370 375 380	1152
ggg tgc acg gta ggt act att att cgt act gat aat tac gaa atg ggg Gly Ser Thr Val Gly Thr Ile Ile Arg Thr Asp Asn Tyr Glu Met Gly 385 390 395 400	1200
aac cga gat att tac acc att act tca gaa gct gtt ggc gcc ctt tgg Asn Arg Asp Ile Tyr Thr Ile Thr Ser Glu Ala Val Gly Ala Leu Trp 405 410 415	1248
cca cat ggt caa act gtg ttg gga gtc gct tgc gct aga ttt act tta Pro His Gly Gln Thr Val Leu Gly Val Ala Ser Ala Arg Phe Thr Leu 420 425 430	1296
aga cat ctt tcc aat aat ttt aca cag gtg ctg gtg tat gag aat cca Arg His Leu Ser Asn Asn Phe Thr Gln Val Leu Val Tyr Glu Asn Pro 435 440 445	1344
ata agt aat agt ttt aat aga tca act gta act agt gaa tta cct gga Ile Ser Asn Ser Phe Asn Arg Ser Thr Val Thr Ser Glu Leu Pro Gly 450 455 460	1392
gaa aac tca gat agg cca act gat agc gat tat agt cat aga cta acg Glu Asn Ser Asp Arg Pro Thr Asp Ser Tyr Ser His Arg Leu Thr 465 470 475 480	1440
tgt atc aca gct ttt cga gct gga aat aat ggt acg gtt cca gta ttt Cys Ile Thr Ala Phe Arg Ala Gly Asn Asn Gly Thr Val Pro Val Phe 485 490 495	1488
ggc tgg aca tct aga act gtt aat cgc gac aat ata att gag caa aac Gly Trp Thr Ser Arg Thr Val Asn Arg Asp Asn Ile Ile Glu Gln Asn	1536

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500	505	510	
aaa att aca caa ttc cca ggt gtt aag tca cac act ctc aac aat tgt Lys Ile Thr Gln Phe Pro Gly Val Lys Ser His Thr Leu Asn Asn Cys 515 520 525			1584
caa gta gtt aga ggg act gga ttt act gga gga gac tgg ttg aga cca Gln Val Val Arg Gly Thr Gly Phe Thr Gly Gly Asp Trp Leu Arg Pro 530 535 540			1632
aat aat aat ggt aca ttt aga cta act att act tca ttc tcg agc caa Asn Asn Asn Gly Thr Phe Arg Leu Thr Ile Thr Ser Phe Ser Ser Gln 545 550 555 560			1680
tct tac cga atc cgc tta cgt tat gct act tca gta ggg aat act tct Ser Tyr Arg Ile Arg Leu Arg Tyr Ala Thr Ser Val Gly Asn Thr Ser 565 570 575			1728
tta gtt ata tct tct tct gat gca ggt att tct tcc aca aca att ccg Leu Val Ile Ser Ser Ser Asp Ala Gly Ile Ser Ser Thr Thr Ile Pro 580 585 590			1776
ctt act tca aca ata aca tca ctg ccc caa act gta cca tac cag gct Leu Thr Ser Thr Ile Thr Ser Leu Pro Gln Thr Val Pro Tyr Gln Ala 595 600 605			1824
ttt agg gtt gta gat tta cct att act ttt aca aca cct act acc caa Phe Arg Val Val Asp Leu Pro Ile Thr Phe Thr Thr Pro Thr Thr Gln 610 615 620			1872
aga aat tat acg ttt gat ttc cgt ctc caa aat cca tct aac gca aat Arg Asn Tyr Thr Phe Asp Phe Arg Leu Gln Asn Pro Ser Asn Ala Asn 625 630 635 640			1920
gta ttc att gat aga ttt gaa ttt gtt cca att ggg ggt tct ttg tct Val Phe Ile Asp Arg Phe Glu Phe Val Pro Ile Gly Gly Ser Leu Ser 645 650 655			1968
gag tat gaa acc aaa cat cag cta gaa aaa gca agg aaa gcg gtg aac Glu Tyr Glu Thr Lys His Gln Leu Glu Lys Ala Arg Lys Ala Val Asn 660 665 670			2016
gat ttg ttt acc aat gaa tcg aaa aat gtg tta aaa aaa gaa act act Asp Leu Phe Thr Asn Glu Ser Lys Asn Val Leu Lys Lys Glu Thr Thr 675 680 685			2064
gat tat gac ata gat caa gca gca aac ttg gta gaa tgt ata tca gat Asp Tyr Asp Ile Asp Gln Ala Ala Asn Leu Val Glu Cys Ile Ser Asp 690 695 700			2112
gaa tgt gca aat gca aaa atg atc cta tta gat gaa gta aaa tat gcg Glu Cys Ala Asn Ala Lys Met Ile Leu Leu Asp Glu Val Lys Tyr Ala 705 710 715 720			2160
aaa caa ctc agc gaa gcc cgc aat cta ctt cta aat ggt aat ttt gaa Lys Gln Leu Ser Glu Ala Arg Asn Leu Leu Leu Asn Gly Asn Phe Glu 725 730 735			2208
tac caa gat aga gat ggg gag aat cca tgg aaa aca agt ccc aat gtt Tyr Gln Asp Arg Asp Gly Glu Asn Pro Trp Lys Thr Ser Pro Asn Val 740 745 750			2256
acc atc caa gag aat aat ccc att ttt aaa ggc cgt tat ctc agt atg Thr Ile Gln Glu Asn Asn Pro Ile Phe Lys Gly Arg Tyr Leu Ser Met 755 760 765			2304
tcg ggt gcg aac aat atc gag gta aca aat gat ata ttc ccc act tat Ser Gly Ala Asn Asn Ile Glu Val Thr Asn Asp Ile Phe Pro Thr Tyr 770 775 780			2352
gca tac caa aaa att gat gaa tcc aaa tta aaa ccc tat acg cgt tat Ala Tyr Gln Lys Ile Asp Glu Ser Lys Leu Lys Pro Tyr Thr Arg Tyr 785 790 795 800			2400
aaa gtt cga ggg ttt gtt gga aat agt aaa gat tta gag ttg ttg att Lys Val Arg Gly Phe Val Gly Asn Ser Lys Asp Leu Glu Leu Leu Ile 805 810 815			2448
aca cga tat aat gaa gaa gta gat gcg att tta aat gta gca aat gat			2496

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Thr	Arg	Tyr	Asn	Glu	Glu	Val	Asp	Ala	Ile	Leu	Asn	Val	Ala	Asn	Asp	
			820					825					830			
ata	cca	cat	gct	ccg	aca	cct	ttc	tgc	ggt	gga	ttt	gat	cga	tgc	aag	2544
Ile	Pro	His	Ala	Pro	Thr	Pro	Phe	Cys	Gly	Gly	Phe	Asp	Arg	Cys	Lys	
			835					840					845			
cca	cat	tct	tat	cct	cct	atg	aat	cca	gaa	tgt	cac	cat	gat	gta	ata	2592
Pro	His	Ser	Tyr	Pro	Pro	Met	Asn	Pro	Glu	Cys	His	His	Asp	Val	Ile	
			850				855						860			
aat	aac	att	gaa	ata	tcc	tct	cct	tgc	cat	cac	aat	aag	atg	gtt	gat	2640
Asn	Asn	Ile	Glu	Ile	Ser	Ser	Pro	Cys	His	His	Asn	Lys	Met	Val	Asp	
			865				870						875		880	
aac	gct	gat	aca	tct	tct	cgc	cat	agt	gaa	tta	ggt	aaa	aaa	cat	ggc	2688
Asn	Ala	Asp	Thr	Ser	Ser	Arg	His	Ser	Glu	Leu	Gly	Lys	Lys	His	Gly	
				885						890					895	
att	tgt	cat	gaa	tct	cat	cat	ttt	gaa	ttc	cat	att	gat	aca	gga	aaa	2736
Ile	Cys	His	Glu	Ser	His	His	Phe	Glu	Phe	His	Ile	Asp	Thr	Gly	Lys	
			900					905								
atc	gat	ttg	gtc	gaa	aat	ttg	gga	att	tgg	gtt	ata	ttt	aaa	ata	tgt	2784
Ile	Asp	Leu	Val	Glu	Asn	Leu	Gly	Ile	Trp	Val	Ile	Phe	Lys	Ile	Cys	
			915					920					925			
tcc	aca	gat	ggt	tac	gca	aca	tta	gat	aat	ttg	gaa	gtt	att	gaa	gag	2832
Ser	Thr	Asp	Gly	Tyr	Ala	Thr	Leu	Asp	Asn	Leu	Glu	Val	Ile	Glu	Glu	
			930				935					940				
ggt	cct	tta	gga	gcc	gaa	tca	tta	gaa	cgt	gtg	aaa	aga	aga	gaa	aag	2880
Gly	Pro	Leu	Gly	Ala	Glu	Ser	Leu	Glu	Arg	Val	Lys	Arg	Arg	Glu	Lys	
			945			950				955					960	
aaa	tgg	aaa	cat	cac	atg	gaa	cac	aag	tgt	tca	gaa	act	aaa	cat	gta	2928
Lys	Trp	Lys	His	Met	Glu	His	Lys	Cys	Ser	Glu	Thr	Lys	His	Val		
				965					970					975		
tac	cat	gct	gcg	aaa	caa	gcg	gtg	gtg	gcg	tta	ttc	aca	aac	act	caa	2976
Tyr	His	Ala	Ala	Lys	Gln	Ala	Val	Val	Ala	Leu	Phe	Thr	Asn	Thr	Gln	
			980					985						990		
tat	gat	aga	ttg	aag	ttc	gaa	aca	acc	ata	tcc	aat	att	cta	ttt	gct	3024
Tyr	Asp	Arg	Leu	Lys	Phe	Glu	Thr	Thr	Ile	Ser	Asn	Ile	Leu	Phe	Ala	
			995				1000						1005			
gat	tat	ctc	gtg	tcg	tca	att	ccg	ttt	gta	tat	aat	aaa	tgg	tta	cca	3072
Asp	Tyr	Leu	Val	Ser	Ser	Ile	Pro	Phe	Val	Tyr	Asn	Lys	Trp	Leu	Pro	
			1010				1015					1020				
gat	gtt	cca	ggt	atg	aat	tat	gat	atc	tat	aca	gaa	tta	aaa	aat	ctg	3120
Asp	Val	Pro	Gly	Met	Asn	Tyr	Asp	Ile	Tyr	Thr	Glu	Leu	Lys	Asn	Leu	
			1025			1030				1035					1040	
att	acg	gga	gct	ttc	aat	cta	tac	gat	caa	cga	aat	att	ata	aaa	aat	3168
Ile	Thr	Gly	Ala	Phe	Asn	Leu	Tyr	Asp	Gln	Arg	Asn	Ile	Ile	Lys	Asn	
				1045					1050					1055		
gga	gac	ttt	aat	aac	gga	ctc	atg	cat	tgg	cat	gcg	aca	cct	cat	gcg	3216
Gly	Asp	Phe	Asn	Asn	Gly	Leu	Met	His	Trp	His	Ala	Thr	Pro	His	Ala	
			1060					1065					1070			
aga	gta	gag	caa	ata	gat	aat	agg	tct	gtg	ctg	gtg	ctt	cca	aat	tat	3264
Arg	Val	Glu	Gln	Ile	Asp	Asn	Arg	Ser	Val	Leu	Val	Leu	Pro	Asn	Tyr	
			1075				1080					1085				
gct	gcc	aat	gtt	tca	caa	gag	gtt	tgt	tta	gaa	cac	aat	cgt	ggt	tat	3312
Ala	Ala	Asn	Val	Ser	Gln	Glu	Val	Cys	Leu	Glu	His	Asn	Arg	Gly	Tyr	
			1090				1095					1100				
gta	tta	cgt	gta	acg	gcg	aaa	aaa	gaa	ggt	cct	gga	att	gga	tat	gtt	3360
Val	Leu	Arg	Val	Thr	Ala	Lys	Lys	Glu	Gly	Pro	Gly	Ile	Gly	Tyr	Val	
			1105			1110				1115					1120	
acg	ttc	agt	gat	tgt	gca	aat	aat	ata	gaa	aaa	ctg	aca	ttt	act	tct	3408
Thr	Phe	Ser	Asp	Cys	Ala	Asn	Asn	Ile	Glu	Lys	Leu	Thr	Phe	Thr	Ser	
				1125					1130					1135		

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tgc gat tat ggt aca aac gaa gtg cca tat gag caa tct aat tat cct	3456
Cys Asp Tyr Gly Thr Asn Glu Val Pro Tyr Glu Gln Ser Asn Tyr Pro	
1140 1145 1150	
aca gac gga gtt tca tac gga caa cat ggt tgt aat ata gac aga gta	3504
Thr Asp Gly Val Ser Tyr Gly Gln His Gly Cys Asn Ile Asp Arg Val	
1155 1160 1165	
ccg tac gaa caa tct ggt tat cct aca gac gga gta tgc tac gaa caa	3552
Pro Tyr Glu Gln Ser Gly Tyr Pro Thr Asp Gly Val Ser Tyr Glu Gln	
1170 1175 1180	
tct ggt tat cgt aca gac gga gta ctg tac gaa caa tct ggt tat cgt	3600
Ser Gly Tyr Arg Thr Asp Gly Val Leu Tyr Glu Gln Ser Gly Tyr Arg	
1185 1190 1195 1200	
aca gac gga gta cca tgc gaa caa cat ggt tgt cat aca gac gga gta	3648
Thr Asp Gly Val Pro Cys Glu Gln His Gly Cys His Thr Asp Gly Val	
1205 1210 1215	
cca tac aaa caa cat ggt tgt cat aca gac aga tca aga gat gaa caa	3696
Pro Tyr Lys Gln His Gly Cys His Thr Asp Arg Ser Arg Asp Glu Gln	
1220 1225 1230	
ctt ggt tat gtg aca aaa acg att gat gta ttc cct gat aca gat aaa	3744
Leu Gly Tyr Val Thr Lys Thr Ile Asp Val Phe Pro Asp Thr Asp Lys	
1235 1240 1245	
gta cgt atc gac att gga gaa acc gaa ggt acc ttt aaa gta gaa agt	3792
Val Arg Ile Asp Ile Gly Glu Thr Glu Gly Thr Phe Lys Val Glu Ser	
1250 1255 1260	
gtg gaa ctg att tgt atg gaa gag taa	3819
Val Glu Leu Ile Cys Met Glu Glu	
1265 1270	

&lt;210&gt; SEQ ID NO 2

&lt;211&gt; LENGTH: 1272

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Brevibacillus

&lt;400&gt; SEQUENCE: 2

Met Asn Gln Asn Gln Asn Gln Asn Gln Asn Lys Asn Glu Leu	
1 5 10 15	
Gln Ile Ile Glu Pro Ser Ser Asp Ser Phe Leu Tyr Ser His Asn Asn	
20 25 30	
Tyr Pro Tyr Ser Thr Asp Pro Asn Thr Val Leu His Gly Arg Asn Tyr	
35 40 45	
Lys Glu Trp Leu Asn Met Cys Thr Gly Thr Asp Asp Ser Arg Gly Pro	
50 55 60	
Glu Ala Ala Ser Thr Ala Arg Ser Ala Ile Ser Val Ala Ile Thr Ile	
65 70 75 80	
Ser Thr Thr Ile Leu Gly Leu Leu Gly Val Pro Phe Ala Ser Gln Ile	
85 90 95	
Gly Ala Phe Tyr Asn Phe Val Leu Asn Thr Val Trp Pro Gln Gly Asn	
100 105 110	
Asn Gln Trp Glu Glu Phe Met Arg His Val Glu Asn Leu Ile Asn Glu	
115 120 125	
Arg Ile Ala Asp Tyr Ala Arg Ser Lys Ala Leu Ala Glu Leu Thr Gly	
130 135 140	
Leu Gly Asn Asn Leu Asn Leu Tyr Arg Glu Ala Phe Glu Asp Trp Arg	
145 150 155 160	
Arg Asn Pro Thr Ser Gln Glu Ala Lys Thr Arg Val Ile Asp Arg Phe	
165 170 175	
Arg Ile Leu Asp Gly Leu Phe Glu Ala Tyr Met Pro Ser Phe Ala Val	
180 185 190	

Gln	Gly	Phe	Glu	Val	Gln	Leu	Leu	Thr	Val	Tyr	Ala	Ser	Ala	Ala	Asn
Ile	His	Leu	Phe	Leu	Leu	Arg	Asp	Ser	Ser	Ile	Tyr	Gly	Leu	Asp	Trp
Gly	Leu	Ser	Gln	Thr	Asn	Val	Asn	Glu	Asn	Tyr	Asn	Arg	Gln	Ile	Arg
His	Thr	Ala	Thr	Tyr	Ala	Asn	His	Cys	Thr	Thr	Trp	Tyr	Gln	Thr	Gly
Leu	Gln	Arg	Leu	Gln	Gly	Thr	Asn	Ala	Thr	Ser	Trp	Gly	Ala	Tyr	Asn
Arg	Phe	Arg	Arg	Glu	Met	Thr	Leu	Thr	Val	Leu	Asp	Ile	Ser	Ser	Leu
Phe	Ser	Asn	Tyr	Asp	Tyr	Arg	Ser	Tyr	Pro	Thr	Glu	Val	Arg	Gly	Glu
Leu	Thr	Arg	Glu	Ile	Tyr	Thr	Asp	Pro	Val	Gly	Phe	Gly	Trp	Gln	Asn
Asn	Ala	Pro	Ser	Phe	Ala	Glu	Ile	Glu	Asn	Leu	Ala	Ile	Arg	Ala	Pro
Arg	Thr	Val	Thr	Trp	Leu	Asn	Ser	Thr	Arg	Ile	His	Thr	Gly	Thr	Leu
Gln	Gly	Trp	Ser	Gly	Ser	Asn	Arg	Tyr	Trp	Ala	Ala	His	Met	Gln	Asn
Phe	Ser	Glu	Thr	Asn	Ser	Gly	Asn	Ile	Arg	Phe	Asp	Gly	Pro	Leu	Tyr
Gly	Ser	Thr	Val	Gly	Thr	Ile	Ile	Arg	Thr	Asp	Asn	Tyr	Glu	Met	Gly
Asn	Arg	Asp	Ile	Tyr	Thr	Ile	Thr	Ser	Glu	Ala	Val	Gly	Ala	Leu	Trp
Pro	His	Gly	Gln	Thr	Val	Leu	Gly	Val	Ala	Ser	Ala	Arg	Phe	Thr	Leu
Arg	His	Leu	Ser	Asn	Asn	Phe	Thr	Gln	Val	Leu	Val	Tyr	Glu	Asn	Pro
Ile	Ser	Asn	Ser	Phe	Asn	Arg	Ser	Thr	Val	Thr	Ser	Glu	Leu	Pro	Gly
Glu	Asn	Ser	Asp	Arg	Pro	Thr	Asp	Ser	Asp	Tyr	Ser	His	Arg	Leu	Thr
Cys	Ile	Thr	Ala	Phe	Arg	Ala	Gly	Asn	Asn	Gly	Thr	Val	Pro	Val	Phe
Gly	Trp	Thr	Ser	Arg	Thr	Val	Asn	Arg	Asp	Asn	Ile	Ile	Glu	Gln	Asn
Lys	Ile	Thr	Gln	Phe	Pro	Gly	Val	Lys	Ser	His	Thr	Leu	Asn	Asn	Cys
Gln	Val	Val	Arg	Gly	Thr	Gly	Phe	Thr	Gly	Gly	Asp	Trp	Leu	Arg	Pro
Asn	Asn	Asn	Gly	Thr	Phe	Arg	Leu	Thr	Ile	Thr	Ser	Phe	Ser	Ser	Gln
Ser	Tyr	Arg	Ile	Arg	Leu	Arg	Tyr	Ala	Thr	Ser	Val	Gly	Asn	Thr	Ser
Leu	Val	Ile	Ser	Ser	Ser	Asp	Ala	Gly	Ile	Ser	Ser	Thr	Thr	Ile	Pro
Leu	Thr	Ser	Thr	Ile	Thr	Ser	Leu	Pro	Gln	Thr	Val	Pro	Tyr	Gln	Ala

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Phe	Arg	Val	Val	Asp	Leu	Pro	Ile	Thr	Phe	Thr	Thr	Pro	Thr	Thr	Gln
610						615					620				
Arg	Asn	Tyr	Thr	Phe	Asp	Phe	Arg	Leu	Gln	Asn	Pro	Ser	Asn	Ala	Asn
625					630					635					640
Val	Phe	Ile	Asp	Arg	Phe	Glu	Phe	Val	Pro	Ile	Gly	Gly	Ser	Leu	Ser
				645					650					655	
Glu	Tyr	Glu	Thr	Lys	His	Gln	Leu	Glu	Lys	Ala	Arg	Lys	Ala	Val	Asn
			660					665					670		
Asp	Leu	Phe	Thr	Asn	Glu	Ser	Lys	Asn	Val	Leu	Lys	Lys	Glu	Thr	Thr
		675					680					685			
Asp	Tyr	Asp	Ile	Asp	Gln	Ala	Ala	Asn	Leu	Val	Glu	Cys	Ile	Ser	Asp
	690				695						700				
Glu	Cys	Ala	Asn	Ala	Lys	Met	Ile	Leu	Leu	Asp	Glu	Val	Lys	Tyr	Ala
705					710					715					720
Lys	Gln	Leu	Ser	Glu	Ala	Arg	Asn	Leu	Leu	Leu	Asn	Gly	Asn	Phe	Glu
			725					730						735	
Tyr	Gln	Asp	Arg	Asp	Gly	Glu	Asn	Pro	Trp	Lys	Thr	Ser	Pro	Asn	Val
			740					745						750	
Thr	Ile	Gln	Glu	Asn	Asn	Pro	Ile	Phe	Lys	Gly	Arg	Tyr	Leu	Ser	Met
		755					760					765			
Ser	Gly	Ala	Asn	Asn	Ile	Glu	Val	Thr	Asn	Asp	Ile	Phe	Pro	Thr	Tyr
	770					775					780				
Ala	Tyr	Gln	Lys	Ile	Asp	Glu	Ser	Lys	Leu	Lys	Pro	Tyr	Thr	Arg	Tyr
785					790					795					800
Lys	Val	Arg	Gly	Phe	Val	Gly	Asn	Ser	Lys	Asp	Leu	Glu	Leu	Leu	Ile
			805					810						815	
Thr	Arg	Tyr	Asn	Glu	Glu	Val	Asp	Ala	Ile	Leu	Asn	Val	Ala	Asn	Asp
			820					825						830	
Ile	Pro	His	Ala	Pro	Thr	Pro	Phe	Cys	Gly	Gly	Phe	Asp	Arg	Cys	Lys
		835					840					845			
Pro	His	Ser	Tyr	Pro	Pro	Met	Asn	Pro	Glu	Cys	His	His	Asp	Val	Ile
	850					855					860				
Asn	Asn	Ile	Glu	Ile	Ser	Ser	Pro	Cys	His	His	Asn	Lys	Met	Val	Asp
865					870					875					880
Asn	Ala	Asp	Thr	Ser	Ser	Arg	His	Ser	Glu	Leu	Gly	Lys	Lys	His	Gly
			885						890					895	
Ile	Cys	His	Glu	Ser	His	His	Phe	Glu	Phe	His	Ile	Asp	Thr	Gly	Lys
		900						905					910		
Ile	Asp	Leu	Val	Glu	Asn	Leu	Gly	Ile	Trp	Val	Ile	Phe	Lys	Ile	Cys
	915						920					925			
Ser	Thr	Asp	Gly	Tyr	Ala	Thr	Leu	Asp	Asn	Leu	Glu	Val	Ile	Glu	Glu
	930					935					940				
Gly	Pro	Leu	Gly	Ala	Glu	Ser	Leu	Glu	Arg	Val	Lys	Arg	Arg	Glu	Lys
945					950					955					960
Lys	Trp	Lys	His	His	Met	Glu	His	Lys	Cys	Ser	Glu	Thr	Lys	His	Val
			965					970						975	
Tyr	His	Ala	Ala	Lys	Gln	Ala	Val	Val	Ala	Leu	Phe	Thr	Asn	Thr	Gln
			980					985					990		
Tyr	Asp	Arg	Leu	Lys	Phe	Glu	Thr	Thr	Ile	Ser	Asn	Ile	Leu	Phe	Ala
	995							1000					1005		
Asp	Tyr	Leu	Val	Ser	Ser	Ile	Pro	Phe	Val	Tyr	Asn	Lys	Trp	Leu	Pro
	1010					1015					1020				
Asp	Val	Pro	Gly	Met	Asn	Tyr	Asp	Ile	Tyr	Thr	Glu	Leu	Lys	Asn	Leu

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1025	1030	1035	1040
Ile Thr Gly Ala Phe Asn Leu Tyr Asp Gln Arg Asn Ile Ile Lys Asn			
	1045	1050	1055
Gly Asp Phe Asn Asn Gly Leu Met His Trp His Ala Thr Pro His Ala			
	1060	1065	1070
Arg Val Glu Gln Ile Asp Asn Arg Ser Val Leu Val Leu Pro Asn Tyr			
	1075	1080	1085
Ala Ala Asn Val Ser Gln Glu Val Cys Leu Glu His Asn Arg Gly Tyr			
	1090	1095	1100
Val Leu Arg Val Thr Ala Lys Lys Glu Gly Pro Gly Ile Gly Tyr Val			
	1105	1110	1115
Thr Phe Ser Asp Cys Ala Asn Asn Ile Glu Lys Leu Thr Phe Thr Ser			
	1125	1130	1135
Cys Asp Tyr Gly Thr Asn Glu Val Pro Tyr Glu Gln Ser Asn Tyr Pro			
	1140	1145	1150
Thr Asp Gly Val Ser Tyr Gly Gln His Gly Cys Asn Ile Asp Arg Val			
	1155	1160	1165
Pro Tyr Glu Gln Ser Gly Tyr Pro Thr Asp Gly Val Ser Tyr Glu Gln			
	1170	1175	1180
Ser Gly Tyr Arg Thr Asp Gly Val Leu Tyr Glu Gln Ser Gly Tyr Arg			
	1185	1190	1195
Thr Asp Gly Val Pro Cys Glu Gln His Gly Cys His Thr Asp Gly Val			
	1205	1210	1215
Pro Tyr Lys Gln His Gly Cys His Thr Asp Arg Ser Arg Asp Glu Gln			
	1220	1225	1230
Leu Gly Tyr Val Thr Lys Thr Ile Asp Val Phe Pro Asp Thr Asp Lys			
	1235	1240	1245
Val Arg Ile Asp Ile Gly Glu Thr Glu Gly Thr Phe Lys Val Glu Ser			
	1250	1255	1260
Val Glu Leu Ile Cys Met Glu Glu			
	1265	1270	

&lt;210&gt; SEQ ID NO 3

&lt;211&gt; LENGTH: 1953

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Brevibacillus

&lt;220&gt; FEATURE:

&lt;221&gt; NAME/KEY: CDS

&lt;222&gt; LOCATION: (1)...(1953)

&lt;400&gt; SEQUENCE: 3

atg aat caa aat caa aat cag aat cag aat caa aat aaa aat gaa ctt	48
Met Asn Gln Asn Gln Asn Gln Asn Gln Asn Lys Asn Glu Leu	
1 5 10 15	
caa atc ata gaa cct tca agc gat tct ttt ctt tat agt cac aac aat	96
Gln Ile Ile Glu Pro Ser Ser Asp Ser Phe Leu Tyr Ser His Asn Asn	
20 25 30	
tat ccg tat tcc act gat cca aat aca gta tta cac ggt agg aat tac	144
Tyr Pro Tyr Ser Thr Asp Pro Asn Thr Val Leu His Gly Arg Asn Tyr	
35 40 45	
aaa gag tgg cta aac atg tgt aca ggt aca gac gat tca cga ggt ccc	192
Lys Glu Trp Leu Asn Met Cys Thr Gly Thr Asp Asp Ser Arg Gly Pro	
50 55 60	
gaa gct gct tct act gca aga tca gct ata tcg gtt gcg att act ata	240
Glu Ala Ala Ser Thr Ala Arg Ser Ala Ile Ser Val Ala Ile Thr Ile	
65 70 75 80	
agc acc aca att ctt ggc tta cta ggt gtt ccg ttt gca tct cag atc	288



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Ser	Thr	Thr	Ile	Leu	Gly	Leu	Leu	Gly	Val	Pro	Phe	Ala	Ser	Gln	Ile	
				85					90					95		
ggg	gca	ttt	tat	aac	ttc	gta	ttg	aat	acg	gta	tgg	cct	cag	gga	aat	336
Gly	Ala	Phe	Tyr	Asn	Phe	Val	Leu	Asn	Thr	Val	Trp	Pro	Gln	Gly	Asn	
			100					105					110			
aac	caa	tgg	gaa	gag	ttc	atg	aga	cat	gta	gaa	aat	ctc	ata	aac	gaa	384
Asn	Gln	Trp	Glu	Glu	Phe	Met	Arg	His	Val	Glu	Asn	Leu	Ile	Asn	Glu	
			115				120					125				
cga	ata	gct	gat	tat	gca	aga	agt	aag	gca	ctt	gca	gaa	tta	acg	ggt	432
Arg	Ile	Ala	Asp	Tyr	Ala	Arg	Ser	Lys	Ala	Leu	Ala	Glu	Leu	Thr	Gly	
	130					135					140					
tta	ggt	aat	aac	tta	aat	tta	tat	aga	gag	gct	ttt	gaa	gat	tgg	aga	480
Leu	Gly	Asn	Asn	Leu	Asn	Leu	Tyr	Arg	Glu	Ala	Phe	Glu	Asp	Trp	Arg	
	145				150					155					160	
cga	aat	cct	act	agt	caa	gaa	gct	aaa	acc	cgc	gta	ata	gat	aga	ttc	528
Arg	Asn	Pro	Thr	Ser	Gln	Glu	Ala	Lys	Thr	Arg	Val	Ile	Asp	Arg	Phe	
			165					170					175			
cgt	ata	cta	gat	ggc	tta	ttt	gaa	gca	tat	atg	cca	tca	ttt	gca	gta	576
Arg	Ile	Leu	Asp	Gly	Leu	Phe	Glu	Ala	Tyr	Met	Pro	Ser	Phe	Ala	Val	
			180					185					190			
caa	ggt	ttt	gaa	gta	caa	tta	tta	aca	gtg	tat	gca	tcc	gct	gca	aat	624
Gln	Gly	Phe	Glu	Val	Gln	Leu	Leu	Thr	Val	Tyr	Ala	Ser	Ala	Ala	Asn	
		195					200					205				
atc	cat	tta	ttt	tta	tta	aga	gat	agc	tct	att	tac	ggt	ttg	gat	tgg	672
Ile	His	Leu	Phe	Leu	Leu	Arg	Asp	Ser	Ser	Ile	Tyr	Gly	Leu	Asp	Trp	
	210					215					220					
gga	tta	agt	caa	act	aat	gtt	aac	gaa	aat	tac	aat	cgc	caa	ata	agg	720
Gly	Leu	Ser	Gln	Thr	Asn	Val	Asn	Glu	Asn	Tyr	Asn	Arg	Gln	Ile	Arg	
	225				230					235				240		
cac	acc	gca	acg	tat	gca	aat	cat	tgt	aca	act	tgg	tat	caa	act	ggt	768
His	Thr	Ala	Thr	Tyr	Ala	Asn	His	Cys	Thr	Thr	Trp	Tyr	Gln	Thr	Gly	
			245					250					255			
tta	caa	aga	ttg	caa	ggt	acc	aat	gct	acc	agt	tgg	ggc	gct	tat	aat	816
Leu	Gln	Arg	Leu	Gln	Gly	Thr	Asn	Ala	Thr	Ser	Trp	Gly	Ala	Tyr	Asn	
			260				265						270			
aga	ttt	aga	agg	gaa	atg	acg	tta	aca	gta	tta	gat	att	agt	tca	tta	864
Arg	Phe	Arg	Arg	Glu	Met	Thr	Leu	Thr	Val	Leu	Asp	Ile	Ser	Ser	Leu	
		275					280					285				
ttt	tca	aat	tat	gat	tat	cgt	agt	tat	cca	aca	gag	gta	agg	gga	gag	912
Phe	Ser	Asn	Tyr	Asp	Tyr	Arg	Ser	Tyr	Pro	Thr	Glu	Val	Arg	Gly	Glu	
	290					295					300					
ctt	acg	aga	gaa	att	tat	acg	gat	cca	gta	ggc	ttt	ggc	tgg	cag	aat	960
Leu	Thr	Arg	Glu	Ile	Tyr	Thr	Asp	Pro	Val	Gly	Phe	Gly	Trp	Gln	Asn	
	305				310					315				320		
aat	gca	cca	tca	ttc	gct	gaa	ata	gaa	aat	cta	gca	att	agg	gca	cca	1008
Asn	Ala	Pro	Ser	Phe	Ala	Glu	Ile	Glu	Asn	Leu	Ala	Ile	Arg	Ala	Pro	
			325						330				335			
aga	acc	gtt	act	tgg	tta	aat	tca	aca	aga	att	cat	aca	ggg	acc	ttg	1056
Arg	Thr	Val	Thr	Trp	Leu	Asn	Ser	Thr	Arg	Ile	His	Thr	Gly	Thr	Leu	
			340					345					350			
cag	ggc	tgg	agt	ggt	tct	aac	aga	tat	tgg	gca	gct	cac	atg	caa	aac	1104
Gln	Gly	Trp	Ser	Gly	Ser	Asn	Arg	Tyr	Trp	Ala	Ala	His	Met	Gln	Asn	
		355					360					365				
ttt	tca	gaa	acc	aat	tca	gga	aat	ata	aga	ttt	gac	ggt	cct	ctc	tat	1152
Phe	Ser	Glu	Thr	Asn	Ser	Gly	Asn	Ile	Arg	Phe	Asp	Gly	Pro	Leu	Tyr	
	370					375					380					
ggg	tcg	acg	gta	ggt	act	att	att	cgt	act	gat	aat	tac	gaa	atg	ggg	1200
Gly	Ser	Thr	Val	Gly	Thr	Ile	Ile	Arg	Thr	Asp	Asn	Tyr	Glu	Met	Gly	
	385					390				395					400	

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aac cga gat att tac acc att act tca gaa gct gtt ggc gcc ctt tgg	1248
Asn Arg Asp Ile Tyr Thr Ile Thr Ser Glu Ala Val Gly Ala Leu Trp	
405 410 415	
cca cat ggt caa act gtg ttg gga gtc gct tcg gct aga ttt act tta	1296
Pro His Gly Gln Thr Val Leu Gly Val Ala Ser Ala Arg Phe Thr Leu	
420 425 430	
aga cat ctt tcc aat aat ttt aca cag gtg ctg gtg tat gag aat cca	1344
Arg His Leu Ser Asn Asn Phe Thr Gln Val Leu Val Tyr Glu Asn Pro	
435 440 445	
ata agt aat agt ttt aat aga tca act gta act agt gaa tta cct gga	1392
Ile Ser Asn Ser Phe Asn Arg Ser Thr Val Thr Ser Glu Leu Pro Gly	
450 455 460	
gaa aac tca gat agg cca act gat agc gat tat agt cat aga cta acg	1440
Glu Asn Ser Asp Arg Pro Thr Asp Ser Asp Tyr Ser His Arg Leu Thr	
465 470 475 480	
tgt atc aca gct ttt cga gct gga aat aat ggt acg gtt cca gta ttt	1488
Cys Ile Thr Ala Phe Arg Ala Gly Asn Asn Gly Thr Val Pro Val Phe	
485 490 495	
ggc tgg aca tct aga act gtt aat cgc gac aat ata att gag caa aac	1536
Gly Trp Thr Ser Arg Thr Val Asn Arg Asp Asn Ile Ile Glu Gln Asn	
500 505 510	
aaa att aca caa ttc cca ggt gtt aag tca cac act ctc aac aat tgt	1584
Lys Ile Thr Gln Phe Pro Gly Val Lys Ser His Thr Leu Asn Asn Cys	
515 520 525	
caa gta gtt aga ggg act gga ttt act gga gga gac tgg ttg aga cca	1632
Gln Val Val Arg Gly Thr Gly Phe Thr Gly Gly Asp Trp Leu Arg Pro	
530 535 540	
aat aat aat ggt aca ttt aga cta act att act tca ttc tcg agc caa	1680
Asn Asn Asn Gly Thr Phe Arg Leu Thr Ile Thr Ser Phe Ser Ser Gln	
545 550 555 560	
tct tac cga atc cgc tta cgt tat gct act tca gta ggg aat act tct	1728
Ser Tyr Arg Ile Arg Leu Arg Tyr Ala Thr Ser Val Gly Asn Thr Ser	
565 570 575	
tta gtt ata tct tct tct gat gca ggt att tct tcc aca aca att ccg	1776
Leu Val Ile Ser Ser Asp Ala Gly Ile Ser Ser Thr Thr Ile Pro	
580 585 590	
ctt act tca aca ata aca tca ctg ccc caa act gta cca tac cag gct	1824
Leu Thr Ser Thr Ile Thr Ser Leu Pro Gln Thr Val Pro Tyr Gln Ala	
595 600 605	
ttt agg gtt gta gat tta cct att act ttt aca aca cct act acc caa	1872
Phe Arg Val Val Asp Leu Pro Ile Thr Phe Thr Thr Pro Thr Thr Gln	
610 615 620	
aga aat tat acg ttt gat ttc cgt ctc caa aat cca tct aac gca aat	1920
Arg Asn Tyr Thr Phe Asp Phe Arg Leu Gln Asn Pro Ser Asn Ala Asn	
625 630 635 640	
gta ttc att gat aga ttt gaa ttt gtt cca att	1953
Val Phe Ile Asp Arg Phe Glu Phe Val Pro Ile	
645 650	

&lt;210&gt; SEQ ID NO 4

&lt;211&gt; LENGTH: 651

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Brevibacillus

&lt;400&gt; SEQUENCE: 4

Met Asn Gln Asn Gln Asn Gln Asn Gln Asn Gln Asn Lys Asn Glu Leu
1 5 10 15

Gln Ile Ile Glu Pro Ser Ser Asp Ser Phe Leu Tyr Ser His Asn Asn
20 25 30

Tyr Pro Tyr Ser Thr Asp Pro Asn Thr Val Leu His Gly Arg Asn Tyr
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35	40	45
Lys Glu Trp Leu Asn Met	Cys Thr Gly Thr Asp	Asp Ser Arg Gly Pro
50	55	60
Glu Ala Ala Ser Thr Ala	Arg Ser Ala Ile Ser	Val Ala Ile Thr Ile
65	70	75
Ser Thr Thr Ile Leu Gly	Leu Leu Gly Val Pro	Phe Ala Ser Gln Ile
85	90	95
Gly Ala Phe Tyr Asn Phe	Val Leu Asn Thr Val	Trp Pro Gln Gly Asn
100	105	110
Asn Gln Trp Glu Glu Phe	Met Arg His Val Glu	Asn Leu Ile Asn Glu
115	120	125
Arg Ile Ala Asp Tyr Ala	Arg Ser Lys Ala Leu	Ala Glu Leu Thr Gly
130	135	140
Leu Gly Asn Asn Leu Asn	Leu Tyr Arg Glu Ala	Phe Glu Asp Trp Arg
145	150	155
Arg Asn Pro Thr Ser Gln	Glu Ala Lys Thr Arg	Val Ile Asp Arg Phe
165	170	175
Arg Ile Leu Asp Gly Leu	Phe Glu Ala Tyr Met	Pro Ser Phe Ala Val
180	185	190
Gln Gly Phe Glu Val Gln	Leu Leu Thr Val Tyr	Ala Ser Ala Ala Asn
195	200	205
Ile His Leu Phe Leu Leu	Arg Asp Ser Ser Ile	Tyr Gly Leu Asp Trp
210	215	220
Gly Leu Ser Gln Thr Asn	Val Asn Glu Asn Tyr	Asn Arg Gln Ile Arg
225	230	235
His Thr Ala Thr Tyr Ala	Asn His Cys Thr Thr	Trp Tyr Gln Thr Gly
245	250	255
Leu Gln Arg Leu Gln Gly	Thr Asn Ala Thr Ser	Trp Gly Ala Tyr Asn
260	265	270
Arg Phe Arg Arg Glu Met	Thr Leu Thr Val Leu	Asp Ile Ser Ser Leu
275	280	285
Phe Ser Asn Tyr Asp Tyr	Arg Ser Tyr Pro Thr	Glu Val Arg Gly Glu
290	295	300
Leu Thr Arg Glu Ile Tyr	Thr Asp Pro Val Gly	Phe Gly Trp Gln Asn
305	310	315
Asn Ala Pro Ser Phe Ala	Glu Ile Glu Asn Leu	Ala Ile Arg Ala Pro
325	330	335
Arg Thr Val Thr Trp Leu	Asn Ser Thr Arg Ile	His Thr Gly Thr Leu
340	345	350
Gln Gly Trp Ser Gly Ser	Asn Arg Tyr Trp Ala	Ala His Met Gln Asn
355	360	365
Phe Ser Glu Thr Asn Ser	Gly Asn Ile Arg Phe	Asp Gly Pro Leu Tyr
370	375	380
Gly Ser Thr Val Gly Thr	Ile Ile Arg Thr Asp	Asn Tyr Glu Met Gly
385	390	395
Asn Arg Asp Ile Tyr Thr	Ile Thr Ser Glu Ala	Val Gly Ala Leu Trp
405	410	415
Pro His Gly Gln Thr Val	Leu Gly Val Ala Ser	Ala Arg Phe Thr Leu
420	425	430
Arg His Leu Ser Asn Asn	Phe Thr Gln Val Leu	Val Tyr Glu Asn Pro
435	440	445
Ile Ser Asn Ser Phe Asn	Arg Ser Thr Val Thr	Ser Glu Leu Pro Gly
450	455	460

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Glu Asn Ser Asp Arg Pro Thr Asp Ser Asp Tyr Ser His Arg Leu Thr  
 465 470 475 480  
 Cys Ile Thr Ala Phe Arg Ala Gly Asn Asn Gly Thr Val Pro Val Phe  
 485 490 495  
 Gly Trp Thr Ser Arg Thr Val Asn Arg Asp Asn Ile Ile Glu Gln Asn  
 500 505 510  
 Lys Ile Thr Gln Phe Pro Gly Val Lys Ser His Thr Leu Asn Asn Cys  
 515 520 525  
 Gln Val Val Arg Gly Thr Gly Phe Thr Gly Gly Asp Trp Leu Arg Pro  
 530 535 540  
 Asn Asn Asn Gly Thr Phe Arg Leu Thr Ile Thr Ser Phe Ser Ser Gln  
 545 550 555 560  
 Ser Tyr Arg Ile Arg Leu Arg Tyr Ala Thr Ser Val Gly Asn Thr Ser  
 565 570 575  
 Leu Val Ile Ser Ser Ser Asp Ala Gly Ile Ser Ser Thr Thr Ile Pro  
 580 585 590  
 Leu Thr Ser Thr Ile Thr Ser Leu Pro Gln Thr Val Pro Tyr Gln Ala  
 595 600 605  
 Phe Arg Val Val Asp Leu Pro Ile Thr Phe Thr Thr Pro Thr Thr Gln  
 610 615 620  
 Arg Asn Tyr Thr Phe Asp Phe Arg Leu Gln Asn Pro Ser Asn Ala Asn  
 625 630 635 640  
 Val Phe Ile Asp Arg Phe Glu Phe Val Pro Ile  
 645 650

&lt;210&gt; SEQ ID NO 5

&lt;211&gt; LENGTH: 4150

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Brevibacillus

&lt;400&gt; SEQUENCE: 5

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ttttactgac ccacattcca aagaaatctc actttgttgt gatgggagcg tatgtgtaga      60
tcatgtactc aagtgtagtg tcggatgttt accagacggt catttagatt gtcaatatgt      120
aactgtatgt gatctacaaa tgacctgtg tcatgagaga gcttgtcaat ttgttaagat      180
tactggagaa tttcaatttt cctccattta aataagtacg aattttgttg ttaaacaccc      240
attattaata aatgggtaac aaataggagg gtaaatatga atcaaaatca aaatcagaat      300
cagaatcaaa ataaaaatga acttcaaatc atagaacctt caagcgattc ttttctttat      360
agtcacaaca attatccgta ttccactgat ccaaatacag tattacacgg taggaattac      420
aaagagtggc taaacatgtg tacaggtaca gacgattcac gaggtcccga agctgcttct      480
actgcaagat cagctatatc ggttgcgatt actataagca ccacaattct tggcttacta      540
ggtgttccgt ttgcatctca gatcggggca ttttataact tcgtattgaa tacggtatgg      600
cctcagggaa ataaccaatg ggaagagtgc atgagacatg tagaaaatct cataaacgaa      660
cgaatagctg attatgcaag aagtaaggca cttgcagaat taacgggttt aggtataaac      720
ttaaatatat atagagaggc ttttgaagat tggagacgaa atcctactag tcaagaagct      780
aaaacccgcg taatagatag attccgtata ctagatggct tatttgaagc atatatgcca      840
tcatttgcag tacaagggtt tgaagtacaa ttattaacag tgtatgcac cgctgcaaat      900
atccatttat ttttattaag agatagctct atttacggtt tggattgggg attaagtcaa      960
actaatgtta acgaaaatta caatcgccaa ataaggcaca ccgcaacgta tgcaaatcat     1020

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tgtacaactt	ggtatcaaac	tggtttacaa	agattgcaag	gtaccaatgc	taccagttgg	1080
ggcgcttata	atagatttag	aagggaaatg	acgttaacag	tattagatat	tagttcatta	1140
ttttcaaatt	atgattatcg	tagttatcca	acagaggtaa	ggggagagct	tacgagagaa	1200
atttatacgg	atccagtagg	ctttggettg	cagaataatg	caccatcatt	cgctgaaata	1260
gaaaatctag	caattagggc	accaagaacc	gttacttggt	taaattcaac	aagaattcat	1320
acagggacct	tgcaggctg	gagtggttct	aacagatatt	gggcagctca	catgcaaac	1380
ttttcagaaa	ccaattcagg	aaatataaga	tttgacggtc	ctctctatgg	gtcgacggta	1440
ggtactatta	ttcgtactga	taattacgaa	atggggaacc	gagatattta	caccattact	1500
tcagaagctg	ttggcgccct	ttggccacat	ggtcaaacctg	tgttgggagt	cgcttcggct	1560
agatttactt	taagacatct	ttccaataat	tttacacagg	tgctggtgta	tgagaatcca	1620
ataagtaata	gttttaatag	atcaactgta	actagtgaat	tacctggaga	aaactcagat	1680
aggccaactg	atagcgatta	tagtcataga	ctaactgtga	tcacagcttt	tcgagctgga	1740
aataatggta	cgggtccagt	atttggettg	acatctagaa	ctgttaatcg	cgacaatata	1800
attgagcaaa	acaaaattac	acaattocca	ggtgttaagt	cacacactct	caacaattgt	1860
caagtagtta	gagggactgg	atttactgga	ggagactggg	tgagaccaaa	taataatggt	1920
acatttagac	taactattac	ttcattctcg	agccaatctt	accgaatccg	cttacgttat	1980
gtactttcag	tagggaatac	ttcttttagt	atatcttctt	ctgatgcagg	tattttcttc	2040
acaacaattc	cgcttacttc	aacaataaca	tcaactgccc	aaactgtacc	ataccaggct	2100
tttaggggtg	tagatttacc	tattactttt	acaacaccta	ctacccaaag	aaattatacg	2160
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tcgggtgcga	acaatatcga	ggtaacaaat	gatataattc	ccacttatgc	ataccaaaaa	2640
attgatgaat	ccaaattaaa	accctatacg	cgttataaag	ttcgagggtt	tgttggaat	2700
agtaaagatt	tagagttggt	gattacacga	tataatgaag	aagtagatgc	gatttttaaat	2760
gtagcaaatg	atataccaca	tgtcccgaca	cctttctcgc	gtggatttga	tcgatgcaag	2820
ccacattctt	atcctcctat	gaatccagaa	tgtcaccatg	atgtaataaa	taacattgaa	2880
atatcctctc	cttgccatca	caataagatg	gttgataacg	ctgatacatc	ttctcgccat	2940
agtgaattag	gtaaaaaaca	tggcatttgt	catgaatctc	atcattttga	attccatatt	3000
gatacaggaa	aaatcgattt	ggtcgaaaat	ttgggaattt	gggttatatt	taaaatatgt	3060
tccacagatg	gttacgcaac	attagataat	ttggaagtta	ttgaagaggg	tccttttagga	3120
gccgaatcat	tagaacgtgt	gaaaagaaga	gaaaagaat	ggaaacatca	catggaacac	3180
aagtgttcag	aaactaaaca	tgtataccat	gctgcgaaac	aagcggtggt	ggcgttattc	3240
acaaacactc	aatatgatag	attgaagttc	gaaacaacca	tatccaatat	tctatttgct	3300
gattatctcg	tgtcgtcaat	tccgtttgta	tataataaat	ggttaccaga	tgttccaggt	3360
atgaattatg	atatctatac	agaattaaaa	aatctgatta	cgggagcttt	caatctatac	3420

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gatcaacgaa atattataaa aaatggagac tttaataacg gactcatgca ttggcatgcg 3480
acacctcatg cgagagtaga gcaaatagat aataggtctg tgctggtgct tccaaattat 3540
gtgccaatg ttccacaaga ggtttgttta gaacacaatc gtggttatgt attacgtgta 3600
acggcgaaaa aagaaggtcc tggaattgga tatgttacgt tcagtgattg tgcaataat 3660
atagaaaaac tgacatttac ttcttgcgat tatggtacaa acgaagtgcc atatgagcaa 3720
tctaattatc ctacagacgg agtttcatac ggacaacatg gttgtaatat agacagagta 3780
ccgtacgaac aatctgggta tctacagac ggagtatcgt acgaacaatc tggttatcgt 3840
acagacggag tactgtacga acaatctggt tatcgtacag acggagtacc atgcgaacaa 3900
catggttgtc atacagacgg agtaccatac aaacaacatg gttgtcatac agacagatca 3960
agagatgaac aacttgggta tgtgacaaaa acgattgatg tattccctga tacagataaa 4020
gtacgtatcg acattggaga aaccgaaggt acctttaaag tagaaagtgt ggaactgatt 4080
tgtatggaag agtaaatcat aacaaagtaa aaggtatggt ttaatcaaaa atttattttc 4140
cgaacaacag 4150

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<210> SEQ ID NO 6
<211> LENGTH: 1059
<212> TYPE: DNA
<213> ORGANISM: Brevibacillus
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (1)...(1059)

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<400> SEQUENCE: 6

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atg atg gaa cca atg aag ttt cgg aaa aat ggt tta tat aat att gta 48
Met Met Glu Pro Met Lys Phe Arg Lys Asn Gly Leu Tyr Asn Ile Val
1 5 10 15

aat gta aac agt gga aat cta gca gtt gta aaa gat gca tca aaa gaa 96
Asn Val Asn Ser Gly Asn Leu Ala Val Val Lys Asp Ala Ser Lys Glu
20 25 30

aat tat gca cct att att caa ttt gat aaa cgg ggc aca gat aat gaa 144
Asn Tyr Ala Pro Ile Ile Gln Phe Asp Lys Arg Gly Thr Asp Asn Glu
35 40 45

aaa ttt gtg ttc ttt cct ttg gat agt aaa ggt aaa agt caa aca tat 192
Lys Phe Val Phe Phe Pro Leu Asp Ser Lys Gly Lys Ser Gln Thr Tyr
50 55 60

gca att gcc gct tac cat agt gga aag att ata tgt gta aaa gat gca 240
Ala Ile Ala Ala Tyr His Ser Gly Lys Ile Ile Cys Val Lys Asp Ala
65 70 75 80

tca aca gaa aat tat gca cct att atc caa ttt aat tgg aat aac act 288
Ser Thr Glu Asn Tyr Ala Pro Ile Ile Gln Phe Asn Trp Asn Asn Thr
85 90 95

aca aat gaa caa tgg aat att ata cct gat aat tcg tgg ggg tat aat 336
Thr Asn Glu Gln Trp Asn Ile Ile Pro Asp Asn Ser Trp Gly Tyr Asn
100 105 110

atc gtg aat caa aac agt gga aat cta gca gtt gta aaa gat gca tca 384
Ile Val Asn Gln Asn Ser Gly Asn Leu Ala Val Val Lys Asp Ala Ser
115 120 125

aaa gaa aat tat gca cct att att caa ttt gat aaa cgg ggc aca atg 432
Lys Glu Asn Tyr Ala Pro Ile Ile Gln Phe Asp Lys Arg Gly Thr Met
130 135 140

aac gaa gat tgg aaa ttt caa gag gta agc tgg ttt cca gta cct gaa 480
Asn Glu Asp Trp Lys Phe Gln Glu Val Ser Trp Phe Pro Val Pro Glu
145 150 155 160

act cct aca gta gaa act cta cca aaa gcc cct caa ttt aat gat gtt 528

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Thr	Pro	Thr	Val	Glu	Thr	Leu	Pro	Lys	Ala	Pro	Gln	Phe	Asn	Asp	Val	
				165					170					175		
cat	caa	aat	tta	cct	cag	gta	act	gac	gag	ata	ctt	aca	ggt	tac	gca	576
His	Gln	Asn	Leu	Pro	Gln	Val	Thr	Asp	Glu	Ile	Leu	Thr	Gly	Tyr	Ala	
			180					185					190			
atg	att	cct	tgc	att	atg	gtt	aga	gac	cat	aat	tgg	tcc	gat	gaa	tct	624
Met	Ile	Pro	Cys	Ile	Met	Val	Arg	Asp	His	Asn	Trp	Ser	Asp	Glu	Ser	
		195					200					205				
aaa	atg	aaa	act	tct	cct	tac	tat	att	ttg	aaa	aaa	tat	caa	ttt	tgg	672
Lys	Met	Lys	Thr	Ser	Pro	Tyr	Tyr	Ile	Leu	Lys	Lys	Tyr	Gln	Phe	Trp	
	210					215					220					
gag	ttg	ttg	gcg	agt	ttt	cag	ctt	ttt	aat	ggt	gaa	act	caa	aag	agg	720
Glu	Leu	Leu	Ala	Ser	Phe	Gln	Leu	Phe	Asn	Gly	Glu	Thr	Gln	Lys	Arg	
225					230					235					240	
act	tat	aag	gtt	ggt	atg	aat	atg	aca	gat	caa	agg	tca	atg	gaa	aat	768
Thr	Tyr	Lys	Val	Gly	Met	Asn	Met	Thr	Asp	Gln	Arg	Ser	Met	Glu	Asn	
			245						250					255		
tca	att	ggc	aca	atg	att	ggt	gca	gat	gct	ggt	ttt	caa	ttt	gat	ggt	816
Ser	Ile	Gly	Thr	Met	Ile	Gly	Ala	Asp	Ala	Gly	Phe	Gln	Phe	Asp	Gly	
			260					265					270			
cta	act	gat	gcg	ata	aag	tct	gaa	ata	aca	aca	tca	tta	aaa	gtt	gca	864
Leu	Thr	Asp	Ala	Ile	Lys	Ser	Glu	Ile	Thr	Thr	Ser	Leu	Lys	Val	Ala	
		275					280					285				
atc	tct	aga	gaa	aca	aaa	cta	atg	act	gaa	gaa	acc	ggg	gag	gta	atc	912
Ile	Ser	Arg	Glu	Thr	Lys	Leu	Met	Thr	Glu	Glu	Thr	Gly	Glu	Val	Ile	
	290					295					300					
aga	gaa	aat	aaa	act	ggt	aaa	tta	caa	gca	tat	gca	gag	tat	gta	tgt	960
Arg	Glu	Asn	Lys	Thr	Gly	Lys	Leu	Gln	Ala	Tyr	Ala	Glu	Tyr	Val	Cys	
305					310					315					320	
gtt	agt	aaa	ttt	gtg	cta	gaa	cgt	aca	gat	gga	aca	gaa	gta	gct	tct	1008
Val	Ser	Lys	Phe	Val	Leu	Glu	Arg	Thr	Asp	Gly	Thr	Glu	Val	Ala	Ser	
			325						330					335		
tgg	acc	atg	tcg	aat	cct	aat	aca	ata	agc	aaa	act	gtg	ttc	cca	gga	1056
Trp	Thr	Met	Ser	Asn	Pro	Asn	Thr	Ile	Ser	Lys	Thr	Val	Phe	Pro	Gly	
			340				345						350			
taa																1059

&lt;210&gt; SEQ ID NO 7

&lt;211&gt; LENGTH: 352

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Brevibacillus

&lt;400&gt; SEQUENCE: 7

Met	Met	Glu	Pro	Met	Lys	Phe	Arg	Lys	Asn	Gly	Leu	Tyr	Asn	Ile	Val
1				5					10					15	
Asn	Val	Asn	Ser	Gly	Asn	Leu	Ala	Val	Val	Lys	Asp	Ala	Ser	Lys	Glu
		20						25					30		
Asn	Tyr	Ala	Pro	Ile	Ile	Gln	Phe	Asp	Lys	Arg	Gly	Thr	Asp	Asn	Glu
		35				40					45				
Lys	Phe	Val	Phe	Phe	Pro	Leu	Asp	Ser	Lys	Gly	Lys	Ser	Gln	Thr	Tyr
	50					55					60				
Ala	Ile	Ala	Ala	Tyr	His	Ser	Gly	Lys	Ile	Ile	Cys	Val	Lys	Asp	Ala
65					70					75				80	
Ser	Thr	Glu	Asn	Tyr	Ala	Pro	Ile	Ile	Gln	Phe	Asn	Trp	Asn	Asn	Thr
			85					90					95		
Thr	Asn	Glu	Gln	Trp	Asn	Ile	Ile	Pro	Asp	Asn	Ser	Trp	Gly	Tyr	Asn
		100						105					110		
Ile	Val	Asn	Gln	Asn	Ser	Gly	Asn	Leu	Ala	Val	Val	Lys	Asp	Ala	Ser

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115	120	125
Lys Glu Asn Tyr Ala Pro Ile Ile Gln Phe Asp Lys Arg Gly Thr Met		
130	135	140
Asn Glu Asp Trp Lys Phe Gln Glu Val Ser Trp Phe Pro Val Pro Glu		
145	150	155
Thr Pro Thr Val Glu Thr Leu Pro Lys Ala Pro Gln Phe Asn Asp Val		
	165	170
His Gln Asn Leu Pro Gln Val Thr Asp Glu Ile Leu Thr Gly Tyr Ala		
	180	185
Met Ile Pro Cys Ile Met Val Arg Asp His Asn Trp Ser Asp Glu Ser		
	195	200
Lys Met Lys Thr Ser Pro Tyr Tyr Ile Leu Lys Lys Tyr Gln Phe Trp		
	210	215
Glu Leu Leu Ala Ser Phe Gln Leu Phe Asn Gly Glu Thr Gln Lys Arg		
	225	230
Thr Tyr Lys Val Gly Met Asn Met Thr Asp Gln Arg Ser Met Glu Asn		
	245	250
Ser Ile Gly Thr Met Ile Gly Ala Asp Ala Gly Phe Gln Phe Asp Gly		
	260	265
Leu Thr Asp Ala Ile Lys Ser Glu Ile Thr Thr Ser Leu Lys Val Ala		
	275	280
Ile Ser Arg Glu Thr Lys Leu Met Thr Glu Glu Thr Gly Glu Val Ile		
	290	295
Arg Glu Asn Lys Thr Gly Lys Leu Gln Ala Tyr Ala Glu Tyr Val Cys		
	305	310
Val Ser Lys Phe Val Leu Glu Arg Thr Asp Gly Thr Glu Val Ala Ser		
	325	330
Trp Thr Met Ser Asn Pro Asn Thr Ile Ser Lys Thr Val Phe Pro Gly		
	340	345

&lt;210&gt; SEQ ID NO 8

&lt;211&gt; LENGTH: 1688

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Brevibacillus

&lt;400&gt; SEQUENCE: 8

gcgagaatgc tgccacacta attcataaca tggctaaaaa attgggggta taagggatat	60
gaaaacaaca ggaacaaag cggtattatt tgctggggtg cttggatcat taaagctatt	120
aactgaagcc tacgatata atattatttc tgacgaccag atcaacgcaa ttgtgaatgg	180
tctttcggt gtagtggctg taatagctgc ctttactaat aactttaaat caaaaaccga	240
ttagatcaaa ataccgttca gtcgagtga ctctttcttt ttaaataatat acatatttgt	300
gaaaatatgt aataatacgt aatgatatgc aactaagtga ttttccttca gaatcatgtt	360
tctcattatt ttaccacca aaaaaattaa ggggagatta tgatggaacc aatgaagttt	420
cggaaaaatg gtttatataa tattgttaat gtaaacagtg gaaatctagc agttgtaaaa	480
gatgcatcaa aagaaaatta tgcacctatt attcaatttg ataacgggg cacagataat	540
gaaaaatttg tgttctttcc tttggatagt aaaggtaaaa gtcaaacata tgcaattgcc	600
gcttaccata gtggaaagat tatatgtgta aaagatgcat caacagaaaa ttatgcacct	660
attatccaat ttaattggaa taactacta aatgaacaat ggaatattat acctgataat	720
tcgtgggggt ataatatcgt gaatcaaac agtggaatc tagcagttgt aaaagatgca	780
tcaaaagaaa attatgcacc tattattcaa ttgataaac ggggcacaat gaacgaagat	840



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tggaatttc aagaggttaag ctggtttcca gtacctgaaa ctctacagt agaaactcta    900
ccaaaagccc ctcaatttaa tgatgttcat caaaatttac ctcaggtaac tgacgagata    960
cttacagggt acgcaatgat tccttgcatt atggttagag accataattg gtccgatgaa  1020
tctaaaatga aaacttctcc ttactatatt ttgaaaaaat atcaattttg ggagttgttg  1080
gcgagttttc agctttttta tggtgaaact caaaagagga cttataaggt tggtatgaat  1140
atgacagatc aaaggtcaat ggaaaattca attggcacia tgattggtgc agatgctggt  1200
tttcaatttg atggtctaac tgatgcgata aagtctgaaa taacaacatc attaaaagtt  1260
gcaatctcta gagaacaaaa actaatgact gaagaaaccg gggaggtaat cagagaaaaat  1320
aaaactggta aattacaagc atatgcagag tatgtatgtg ttagtaaatt tgtgctagaa  1380
cgtacagatg gaacagaagt agcttcttgg accatgtcga atcctaatac aataagcaaa  1440
actgtgttcc caggataaga aggagaaagc tctctgacgg aaattaaacc gttggagggc  1500
atTTTTtatt gataaacatg taattttacc cattgcataa tctcctcata ttcaaataaa  1560
ataagaacac acattcgtat ttagagagat gttttgatgg ctagtaaaat tcttgatcca  1620
cttghtaaca aattcatctt gccagaacat gcagaaatgt tacgtcagta tcacgaggat  1680
aagaaact                                     1688

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<210> SEQ ID NO 9
<211> LENGTH: 1893
<212> TYPE: DNA
<213> ORGANISM: Brevibacillus
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (1)...(1893)

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<400> SEQUENCE: 9

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atg cta cac aga aat aaa atg ctg aaa gtc ctg agt aca act acg atg    48
Met Leu His Arg Asn Lys Met Leu Lys Val Leu Ser Thr Thr Thr Met
  1             5             10            15

ctg ttg gct tta aca gcc act tct cca gcg ttt tcc tat att act cat    96
Leu Leu Ala Leu Thr Ala Thr Ser Pro Ala Phe Ser Tyr Ile Thr His
  20            25            30

gcc gca aat gga ata cat gat gta gaa gat aaa aag aaa gag gat aaa  144
Ala Ala Asn Gly Ile His Asp Val Glu Asp Lys Lys Lys Glu Asp Lys
  35            40            45

gaa aaa aaa gag aaa gaa gat aaa gaa aag aaa gag cga gag aaa aaa  192
Glu Lys Lys Glu Lys Glu Asp Lys Glu Lys Lys Glu Arg Glu Lys Lys
  50            55            60

gcc aga gaa gaa aga atg aaa gaa att agt aaa gga att gta aca aca  240
Ala Arg Glu Glu Arg Met Lys Glu Ile Ser Lys Gly Ile Val Thr Thr
  65            70            75            80

gag ttt aac agt gaa gaa gaa caa cga tta caa gat acc caa gcc cta  288
Glu Phe Asn Ser Glu Glu Gln Arg Leu Gln Asp Thr Gln Ala Leu
  85            90            95

tta aaa aaa ctt tcg cct gaa gta ttg gaa atg tat gaa aag gtg gga  336
Leu Lys Lys Leu Ser Pro Glu Val Leu Glu Met Tyr Glu Lys Val Gly
 100            105            110

gga aaa att cat ctg aca gat aaa agt att gca gaa aat cct act gtc  384
Gly Lys Ile His Leu Thr Asp Lys Ser Ile Ala Glu Asn Pro Thr Val
 115            120            125

cgg gat atc agt gaa aaa gaa aag cag ata aaa gat agc gaa gga aat  432
Arg Asp Ile Ser Glu Lys Glu Lys Gln Ile Lys Asp Ser Glu Gly Asn
 130            135            140

gaa gtt tcc tta gat tct cat ttt gta ttt tca ata ggt ggt aaa aac  480

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Glu Val Ser Leu Asp Ser His Phe Val Phe Ser Ile Gly Gly Lys Asn	
145 150 155 160	
cca gct ctg att atc cat aca gaa gag tat tcg gaa agc cac agc aaa	528
Pro Ala Leu Ile Ile His Thr Glu Glu Tyr Ser Glu Ser His Ser Lys	
165 170 175	
agc aaa gag gta tat tat gag gta gga aaa gca atc gct cgt gac acg	576
Ser Lys Glu Val Tyr Tyr Glu Val Gly Lys Ala Ile Ala Arg Asp Thr	
180 185 190	
tta gat gaa agt act ttt gca aat gaa gcg ttt tta gat gcg cta cat	624
Leu Asp Glu Ser Thr Phe Ala Asn Glu Ala Phe Leu Asp Ala Leu His	
195 200 205	
caa gca aaa gca gac gaa gat gca agc gcc tta ctt ctt tca cat cta	672
Gln Ala Lys Ala Asp Glu Asp Ala Ser Ala Leu Leu Leu Ser His Leu	
210 215 220	
cct cct cat gaa ggt gag tat gat gcc gca tat gtg aaa gaa cac atc	720
Pro Pro His Glu Gly Glu Tyr Asp Ala Ala Tyr Val Lys Glu His Ile	
225 230 235 240	
aat gag ttt cga gag gtg ttt gca cag gcc ttt gcg tat tat tat gaa	768
Asn Glu Phe Arg Glu Val Phe Ala Gln Ala Phe Ala Tyr Tyr Tyr Glu	
245 250 255	
cct agt tat aaa cct gtg tta aaa gct tat tca ccg gaa atg ttt agg	816
Pro Ser Tyr Lys Pro Val Leu Lys Ala Tyr Ser Pro Glu Met Phe Arg	
260 265 270	
tac atg gat gac atg agc aaa aaa gga ttt gag gaa ata aat aag agt	864
Tyr Met Asp Asp Met Ser Lys Lys Gly Phe Glu Glu Ile Asn Lys Ser	
275 280 285	
tca aat gaa aca caa aaa aca gaa cga aaa gat ttc aaa gaa gat gta	912
Ser Asn Glu Thr Gln Lys Thr Glu Arg Lys Asp Phe Lys Glu Asp Val	
290 295 300	
aca gca gct gac aag tgg tat agg gaa atg ttt aag caa tat agt caa	960
Thr Ala Ala Asp Lys Trp Tyr Arg Glu Met Phe Lys Gln Tyr Ser Gln	
305 310 315 320	
aag ctc aaa cct gaa caa aag tca gcc atc caa tta tat acc acg caa	1008
Lys Leu Lys Pro Glu Gln Lys Ser Ala Ile Gln Leu Tyr Thr Thr Gln	
325 330 335	
aat tat aaa acg atc aat aaa gga tta cga gag gac aat ttg cct gta	1056
Asn Tyr Lys Thr Ile Asn Lys Gly Leu Arg Glu Asp Asn Leu Pro Val	
340 345 350	
gac aag ata aaa gaa gtg cga gac atg tcg aag gct tta gcc aag tcc	1104
Asp Lys Ile Lys Glu Val Arg Asp Met Ser Lys Ala Leu Ala Lys Ser	
355 360 365	
cct att tca gaa gca gga gtt gtg tat aga aaa gtt ggg aaa gat gcg	1152
Pro Ile Ser Glu Ala Gly Val Val Tyr Arg Lys Val Gly Lys Asp Ala	
370 375 380	
cta ggt att gac atc acg act aac ttt aaa aat caa aat gtt gta acg	1200
Leu Gly Ile Asp Ile Thr Thr Asn Phe Lys Asn Gln Asn Val Val Thr	
385 390 395 400	
aaa ttg aaa aat gac tta gaa ggt tca atc aga gaa gag aaa gct ttt	1248
Lys Leu Lys Asn Asp Leu Glu Gly Ser Ile Arg Glu Glu Lys Ala Phe	
405 410 415	
ctt agt acc tca gta gcg aac cac ttt agt gaa tcc ttc gat gca aaa	1296
Leu Ser Thr Ser Val Ala Asn His Phe Ser Glu Ser Phe Asp Ala Lys	
420 425 430	
aca gtt cta ttt aaa ata aat atc cca gaa gga aca cat gct gct tat	1344
Thr Val Leu Phe Lys Ile Asn Ile Pro Glu Gly Thr His Ala Ala Tyr	
435 440 445	
att ttt gga gac ctt gct acc tac caa gga gaa tcc gaa cta atc ata	1392
Ile Phe Gly Asp Leu Ala Thr Tyr Gln Gly Glu Ser Glu Leu Ile Ile	
450 455 460	

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gat aaa ggc tct tct tac agg att gat aaa att aat acg tat gaa tac	1440
Asp Lys Gly Ser Ser Tyr Arg Ile Asp Lys Ile Asn Thr Tyr Glu Tyr	
465 470 475 480	
acg aaa aaa tct gga gtt aaa caa aca aat tta cta gta gaa gca aca	1488
Thr Lys Lys Ser Gly Val Lys Gln Thr Asn Leu Leu Val Glu Ala Thr	
485 490 495	
tta ctt cca agt gat ctt gca gac aat atc aat acg gca gca aaa gag	1536
Leu Leu Pro Ser Asp Leu Ala Asp Asn Ile Asn Thr Ala Ala Lys Glu	
500 505 510	
ctg gaa aag cat gga ttg aag gat cag caa gat aat ata ttg gaa aaa	1584
Leu Glu Lys His Gly Leu Lys Asp Gln Gln Asp Asn Ile Leu Glu Lys	
515 520 525	
ttt att gat tta gat gag tct tta tct gat cta gac cga cta tta aaa	1632
Phe Ile Asp Leu Asp Glu Ser Leu Ser Asp Leu Asp Arg Leu Leu Lys	
530 535 540	
aaa tcg aat gaa atg aat gaa gaa caa acg cta gaa tat ttt aaa gca	1680
Lys Ser Asn Glu Met Asn Glu Glu Gln Thr Leu Glu Tyr Phe Lys Ala	
545 550 555 560	
att gtt gat aat gtc agt cat gta aat gaa cat gat gct act att cta	1728
Ile Val Asp Asn Val Ser His Val Asn Glu His Asp Ala Thr Ile Leu	
565 570 575	
aac aca tta tta acg aat agc aaa gaa aat aca gaa ttt act act tgg	1776
Asn Thr Leu Leu Thr Asn Ser Lys Glu Asn Thr Glu Phe Thr Thr Trp	
580 585 590	
tta gaa gat gta aaa aca atg tac ggg cat att gaa acg ata caa aaa	1824
Leu Glu Asp Val Lys Thr Met Tyr Gly His Ile Glu Thr Ile Gln Lys	
595 600 605	
tta agc gac aat gaa ata att gat tac cta aca aca tta aaa ggt aaa	1872
Leu Ser Asp Asn Glu Ile Ile Asp Tyr Leu Thr Thr Leu Lys Gly Lys	
610 615 620	
tta gac tct gat aac agc taa	1893
Leu Asp Ser Asp Asn Ser	
625 630	

&lt;210&gt; SEQ ID NO 10

&lt;211&gt; LENGTH: 630

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Brevibacillus

&lt;400&gt; SEQUENCE: 10

Met Leu His Arg Asn Lys Met Leu Lys Val Leu Ser Thr Thr Thr Met	
1 5 10 15	
Leu Leu Ala Leu Thr Ala Thr Ser Pro Ala Phe Ser Tyr Ile Thr His	
20 25 30	
Ala Ala Asn Gly Ile His Asp Val Glu Asp Lys Lys Lys Glu Asp Lys	
35 40 45	
Glu Lys Lys Glu Lys Glu Asp Lys Glu Lys Lys Glu Arg Glu Lys Lys	
50 55 60	
Ala Arg Glu Glu Arg Met Lys Glu Ile Ser Lys Gly Ile Val Thr Thr	
65 70 75 80	
Glu Phe Asn Ser Glu Glu Glu Gln Arg Leu Gln Asp Thr Gln Ala Leu	
85 90 95	
Leu Lys Lys Leu Ser Pro Glu Val Leu Glu Met Tyr Glu Lys Val Gly	
100 105 110	
Gly Lys Ile His Leu Thr Asp Lys Ser Ile Ala Glu Asn Pro Thr Val	
115 120 125	
Arg Asp Ile Ser Glu Lys Glu Lys Gln Ile Lys Asp Ser Glu Gly Asn	
130 135 140	

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Glu Val Ser Leu Asp	Ser His Phe Val Phe Ser Ile Gly Gly Lys Asn
145	150 155 160
Pro Ala Leu Ile Ile His Thr Glu Glu Tyr Ser Glu Ser His Ser Lys	
	165 170 175
Ser Lys Glu Val Tyr Tyr Glu Val Gly Lys Ala Ile Ala Arg Asp Thr	
	180 185 190
Leu Asp Glu Ser Thr Phe Ala Asn Glu Ala Phe Leu Asp Ala Leu His	
	195 200 205
Gln Ala Lys Ala Asp Glu Asp Ala Ser Ala Leu Leu Leu Ser His Leu	
	210 215 220
Pro Pro His Glu Gly Glu Tyr Asp Ala Ala Tyr Val Lys Glu His Ile	
	225 230 235 240
Asn Glu Phe Arg Glu Val Phe Ala Gln Ala Phe Ala Tyr Tyr Tyr Glu	
	245 250 255
Pro Ser Tyr Lys Pro Val Leu Lys Ala Tyr Ser Pro Glu Met Phe Arg	
	260 265 270
Tyr Met Asp Asp Met Ser Lys Lys Gly Phe Glu Glu Ile Asn Lys Ser	
	275 280 285
Ser Asn Glu Thr Gln Lys Thr Glu Arg Lys Asp Phe Lys Glu Asp Val	
	290 295 300
Thr Ala Ala Asp Lys Trp Tyr Arg Glu Met Phe Lys Gln Tyr Ser Gln	
	305 310 315 320
Lys Leu Lys Pro Glu Gln Lys Ser Ala Ile Gln Leu Tyr Thr Thr Gln	
	325 330 335
Asn Tyr Lys Thr Ile Asn Lys Gly Leu Arg Glu Asp Asn Leu Pro Val	
	340 345 350
Asp Lys Ile Lys Glu Val Arg Asp Met Ser Lys Ala Leu Ala Lys Ser	
	355 360 365
Pro Ile Ser Glu Ala Gly Val Val Tyr Arg Lys Val Gly Lys Asp Ala	
	370 375 380
Leu Gly Ile Asp Ile Thr Thr Asn Phe Lys Asn Gln Asn Val Val Thr	
	385 390 395 400
Lys Leu Lys Asn Asp Leu Glu Gly Ser Ile Arg Glu Glu Lys Ala Phe	
	405 410 415
Leu Ser Thr Ser Val Ala Asn His Phe Ser Glu Ser Phe Asp Ala Lys	
	420 425 430
Thr Val Leu Phe Lys Ile Asn Ile Pro Glu Gly Thr His Ala Ala Tyr	
	435 440 445
Ile Phe Gly Asp Leu Ala Thr Tyr Gln Gly Glu Ser Glu Leu Ile Ile	
	450 455 460
Asp Lys Gly Ser Ser Tyr Arg Ile Asp Lys Ile Asn Thr Tyr Glu Tyr	
	465 470 475 480
Thr Lys Lys Ser Gly Val Lys Gln Thr Asn Leu Leu Val Glu Ala Thr	
	485 490 495
Leu Leu Pro Ser Asp Leu Ala Asp Asn Ile Asn Thr Ala Ala Lys Glu	
	500 505 510
Leu Glu Lys His Gly Leu Lys Asp Gln Gln Asp Asn Ile Leu Glu Lys	
	515 520 525
Phe Ile Asp Leu Asp Glu Ser Leu Ser Asp Leu Asp Arg Leu Leu Lys	
	530 535 540
Lys Ser Asn Glu Met Asn Glu Glu Gln Thr Leu Glu Tyr Phe Lys Ala	
	545 550 555 560
Ile Val Asp Asn Val Ser His Val Asn Glu His Asp Ala Thr Ile Leu	

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565	570	575
Asn Thr Leu Leu Thr Asn Ser Lys Glu Asn Thr Glu Phe Thr Thr Trp		
580	585	590
Leu Glu Asp Val Lys Thr Met Tyr Gly His Ile Glu Thr Ile Gln Lys		
595	600	605
Leu Ser Asp Asn Glu Ile Ile Asp Tyr Leu Thr Thr Leu Lys Gly Lys		
610	615	620
Leu Asp Ser Asp Asn Ser		
625	630	
<210> SEQ ID NO 11		
<211> LENGTH: 2287		
<212> TYPE: DNA		
<213> ORGANISM: Brevibacillus		
<400> SEQUENCE: 11		
taaacatgaa ggtaatgact gaacctcccg ctatcccaat agatgtcaaa aacagccctc	60	
ttttccatag taatcacata atagatgtac taaaaaagag ggctgttttc ttcatttgtc	120	
gaggacactg ataacagagt gattttatct tcgatggata tacaaaagac tctactacct	180	
tcgggggttc cactacatgt aaggaggaat ttagatgcta cacagaaata aaatgctgaa	240	
agtcctgagt acaactacga tgctgttggc tttaacagcc acttctccag cgttttctca	300	
tattactcat gccgcaaatg gaatacatga tgtagaagat aaaaagaaa aggataaaga	360	
aaaaaaagag aaagaagata aagaaaagaa agagcgagag aaaaagcca gagaagaaag	420	
aatgaaagaa attagtaaag gaattgtaac aacagagttt aacagtgaag aagaacaacg	480	
attacaagat acccaagccc tattaaaaaa actttcgctt gaagtattgg aaatgtatga	540	
aaagtgggga ggaaaaattc atctgacaga taaaagtatt gcagaaaatc ctactgtccg	600	
ggatatcagt gaaaaagaaa agcagataaa agatagcgaa ggaaatgaag ttcccttaga	660	
ttctcatttt gtattttcaa taggtggtaa aaaccagct ctgattatcc atacagaaga	720	
gtattcggaa agccacagca aaagcaaaga ggtatattat gaggtaggaa aagcaatcgc	780	
tcgtgacacg ttagatgaaa gtacttttgc aaatgaagcg tttttagatg cgctacatca	840	
agcaaaagca gacgaagatg caagcgccct acttctttca catctacctc ctcatgaagg	900	
tgagtatgat gccgcatatg tgaaagaaca catcaatgag ttctcgagagg tgtttgcaca	960	
ggcctttgcg tattattatg aacctagtta taaacctgtg ttaaaagctt attcaccgga	1020	
aatgtttagg tacatggatg acatgagcaa aaaaggattt gaggaaataa ataagagttc	1080	
aaatgaaaca caaaaaacag aacgaaaaga tttcaaagaa gatgtaacag cagctgacaa	1140	
gtggtatagg gaaatgttta agcaatatag tcaaaagctc aaacctgaac aaaagtcagc	1200	
catccaatta tataccacgc aaaattataa aacgatcaat aaaggattac gagaggacaa	1260	
tttgcttgta gacaagataa aagaagtgcg agacatgtcg aaggctttag ccaagtcgcc	1320	
tatttcagaa gcaggagtgt tgtatagaaa agttgggaaa gatgcgctag gtattgacat	1380	
cacgactaac tttaaaaatc aaaatgttgt aacgaaattg aaaaatgact tagaagggtc	1440	
aatcagagaa gagaagcgtt ttcttagtac ctcatgtagc aaccacttta gtgaatcctt	1500	
cgatgcaaaa acagtcttat ttaaaataaa tatcccagaa ggaacacatg ctgcttatat	1560	
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ttacaggatt gataaaatta atacgtatga atacacgaaa aaatctggag ttaaacaaac	1680	
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agcaaaagag ctggaaaagc atggattgaa ggatcagcaa gataatatat tgaaaaaatt 1800
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gaatgaagaa caaacgctag aatattttta agcaattggt gataatgtca gtcattgttaa 1920
tgaacatgat gctactattc taaacacatt attaacgaat agcaagaaa atacagaatt 1980
tactacttgg ttagaagatg taaaaacaat gtacgggcat attgaaacga tacaaaaatt 2040
aagcgacaat gaaataattg attacctaac aacattaaaa ggtaaattag actctgataa 2100
cagctaaaag aatctaagat gcttttccat actctaagtc attagcgagt atcctccac 2160
atccagatcg ctgtaaaaat gcaccctaa cgttcacggt taaactcaaa aggtttttcc 2220
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<211> LENGTH: 2031
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
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<221> NAME/KEY: CDS
<222> LOCATION: (1)...(2031)

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<400> SEQUENCE: 12

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1 5 10 15

cag atc atc gag cca agc tcc gac agc ttc ctc tac agc cac aac aac 96
Gln Ile Ile Glu Pro Ser Ser Asp Ser Phe Leu Tyr Ser His Asn Asn
20 25 30

tac ccc tac agc acc gac ccc aac acc gtg ctg cat gga agg aac tac 144
Tyr Pro Tyr Ser Thr Asp Pro Asn Thr Val Leu His Gly Arg Asn Tyr
35 40 45

aag gag tgg ctg aac atg tgc acc ggc act gat gat tca aga gga cca 192
Lys Glu Trp Leu Asn Met Cys Thr Gly Thr Asp Asp Ser Arg Gly Pro
50 55 60

gaa gct gct tca aca gca aga agc gcc atc tcc gtc gcc atc acc atc 240
Glu Ala Ala Ser Thr Ala Arg Ser Ala Ile Ser Val Ala Ile Thr Ile
65 70 75 80

tcc acc acc atc ctc ggc ctc ctc ggc gtg cca ttt gca agc cag atc 288
Ser Thr Thr Ile Leu Gly Leu Leu Gly Val Pro Phe Ala Ser Gln Ile
85 90 95

ggc gcc ttc tac aac ttc gtg ctc aac acc gtc tgg cct caa ggc aac 336
Gly Ala Phe Tyr Asn Phe Val Leu Asn Thr Val Trp Pro Gln Gly Asn
100 105 110

aac caa tgg gag gag ttc atg agg cat gtg gag aac ctc atc aat gaa 384
Asn Gln Trp Glu Glu Phe Met Arg His Val Glu Asn Leu Ile Asn Glu
115 120 125

agg atc gcc gac tat gca aga tca aag gcg ctg gcg gag ctc acc ggc 432
Arg Ile Ala Asp Tyr Ala Arg Ser Lys Ala Leu Ala Glu Leu Thr Gly
130 135 140

ctc ggc aac aac ctc aac ctc tac aga gaa gca ttt gaa gat tgg aga 480
Leu Gly Asn Asn Leu Asn Leu Tyr Arg Glu Ala Phe Glu Asp Trp Arg
145 150 155 160

aga aat cca aca agc caa gaa gcc aag aca agg gtg atc gac cgc ttc 528
Arg Asn Pro Thr Ser Gln Glu Ala Lys Thr Arg Val Ile Asp Arg Phe
165 170 175

aga atc ttg gat ggc ctc ttc gag gcc tac atg cca tca ttt gct gtt 576
Arg Ile Leu Asp Gly Leu Phe Glu Ala Tyr Met Pro Ser Phe Ala Val

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caa gga ttt gag gtg cag ctg ctc acc gtc tac gcc tcc gcc gcc aac Gln Gly Phe Glu Val Gln Leu Leu Thr Val Tyr Ala Ser Ala Ala Asn 195 200 205			624
atc cac ctc ttc ctg ctg aga gat tca agc atc tat ggg ctg gac tgg Ile His Leu Phe Leu Leu Arg Asp Ser Ser Ile Tyr Gly Leu Asp Trp 210 215 220			672
ggc ctc agc caa aca aat gtc aat gag aac tac aac agg cag atc cgc Gly Leu Ser Gln Thr Asn Val Asn Glu Asn Tyr Asn Arg Gln Ile Arg 225 230 235 240			720
cac acc gcc acc tac gcc aac cac tgc acc acc tgg tac caa act ggg His Thr Ala Thr Tyr Ala Asn His Cys Thr Thr Trp Tyr Gln Thr Gly 245 250 255			768
ctg caa agg ctg caa gga aca aat gca aca agc tgg ggc gcc tac aac Leu Gln Arg Leu Gln Gly Thr Asn Ala Thr Ser Trp Gly Ala Tyr Asn 260 265 270			816
agg ttc aga agg gag atg aca ttg acg gtg ctg gac atc tca agc ctc Arg Phe Arg Arg Glu Met Thr Leu Thr Val Leu Asp Ile Ser Ser Leu 275 280 285			864
ttc agc aac tat gac tac aga agc tac cca aca gaa gtt cga gga gag Phe Ser Asn Tyr Asp Tyr Arg Ser Tyr Pro Thr Glu Val Arg Gly Glu 290 295 300			912
ctg aca agg gag atc tac aca gat cct gtt gga ttt gga tgg cag aac Leu Thr Arg Glu Ile Tyr Thr Asp Pro Val Gly Phe Gly Trp Gln Asn 305 310 315 320			960
aat gct cct tcc ttc gcc gag att gag aac ctc gcc atc agg gcg cca Asn Ala Pro Ser Phe Ala Glu Ile Glu Asn Leu Ala Ile Arg Ala Pro 325 330 335			1008
agg acg gtg aca tgg ctc aac tca aca aga atc cac acc ggc acc ttg Arg Thr Val Thr Trp Leu Asn Ser Thr Arg Ile His Thr Gly Thr Leu 340 345 350			1056
caa gga tgg agc ggc agc aac aga tat tgg gcg gcg cac atg caa aac Gln Gly Trp Ser Gly Ser Asn Arg Tyr Trp Ala Ala His Met Gln Asn 355 360 365			1104
ttc tca gaa acc aac agc ggc aac ata aga ttt gat ggg ccg ctc tat Phe Ser Glu Thr Asn Ser Gly Asn Ile Arg Phe Asp Gly Pro Leu Tyr 370 375 380			1152
gga agc acc gtc ggc acc atc atc agg acg gac aac tac gag atg ggc Gly Ser Thr Val Gly Thr Ile Ile Arg Thr Asp Asn Tyr Glu Met Gly 385 390 395 400			1200
aac agg gac atc tac acc atc acc tcc gag gcc gtc ggc gcg ctg tgg Asn Arg Asp Ile Tyr Thr Thr Ile Thr Ser Glu Ala Val Gly Ala Leu Trp 405 410 415			1248
cct cat ggc cag acg gtg cta gga gtt gct tca gca agg ttc acc ctc Pro His Gly Gln Thr Val Leu Gly Val Ala Ser Ala Arg Phe Thr Leu 420 425 430			1296
cgc cac ctc agc aac aac ttc acc caa gtg ctg gtg tat gag aac ccc Arg His Leu Ser Asn Asn Phe Thr Gln Val Leu Val Tyr Glu Asn Pro 435 440 445			1344
atc agc aac agc ttc aac aga agc acc gtc acc tcc gag ctg cca gga Ile Ser Asn Ser Phe Asn Arg Ser Thr Val Thr Ser Glu Leu Pro Gly 450 455 460			1392
gaa aat tca gat cgg cca aca gat tct gac tac agc cac cgg ctg acc Glu Asn Ser Asp Arg Pro Thr Asp Ser Asp Tyr Ser His Arg Leu Thr 465 470 475 480			1440
tgc atc acc gcc ttc cgc gcc ggc aac aat ggc acc gtg ccg gtg ttt Cys Ile Thr Ala Phe Arg Ala Gly Asn Asn Gly Thr Val Pro Val Phe 485 490 495			1488
gga tgg aca tca agg acg gtg aac agg gac aac atc atc gag cag aac			1536

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Gly	Trp	Thr	Ser	Arg	Thr	Val	Asn	Arg	Asp	Asn	Ile	Ile	Glu	Gln	Asn	
			500					505					510			
aag	atc	act	caa	ttt	cct	gga	gtg	aag	agc	cac	acc	ctc	aac	aac	tgc	1584
Lys	Ile	Thr	Gln	Phe	Pro	Gly	Val	Lys	Ser	His	Thr	Leu	Asn	Asn	Cys	
		515					520					525				
caa	gtg	gtg	cgc	ggc	acc	ggc	ttc	acc	ggc	ggt	gat	tgg	ctg	cgg	ccc	1632
Gln	Val	Val	Arg	Gly	Thr	Gly	Phe	Thr	Gly	Gly	Asp	Trp	Leu	Arg	Pro	
	530						535				540					
aac	aac	aat	ggc	acc	ttc	cgc	ctc	acc	atc	acc	agc	ttc	tca	agc	caa	1680
Asn	Asn	Asn	Gly	Thr	Phe	Arg	Leu	Thr	Ile	Thr	Ser	Phe	Ser	Ser	Gln	
	545				550				555						560	
agc	tac	agg	atc	agg	ctg	cgc	tac	gcc	acc	tcc	gtc	ggc	aac	acc	agc	1728
Ser	Tyr	Arg	Ile	Arg	Leu	Arg	Tyr	Ala	Thr	Ser	Val	Gly	Asn	Thr	Ser	
			565					570						575		
ttg	gtg	atc	tca	agc	tct	gat	gct	ggc	atc	agc	agc	acc	acc	atc	ccg	1776
Leu	Val	Ile	Ser	Ser	Ser	Asp	Ala	Gly	Ile	Ser	Ser	Thr	Thr	Ile	Pro	
		580						585					590			
ctg	aca	agc	acc	atc	acc	tgc	ctg	ccg	cag	acg	gtg	cca	tat	caa	gcc	1824
Leu	Thr	Ser	Thr	Ile	Thr	Ser	Leu	Pro	Gln	Thr	Val	Pro	Tyr	Gln	Ala	
		595					600					605				
ttc	cgc	gtg	gtg	gac	ctg	ccc	atc	acc	ttc	acc	acg	ccg	acg	acg	cag	1872
Phe	Arg	Val	Val	Asp	Leu	Pro	Ile	Thr	Phe	Thr	Thr	Pro	Thr	Thr	Gln	
	610					615					620					
agg	aac	tac	acc	ttc	gac	ttc	cgc	ctc	caa	aat	cca	tca	aat	gca	aat	1920
Arg	Asn	Tyr	Thr	Phe	Asp	Phe	Arg	Leu	Gln	Asn	Pro	Ser	Asn	Ala	Asn	
	625				630				635					640		
gtg	ttc	atc	gac	aga	ttt	gaa	ttt	gtt	cca	ata	gga	gga	agc	ctc	tca	1968
Val	Phe	Ile	Asp	Arg	Phe	Glu	Phe	Val	Pro	Ile	Gly	Gly	Ser	Leu	Ser	
			645					650						655		
gaa	tat	gaa	aca	aag	cac	cag	ctg	gag	aag	gca	agg	aag	gcc	gtc	aac	2016
Glu	Tyr	Glu	Thr	Lys	His	Gln	Leu	Glu	Lys	Ala	Arg	Lys	Ala	Val	Asn	
		660					665						670			
gac	ctc	ttc	acc	aac												2031
Asp	Leu	Phe	Thr	Asn												
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	1			5					10					15		
agc	aca	gat	cca	aac	acc	gtg	ctg	cat	gga	agg	aac	tac	aag	gaa	tgg	96
Ser	Thr	Asp	Pro	Asn	Thr	Val	Leu	His	Gly	Arg	Asn	Tyr	Lys	Glu	Trp	
		20					25						30			
ctc	aac	atg	tgc	act	gga	aca	gat	gat	tca	aga	gga	cca	gaa	gct	gct	144
Leu	Asn	Met	Cys	Thr	Gly	Thr	Asp	Asp	Ser	Arg	Gly	Pro	Glu	Ala	Ala	
		35					40					45				
tca	aca	gca	agg	agc	gcc	atc	tcc	gtc	gcc	atc	acc	ata	agc	acc	acc	192
Ser	Thr	Ala	Arg	Ser	Ala	Ile	Ser	Val	Ala	Ile	Thr	Ile	Ser	Thr	Thr	
	50					55					60					
atc	ctc	ggc	ctg	ctg	gga	gtt	ccc	ttc	gcc	agc	cag	atc	ggc	gcc	ttc	240
Ile	Leu	Gly	Leu	Leu	Gly	Val	Pro	Phe	Ala	Ser	Gln	Ile	Gly	Ala	Phe	
	65					70				75					80	



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tac aac ttc gtc ctc aac acc gtc tgg cct caa gga aac aac caa tgg	288
Tyr Asn Phe Val Leu Asn Thr Val Trp Pro Gln Gly Asn Asn Gln Trp	
85 90 95	
gag gag ttc atg agg cat gtg gag aac ctc atc aat gaa agg att gct	336
Glu Glu Phe Met Arg His Val Glu Asn Leu Ile Asn Glu Arg Ile Ala	
100 105 110	
gat tat gca aga tca aag gcg ctg gcg gag ctc acc ggc ctc ggc aac	384
Asp Tyr Ala Arg Ser Lys Ala Leu Ala Glu Leu Thr Gly Leu Gly Asn	
115 120 125	
aac ctc aac ctc tac aga gaa gca ttt gaa gat tgg aga aga aat cca	432
Asn Leu Asn Leu Tyr Arg Glu Ala Phe Glu Asp Trp Arg Arg Asn Pro	
130 135 140	
aca tca caa gaa gca aaa aca agg gtg atc gac agg ttc aga atc ttg	480
Thr Ser Gln Glu Ala Lys Thr Arg Val Ile Asp Arg Phe Arg Ile Leu	
145 150 155 160	
gat ggc ctc ttc gag gcc tac atg ccc tcc ttc gcc gtc caa gga ttt	528
Asp Gly Leu Phe Glu Ala Tyr Met Pro Ser Phe Ala Val Gln Gly Phe	
165 170 175	
gag gtg cag ctg ctc acc gtc tat gct tct gct gcc aac atc cac ctc	576
Glu Val Gln Leu Leu Thr Val Tyr Ala Ser Ala Ala Asn Ile His Leu	
180 185 190	
ttc ctg ctg aga gat tca agc atc tat ggc ctg gac tgg ggc ttg agc	624
Phe Leu Leu Arg Asp Ser Ser Ile Tyr Gly Leu Asp Trp Gly Leu Ser	
195 200 205	
caa aca aat gtc aat gag aac tac aac agg cag atc cgc cac aca gca	672
Gln Thr Asn Val Asn Glu Asn Tyr Asn Arg Gln Ile Arg His Thr Ala	
210 215 220	
aca tat gcc aac cac tgc acc acc tgg tat caa act ggc ctc cag agg	720
Thr Tyr Ala Asn His Cys Thr Thr Trp Tyr Gln Thr Gly Leu Gln Arg	
225 230 235 240	
ctg caa gga aca aat gca aca agc tgg ggc gcc tac aac agg ttc aga	768
Leu Gln Gly Thr Asn Ala Thr Ser Trp Gly Ala Tyr Asn Arg Phe Arg	
245 250 255	
agg gag atg acc ctc acc gtg ctg gac atc agc agc ctc ttc tca aac	816
Arg Glu Met Thr Leu Thr Val Leu Asp Ile Ser Ser Leu Phe Ser Asn	
260 265 270	
tat gac tac aga agc tac cca aca gaa gtt cga gga gag ctg aca agg	864
Tyr Asp Tyr Arg Ser Tyr Pro Thr Glu Val Arg Gly Glu Leu Thr Arg	
275 280 285	
gag atc tac aca gat cct gtt gga ttt gga tgg cag aac aat gct cct	912
Glu Ile Tyr Thr Asp Pro Val Gly Phe Gly Trp Gln Asn Asn Ala Pro	
290 295 300	
tct ttt gct gaa att gag aac ctc gcc atc aga gct cca agg acg gtg	960
Ser Phe Ala Glu Ile Glu Asn Leu Ala Ile Arg Ala Pro Arg Thr Val	
305 310 315 320	
aca tgg ctg aac tca aca aga atc cac acc ggc acc ctc caa gga tgg	1008
Thr Trp Leu Asn Ser Thr Arg Ile His Thr Gly Thr Leu Gln Gly Trp	
325 330 335	
agc ggc agc aac aga tat tgg gcg gcg cac atg caa aac ttc tca gaa	1056
Ser Gly Ser Asn Arg Tyr Trp Ala Ala His Met Gln Asn Phe Ser Glu	
340 345 350	
aca aac agc ggc aac atc aga ttt gat ggg ccg ctc tat gga agc acc	1104
Thr Asn Ser Gly Asn Ile Arg Phe Asp Gly Pro Leu Tyr Gly Ser Thr	
355 360 365	
gtc ggc acc atc atc agg aca gac aac tat gag atg ggc aac agg gac	1152
Val Gly Thr Ile Ile Arg Thr Asp Asn Tyr Glu Met Gly Asn Arg Asp	
370 375 380	
atc tac acc ata aca tca gaa gct gtt gga gct ctc tgg cct cat ggc	1200
Ile Tyr Thr Ile Thr Ser Glu Ala Val Gly Ala Leu Trp Pro His Gly	
385 390 395 400	

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caa aca gtg ctg ggc gtc gcc tct gca agg ttc acc ctc cgc cac ctc	1248
Gln Thr Val Leu Gly Val Ala Ser Ala Arg Phe Thr Leu Arg His Leu	
405 410 415	
tcc aac aac ttc acc cag gtg ctg gtg tat gag aac ccc atc agc aac	1296
Ser Asn Asn Phe Thr Gln Val Leu Val Tyr Glu Asn Pro Ile Ser Asn	
420 425 430	
agc ttc aac aga agc acc gtg aca tca gag ctg cca gga gaa aac agc	1344
Ser Phe Asn Arg Ser Thr Val Thr Ser Glu Leu Pro Gly Glu Asn Ser	
435 440 445	
gac cgg cca aca gat tct gac tac agc cac cgc ctc acc tgc atc acc	1392
Asp Arg Pro Thr Asp Ser Asp Tyr Ser His Arg Leu Thr Cys Ile Thr	
450 455 460	
gcc ttc aga gct ggc aac aat gga aca gtt cct gtt ttt gga tgg aca	1440
Ala Phe Arg Ala Gly Asn Asn Gly Thr Val Pro Val Phe Gly Trp Thr	
465 470 475 480	
tca agg acg gtg aac agg gac aac atc atc gag cag aac aag atc act	1488
Ser Arg Thr Val Asn Arg Asp Asn Ile Ile Glu Gln Asn Lys Ile Thr	
485 490 495	
cag ttc ccc ggc gtc aag agc cac acc ctc aac aac tgc cag gtg gtt	1536
Gln Phe Pro Gly Val Lys Ser His Thr Leu Asn Asn Cys Gln Val Val	
500 505 510	
aga gga act ggc ttc act gga gga gat tgg ctg cgg cca aac aac aat	1584
Arg Gly Thr Gly Phe Thr Gly Asp Trp Leu Arg Pro Asn Asn Asn	
515 520 525	
ggc acc ttc cgc ctc acc atc acc agc ttc tca agc caa agc tac agg	1632
Gly Thr Phe Arg Leu Thr Ile Thr Ser Phe Ser Ser Gln Ser Tyr Arg	
530 535 540	
atc agg ctg aga tat gca act tct gtt ggt aac acc agc ttg gtg atc	1680
Ile Arg Leu Arg Tyr Ala Thr Ser Val Gly Asn Thr Ser Leu Val Ile	
545 550 555 560	
tcc tcc tca gat gct ggc atc tcc tcc acc acc atc ccc ctc acc tcc	1728
Ser Ser Ser Asp Ala Gly Ile Ser Ser Thr Thr Ile Pro Leu Thr Ser	
565 570 575	
acc atc acc agc ctt cct caa aca gtt cca tat caa gcc ttc cgc gtg	1776
Thr Ile Thr Ser Leu Pro Gln Thr Val Pro Tyr Gln Ala Phe Arg Val	
580 585 590	
gta gat ctt ccc atc acc ttc acc acc ccc acc acc caa agg aac tac	1824
Val Asp Leu Pro Ile Thr Phe Thr Thr Pro Thr Thr Gln Arg Asn Tyr	
595 600 605	
acc ttt gat ttc cgc ctc caa aat cca agc aat gca aat gtt ttc atc	1872
Thr Phe Asp Phe Arg Leu Gln Asn Pro Ser Asn Ala Asn Val Phe Ile	
610 615 620	
gac aga ttt gaa ttt gtt cca att gga gga agc ctc tca gaa tat gaa	1920
Asp Arg Phe Glu Phe Val Pro Ile Gly Gly Ser Leu Ser Glu Tyr Glu	
625 630 635 640	
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Thr Lys His	

&lt;210&gt; SEQ ID NO 14

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&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Artificial Sequence

&lt;220&gt; FEATURE:

&lt;223&gt; OTHER INFORMATION: synthetic nucleotide sequence

&lt;221&gt; NAME/KEY: CDS

&lt;222&gt; LOCATION: (1)...(1056)

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Met Met Glu Pro Met Lys Phe Arg Lys Asn Gly Leu Tyr Asn Ile Val	
1 5 10 15	

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aac tat gct ccc atc atc cag ttc gac aag aga gga aca gac aat gag Asn Tyr Ala Pro Ile Ile Gln Phe Asp Lys Arg Gly Thr Asp Asn Glu	144
35 40 45	
aag ttt gtt ttc ttc ccg ctg gac agc aag ggc aag agc caa aca tat Lys Phe Val Phe Phe Pro Leu Asp Ser Lys Gly Lys Ser Gln Thr Tyr	192
50 55 60	
gcc att gct gcc tac cac agc ggc aag atc atc tgc gtg aag gat gct Ala Ile Ala Ala Tyr His Ser Gly Lys Ile Ile Cys Val Lys Asp Ala	240
65 70 75 80	
tca aca gag aac tat gct ccc atc atc cag ttc aac tgg aac aac acc Ser Thr Glu Asn Tyr Ala Pro Ile Ile Gln Phe Asn Trp Asn Asn Thr	288
85 90 95	
acc aat gag caa tgg aac atc atc cct gac aac agc tgg ggc tac aac Thr Asn Glu Gln Trp Asn Ile Ile Pro Asp Asn Ser Trp Gly Tyr Asn	336
100 105 110	
atc gtc aac cag aac agc ggc aac ctc gcc gtg gtg aag gat gca agc Ile Val Asn Gln Asn Ser Gly Asn Leu Ala Val Val Lys Asp Ala Ser	384
115 120 125	
aag gag aac tat gct ccc atc atc cag ttc gac aag aga gga aca atg Lys Glu Asn Tyr Ala Pro Ile Ile Gln Phe Asp Lys Arg Gly Thr Met	432
130 135 140	
aat gaa gat tgg aag ttc caa gaa gtt tca tgg ttc cct gtt cca gaa Asn Glu Asp Trp Lys Phe Gln Glu Val Ser Trp Phe Pro Val Pro Glu	480
145 150 155 160	
acg ccg acg gtg gaa acc ctc ccc aag gcg ccg cag ttc aat gat gtt Thr Pro Thr Val Glu Thr Leu Pro Lys Ala Pro Gln Phe Asn Asp Val	528
165 170 175	
cat caa aat ctt cct cag gtg aca gat gag atc ctc act gga tat gcc His Gln Asn Leu Pro Gln Val Thr Asp Glu Ile Leu Thr Gly Tyr Ala	576
180 185 190	
atg atc ccc tgc atc atg gtg aga gat cac aac tgg agt gat gaa agc Met Ile Pro Cys Ile Met Val Arg Asp His Asn Trp Ser Asp Glu Ser	624
195 200 205	
aag atg aaa act tct cca tac tac atc ctc aag aag tac cag ttc tgg Lys Met Lys Thr Ser Pro Tyr Tyr Ile Leu Lys Lys Tyr Gln Phe Trp	672
210 215 220	
gag ctg ctg gca tca ttc cag ctc ttc aat gga gaa acc cag aag agg Glu Leu Leu Ala Ser Phe Gln Leu Phe Asn Gly Glu Thr Gln Lys Arg	720
225 230 235 240	
acc tac aag gtg ggg atg aac atg aca gat caa aga agc atg gag aac Thr Tyr Lys Val Gly Met Asn Met Thr Asp Gln Arg Ser Met Glu Asn	768
245 250 255	
agc atc ggc acc atg att gga gct gat gct ggc ttc cag ttt gat ggc Ser Ile Gly Thr Met Ile Gly Ala Asp Ala Gly Phe Gln Phe Asp Gly	816
260 265 270	
ctc acc gac gcc atc aag agc gag atc acc acc agc ttg aag gtg gcc Leu Thr Asp Ala Ile Lys Ser Glu Ile Thr Thr Ser Leu Lys Val Ala	864
275 280 285	
atc tca aga gaa aca aag ctg atg aca gaa gaa act gga gaa gtc atc Ile Ser Arg Glu Thr Lys Leu Met Thr Glu Glu Thr Gly Glu Val Ile	912
290 295 300	
agg gag aac aag aca ggg aag ctg caa gcc tat gct gaa tat gtc tgc Arg Glu Asn Lys Thr Gly Lys Leu Gln Ala Tyr Ala Glu Tyr Val Cys	960
305 310 315 320	
gtc agc aag ttc gtg ctg gag agg aca gat gga aca gag gtg gca agc Val Ser Lys Phe Val Leu Glu Arg Thr Asp Gly Thr Glu Val Ala Ser	1008

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	325	330	335	
tgg acc atg agc aac ccc aac acc atc agc aag acg gtg ttc cct gga				1056
Trp Thr Met Ser Asn Pro Asn Thr Ile Ser Lys Thr Val Phe Pro Gly				
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1 5 10 15				
ctg ctg gcg ctc acc gcc acc tcg ccg gcc ttc tca tac atc acc cat				96
Leu Leu Ala Leu Thr Ala Thr Ser Pro Ala Phe Ser Tyr Ile Thr His				
20 25 30				
gct gca aat ggc att cat gat gtg gag gac aag aag aag gag gac aag				144
Ala Ala Asn Gly Ile His Asp Val Glu Asp Lys Lys Lys Glu Asp Lys				
35 40 45				
gag aag aag gag aag gag gac aag gag aag aag gag agg gag aag aag				192
Glu Lys Lys Glu Lys Glu Asp Lys Glu Lys Lys Glu Arg Glu Lys Lys				
50 55 60				
gca aga gaa gaa agg atg aag gag atc agc aag ggc atc gtc acc acc				240
Ala Arg Glu Glu Arg Met Lys Glu Ile Ser Lys Gly Ile Val Thr Thr				
65 70 75 80				
gag ttc aac tca gaa gaa gag cag agg ctg caa gac act caa gct ctg				288
Glu Phe Asn Ser Glu Glu Glu Gln Arg Leu Gln Asp Thr Gln Ala Leu				
85 90 95				
ctg aag aag ctc tcg ccg gag gtg ctg gag atg tat gag aag gtt gga				336
Leu Lys Lys Leu Ser Pro Glu Val Leu Glu Met Tyr Glu Lys Val Gly				
100 105 110				
gga aag atc cac ctc acc gac aag agc att gct gag aac ccc acc gtc				384
Gly Lys Ile His Leu Thr Asp Lys Ser Ile Ala Glu Asn Pro Thr Val				
115 120 125				
agg gac atc tca gag aag gag aag cag atc aag gat tca gaa gga aat				432
Arg Asp Ile Ser Glu Lys Glu Lys Gln Ile Lys Asp Ser Glu Gly Asn				
130 135 140				
gaa gtt tct ctg gat tct cat ttt gtt ttc tcc atc ggc ggc aag aac				480
Glu Val Ser Leu Asp Ser His Phe Val Phe Ser Ile Gly Gly Lys Asn				
145 150 155 160				
ccg gcg ctg atc atc cac aca gaa gaa tat tca gaa agc cac agc aag				528
Pro Ala Leu Ile Ile His Thr Glu Glu Tyr Ser Glu Ser His Ser Lys				
165 170 175				
agc aag gag gtg tac tac gag gtg ggc aag gcc att gca agg gac acc				576
Ser Lys Glu Val Tyr Tyr Glu Val Gly Lys Ala Ile Ala Arg Asp Thr				
180 185 190				
ttg gat gaa agc acc ttc gcc aat gaa gcc ttc ctg gat gct ctt cat				624
Leu Asp Glu Ser Thr Phe Ala Asn Glu Ala Phe Leu Asp Ala Leu His				
195 200 205				
caa gca aaa gct gat gaa gat gct tca gct ctg ctg ctg agc cat ctt				672
Gln Ala Lys Ala Asp Glu Asp Ala Ser Ala Leu Leu Leu Ser His Leu				
210 215 220				
cct cct cat gaa gga gaa tat gat gct gca tat gtc aag gag cac atc				720
Pro Pro His Glu Gly Glu Tyr Asp Ala Ala Tyr Val Lys Glu His Ile				
225 230 235 240				
aat gag ttc aga gaa gtt ttt gct caa gcc ttc gcc tac tac tac gag				768

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Asn	Glu	Phe	Arg	Glu	Val	Phe	Ala	Gln	Ala	Phe	Ala	Tyr	Tyr	Tyr	Glu	
				245					250					255		
cca	tca	tac	aag	ccg	gtg	ctg	aag	gcc	tac	tcg	ccg	gag	atg	ttc	aga	816
Pro	Ser	Tyr	Lys	Pro	Val	Leu	Lys	Ala	Tyr	Ser	Pro	Glu	Met	Phe	Arg	
			260					265					270			
tac	atg	gat	gac	atg	agc	aag	aag	ggc	ttc	gag	gag	atc	aac	aag	agc	864
Tyr	Met	Asp	Asp	Met	Ser	Lys	Lys	Gly	Phe	Glu	Glu	Ile	Asn	Lys	Ser	
		275						280				285				
agc	aat	gaa	aca	caa	aaa	aca	gaa	agg	aag	gac	ttc	aag	gaa	gat	gtc	912
Ser	Asn	Glu	Thr	Gln	Lys	Thr	Glu	Arg	Lys	Asp	Phe	Lys	Glu	Asp	Val	
	290					295					300					
acc	gcc	gcc	gac	aaa	tgg	tac	agg	gag	atg	ttc	aag	cag	tac	agc	cag	960
Thr	Ala	Ala	Asp	Lys	Trp	Tyr	Arg	Glu	Met	Phe	Lys	Gln	Tyr	Ser	Gln	
	305				310					315					320	
aag	ctg	aag	cca	gag	cag	aaa	tca	gcc	atc	cag	ctc	tac	acc	acc	cag	1008
Lys	Leu	Lys	Pro	Glu	Gln	Lys	Ser	Ala	Ile	Gln	Leu	Tyr	Thr	Thr	Gln	
			325						330					335		
aac	tac	aag	acc	atc	aac	aag	ggc	ctc	agg	gag	gac	aac	ctt	cct	gtt	1056
Asn	Tyr	Lys	Thr	Ile	Asn	Lys	Gly	Leu	Arg	Glu	Asp	Asn	Leu	Pro	Val	
		340					345						350			
gac	aag	atc	aag	gag	gtg	agg	gac	atg	agc	aag	gcg	ctg	gcc	aag	agc	1104
Asp	Lys	Ile	Lys	Glu	Val	Arg	Asp	Met	Ser	Lys	Ala	Leu	Ala	Lys	Ser	
		355				360						365				
ccc	atc	tca	gaa	gct	gga	gtg	gtg	tac	agg	aag	gtg	ggc	aag	gat	gct	1152
Pro	Ile	Ser	Glu	Ala	Gly	Val	Val	Tyr	Arg	Lys	Val	Gly	Lys	Asp	Ala	
	370				375						380					
ctt	ggc	atc	gac	atc	acc	acc	aac	ttc	aag	aat	caa	aat	gtt	gtc	acc	1200
Leu	Gly	Ile	Asp	Ile	Thr	Thr	Asn	Phe	Lys	Asn	Gln	Asn	Val	Val	Thr	
	385				390					395				400		
aag	ctg	aag	aat	gat	ctg	gaa	gga	agc	atc	agg	gag	gag	aag	gcc	ttc	1248
Lys	Leu	Lys	Asn	Asp	Leu	Glu	Gly	Ser	Ile	Arg	Glu	Glu	Lys	Ala	Phe	
			405						410					415		
ctc	tcc	acc	tcc	gtc	gcc	aac	cac	ttc	tca	gag	agc	ttt	gat	gca	aaa	1296
Leu	Ser	Thr	Ser	Val	Ala	Asn	His	Phe	Ser	Glu	Ser	Phe	Asp	Ala	Lys	
			420				425						430			
aca	gtg	ctc	ttc	aag	atc	aac	atc	cca	gaa	gga	aca	cat	gct	gcc	tac	1344
Thr	Val	Leu	Phe	Lys	Ile	Asn	Ile	Pro	Glu	Gly	Thr	His	Ala	Ala	Tyr	
		435					440					445				
atc	ttt	gga	gat	ctg	gcc	acc	tac	caa	gga	gaa	agc	gag	ctg	atc	atc	1392
Ile	Phe	Gly	Asp	Leu	Ala	Thr	Tyr	Gln	Gly	Glu	Ser	Glu	Leu	Ile	Ile	
	450					455					460					
gac	aaa	gga	agc	agc	tac	agg	atc	gac	aag	atc	aac	aca	tat	gag	tac	1440
Asp	Lys	Gly	Ser	Ser	Tyr	Arg	Ile	Asp	Lys	Ile	Asn	Thr	Tyr	Glu	Tyr	
	465				470					475				480		
acc	aag	aag	agc	ggc	gtg	aag	caa	aca	aac	ctg	ctg	gtg	gag	gcc	acc	1488
Thr	Lys	Lys	Ser	Gly	Val	Lys	Gln	Thr	Asn	Leu	Leu	Val	Glu	Ala	Thr	
			485						490					495		
ctg	ctg	cca	tca	gat	ctt	gct	gac	aac	atc	aac	acc	gcc	gcc	aag	gag	1536
Leu	Leu	Pro	Ser	Asp	Leu	Ala	Asp	Asn	Ile	Asn	Thr	Ala	Ala	Lys	Glu	
			500					505					510			
ctg	gag	aag	cat	ggc	ctc	aag	gat	cag	cag	gac	aac	atc	ctg	gag	aag	1584
Leu	Glu	Lys	His	Gly	Leu	Lys	Asp	Gln	Gln	Asp	Asn	Ile	Leu	Glu	Lys	
		515					520					525				
ttc	atc	gac	ctg	gat	gaa	agc	ctc	tca	gat	ctg	gac	agg	ctg	ctg	aag	1632
Phe	Ile	Asp	Leu	Asp	Glu	Ser	Leu	Ser	Asp	Leu	Asp	Arg	Leu	Leu	Lys	
	530					535					540					
aag	agc	aat	gag	atg	aat	gag	gag	cag	acg	ctg	gag	tac	ttc	aag	gcc	1680
Lys	Ser	Asn	Glu	Met	Asn	Glu	Glu	Gln	Thr	Leu	Glu	Tyr	Phe	Lys	Ala	
	545				550					555					560	

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atc gtc gac aat gtt tct cat gtc aat gaa cat gat gcc acc atc ctc	1728
Ile Val Asp Asn Val Ser His Val Asn Glu His Asp Ala Thr Ile Leu	
565 570 575	
aac acc ctc ctc acc aac tca aag gag aac aca gag ttc acc acc tgg	1776
Asn Thr Leu Leu Thr Asn Ser Lys Glu Asn Thr Glu Phe Thr Thr Trp	
580 585 590	
ctg gag gat gtc aag acc atg tat ggc cac att gaa acc atc cag aag	1824
Leu Glu Asp Val Lys Thr Met Tyr Gly His Ile Glu Thr Ile Gln Lys	
595 600 605	
ctc tcc gac aat gag atc atc gac tac ctc acc acc ttg aag ggc aag	1872
Leu Ser Asp Asn Glu Ile Ile Asp Tyr Leu Thr Thr Leu Lys Gly Lys	
610 615 620	
ctg gac agc gac aac agc	1890
Leu Asp Ser Asp Asn Ser	
625 630	

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That which is claimed:

1. An isolated or recombinant nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of:

- a) the nucleotide sequence of SEQ ID NO:1 or 5, or the full-length complement thereof;
- b) a nucleotide sequence that encodes a polypeptide comprising the amino acid sequence of SEQ ID NO:4; and
- c) a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence having at least 95% sequence identity to the amino acid sequence of SEQ ID NO:4, wherein said polypeptide has pesticidal activity against a plant pest;

and wherein said nucleotide sequence is operably linked to a heterologous promoter.

2. The recombinant nucleic acid molecule of claim 1, wherein said nucleotide sequence is a synthetic sequence that has been designed for expression in a plant.

3. The recombinant nucleic acid of claim 2, wherein said nucleic acid sequence is SEQ ID NO:12.

4. A vector comprising the nucleic acid molecule of claim 1.

5. The vector of claim 4, further comprising a nucleic acid molecule encoding a heterologous polypeptide.

6. A host cell that contains the nucleic acid molecule of claim 1.

7. The host cell of claim 6, wherein said host cell is a bacterial host cell.

8. The host cell of claim 6, wherein said host cell is a plant cell.

9. A transgenic plant comprising the host cell of claim 8.

10. The transgenic plant of claim 9, wherein said plant is selected from the group consisting of maize, sorghum, wheat, cabbage, sunflower, tomato, crucifers, peppers, potato, cotton, rice, soybean, sugarbeet, sugarcane, tobacco, barley, and oilseed rape.

11. A transgenic seed comprising the nucleic acid molecule of claim 1.

12. An isolated polypeptide with insecticidal activity, selected from the group consisting of:

- a) a polypeptide comprising the amino acid sequence of SEQ ID NO:4;
- b) a polypeptide that is encoded by the nucleotide sequence of SEQ ID NO:1, 3, or 5;
- c) a polypeptide that is at least 95% sequence identity to the amino acid sequence of SEQ ID NO:4, wherein said polypeptide has pesticidal activity against a plant pest;

and wherein said polypeptide is operably linked to a leader sequence, a signal sequence or a transit peptide.

13. A composition comprising the polypeptide of claim 12.

14. The composition of claim 13, wherein said composition is selected from the group consisting of a powder, dust, pellet, granule, spray, emulsion, colloid, and solution.

15. The composition of claim 13, wherein said composition is prepared by desiccation, lyophilization, homogenization, extraction, filtration, centrifugation, sedimentation, or concentration of a culture of *Bacillus thuringiensis* cells.

16. The composition of claim 14, comprising from about 1% to about 99% by weight of said polypeptide.

17. A method for controlling or killing a plant pest population comprising contacting said population with an insecticidally-effective amount of a polypeptide, wherein said polypeptide is selected from the group consisting of:

- a) a polypeptide comprising the amino acid sequence of SEQ ID NO:4;
- b) a polypeptide that is encoded by the nucleotide sequence of SEQ ID NO:1, 3, or 5;
- c) a polypeptide that is at least 95% sequence identity to the amino acid sequence of SEQ ID NO:4, wherein said polypeptide has pesticidal activity against a plant pest.

18. A method for producing a polypeptide with insecticidal activity, comprising culturing the host cell of claim 6 under conditions in which the nucleic acid molecule encoding the polypeptide is expressed.

19. A plant or a plant cell having stably incorporated into its genome a DNA construct comprising a nucleotide sequence that encodes a protein having insecticidal activity, wherein said nucleotide sequence is selected from the group consisting of:

- a) the nucleotide sequence of SEQ ID NO:1 or 5;
- b) a nucleotide sequence that encodes a polypeptide comprising the amino acid sequence of SEQ ID NO:4; and
- c) a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence having at least 95% sequence identity to the amino acid sequence of SEQ ID NO:4, wherein said polypeptide has pesticidal activity against a plant pest;

wherein said nucleotide sequence is operably linked to a promoter that drives expression of a coding sequence in a plant cell.

20. A method for protecting a plant from an insect pest, comprising introducing into said plant or cell thereof at least one expression vector comprising a nucleotide sequence that

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encodes a insecticidal polypeptide, wherein said nucleotide sequence is selected from the group consisting of:

- a) the nucleotide sequence of SEQ ID NO:1 or 5;
- b) a nucleotide sequence that encodes a polypeptide comprising the amino acid sequence of SEQ ID NO:4; and 5
- c) a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence having at least 95% sequence identity to the amino acid sequence of SEQ ID NO:4, wherein said polypeptide has pesticidal activity against a plant pest; and 10

wherein said nucleotide sequence is operably linked to a promoter that drives expression of a coding sequence in a plant cell.

**21.** The isolated or recombinant nucleic acid of claim **1**, wherein said promoter is capable of driving expression of said nucleotide sequence in a plant cell. 15

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